

# Alewife (*Alosa pseudoharengus*)

## Ecological Risk Screening Summary

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Organism Type: Fish

Overall Risk Assessment Category: High



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## 1 Native Range and Status in the United States

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### Native Range

From Froese and Pauly (2025a):

“North America: Atlantic coast from Red Bay, Labrador in Canada to South Carolina in USA; many landlocked populations exist. [...] Native to Lake Ontario; [...]”

### Status in the United States

From Froese and Pauly (2025a):

“North America: Atlantic coast from [...] Canada to South Carolina in USA; many landlocked populations exist. Native to Lake Ontario; introduced into other Great Lakes via Welland Canal

(first taken in Lake Erie in 1931). Introduced elsewhere, including New River in West Virginia and Virginia, and upper Tennessee River system in Tennessee, USA.”

From O’Gorman (2010):

“Established populations of *A. pseudoharengus* in North American waterbodies are mainly in the United States and in the easternmost part of the continent. All but a few are located east of the Mississippi River (USGS, 2009). The lone exceptions are in the north part of the state of Nebraska. Although there are records of alewife in the western state of Colorado (Minckley, 1973; USGS, 2009), the fish was not recently listed as being present in that state (Johnson and Nomanbhoy, 2005).”

“*A. pseudoharengus* had been present in Lake Ontario, the fifth and easternmost of the interconnected Great Lakes, since the mid-1800s and had been abundant there since the late 1870s (O’Gorman and Stewart, 1999). Although Lake Ontario drains to the Atlantic Ocean through the 500-km long St. Lawrence River, it is generally believed that *A. pseudoharengus* is not native to Lake Ontario, mainly because of its absence from the historical record until hundreds of years after the arrival of European colonists. Moreover, there is also genetic evidence that suggests alewife were not indigenous to Lake Ontario (Ihssen et al., 1992).”

From GISD (2017):

“Alewife is native along Atlantic coast of New York, it is however invasive in western New York outside of the Atlantic drainage (Robert O Gorman., pers. comm 2004).”

From Fuller et al. (2025a):

“Introduced populations have been established in 22 US states [...] and throughout the Great Lakes. Introduction to the Youghiogheny River [Maryland] was unsuccessful (Hendricks et al. 1979).”

“The species was first reported from Lake Erie in 1931, Lake Huron in 1933, Lake Michigan in 1949, and Lake Superior in 1954.”

According to Fuller et al. (2025a), *Alosa pseudoharengus* has also been introduced and become established in Georgia, Illinois, Indiana, Kentucky, Maine, Michigan, Minnesota, New Hampshire, North Carolina, Ohio, Pennsylvania, South Carolina, Tennessee, Vermont, Virginia, West Virginia, and Wisconsin. They also report nonindigenous occurrences of *Alosa pseudoharengus* with establishment status unknown in Alabama and Colorado (1973 and 2006, respectively).

Limited records of *Alosa pseudoharengus* in trade in the United States were found (e.g., Anglerssportscenter 2023).

## Regulations

*Alosa pseudoharengus* is regulated in Arkansas (AGFC 2022), Colorado (CPW 2023), Minnesota (Minnesota DNR 2022), Nevada (Nevada Board of Wildlife Commissioners 2022), New Hampshire (NHFG 2022), North Carolina (North Carolina DEQ 2022), Oklahoma (ODWC 2023), Utah (Utah DWR 2023), Washington (Revised Code of Washington 2022), and Wisconsin (Wisconsin DNR 2022). It is regulated at the family level (Clupeidae) in Arizona (Arizona Game and Fish Commission 2022). Please refer back to state agency regulatory documents for details on the regulations, including restrictions on activities involving this species. While effort was made to find all applicable regulations, this list may not be comprehensive. Notably, it does not include regulations that do not explicitly name this species or its genus or family, for example, when omitted from a list of authorized species with blanket regulation for all unnamed species.

From Fuller et al. (2025b):

“While not listed by name, in Ohio it is illegal for any person to possess, import or sell exotic species of fish (including *Alosa pseudoharengus*) or hybrids thereof for introduction or to release into any body of water that is connected to or otherwise drains into a flowing stream or other body of water that would allow egress of the fish into public waters, or waters of the state, without first having obtained permission (OAC Chapter 1501:31-19).”

## Means of Introductions within the United States

From Fuller et al. (2025a):

“Although Smith (1970) brings up the possibility that alewife were introduced into Lake Ontario with American shad stockings in the 1880s, he discounts this possibility in favor of the hypothesis that they reached the lake via the Erie Canal from the Hudson River. He contends that alewife were only able to invade the lake after the decline of predators such as lake trout and Atlantic salmon in the 1860s. Other authors believe, this species was probably native to Lake Ontario (Lee et al. 1980 et seq.) and spread through the Great Lakes via the Welland Canal (Lee et al. 1980 et seq.). [...] The alewife was intentionally stocked in inland waters. The population in the New River, West Virginia, resulted from stockings in Claytor Lake, New River, Virginia (Jenkins and Burkhead 1994). The recently discovered population in Lake St. Catherine, Vermont, is likely a result of an illegal stocking (Good, personal communication). Lakes in the Adirondack Mountains and Otsego Lake, New York were illegally stocked with alewife for forage (Smith 1985; Sinnott, personal communication; D. Warner, personal communication).”

## Remarks

This ERSS was previously published in July 2019. Revisions were completed to incorporate new information and conform to updated standards.

*Alosa pseudoharengus* has landlocked and anadromous populations. The climate matching analysis and the overall risk assessment only consider freshwater populations and life stages; the results are only applicable to freshwater locations.

*Alosa pseudoharengus* has been intentionally stocked outside its native range within the United States by State fishery managers to achieve fishery management objectives. State fish and wildlife management agencies are responsible for balancing multiple fish and wildlife management objectives. The potential for a species to become invasive is now one important consideration when balancing multiple management objectives and advancing sound, science-based management of fish and wildlife and their habitat in the public interest.

From GISD (2017):

“Describing the distribution of alewife is difficult as native and invaded ranges overlap. For example, alewife is native along Atlantic coast of New York, however it is invasive in western New York outside of the Atlantic drainage (Robert O’Gorman, pers. comm. 2004)”

From Fuller et al. (2025a):

“There is apparently disagreement concerning the native status of Alewife in Lake Ontario. Miller (1957) and Smith (1970) point out the first record from Lake Ontario was in 1873. Smith (1970) is of the opinion that it was introduced into the lake. Although Smith (1970) brings up the possibility that Alewife were introduced into Lake Ontario with American Shad stockings in the 1880s, he discounts this possibility in favor of the hypothesis that they reached the lake via the Erie Canal from the Hudson River. He contends that Alewife was only able to invade the lake after the decline of predators such as Lake Trout and Atlantic Salmon in the 1860s. Other authors believe this species was probably native to Lake Ontario (Lee et al. 1980 et seq.) and spread through the Great Lakes via the Welland Canal (Lee et al. 1980 et seq.).”

“Although there is a report of two small alewives taken from the Colorado River, Texas (Bean 1882), we believe this record is in error. Bean (1882) reported that the specimens were sent to Professor Baird at the National Museum. However, a query of the museum's holdings did not return these specimens. We believe the fish are more likely either misidentified *A. chrysochloris* or *A. sapidissima*. *Alosa sapidissima* were stocked in the Colorado River in 1874 (Bean 1882).”

From Fuller et al. (2025b):

“The import, possession, transport, and release of live Alewife in Manitoba is prohibited under articles 6 to 10 of the Canadian Fisheries Act SOR/2015-121.”

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## 2 Biology and Ecology

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### Taxonomic Hierarchy and Taxonomic Standing

From ITIS (2024):

Kingdom Animalia

Subkingdom Bilateria

Infrakingdom Deuterostomia  
Phylum Chordata  
Subphylum Vertebrata  
Infraphylum Gnathostomata  
Superclass Actinopterygii  
Class Teleostei  
Superorder Clupeomorpha  
Order Clupeiformes  
Suborder Clupeoidei  
Family Clupeidae  
Subfamily Alosinae  
Genus *Alosa*  
Species *Alosa pseudoharengus* (Wilson, 1811)

According to Fricke et al. (2025), *Alosa pseudoharengus* (Wilson, 1811) is the current valid name for this species.

## Size, Weight, and Age Range

From Froese and Pauly (2025a):

“Max length : 40.0 cm SL [standard length] male/unsexed; [Robins and Ray 1986]; common length : 30.0 cm SL male/unsexed; [Robins and Ray 1986]; max. published weight: 200.00 g [Robins and Ray 1986]; max. reported age: 9 years [Altman and Dittmer 1962]”

## Environment

From Froese and Pauly (2025a):

“Marine; freshwater; brackish; pelagic-neritic; anadromous [Riede 2004]; depth range 5 - 145 m [Jones et al. 1978], usually 56 - 110 m [Scott and Scott 1988].”

From O’Gorman (2010):

“In the Laurentian Great Lakes, the depths occupied by *A. pseudoharengus* change seasonally and vary somewhat among lakes due to differences in bathymetry and temperature regimes (Wells, 1968; O’Gorman and Schneider, 1986; O’Gorman et al., 2000). In general, *A. pseudoharengus* overwinter offshore in deep water and move shoreward into shallower water in spring to spawn, after which they move back to open waters where they remain throughout the summer, occupying mid to upper levels in the water column (Bergstedt and O’Gorman, 1989; O’Gorman et al., [2013]). In autumn, as the lakes cool, *A. pseudoharengus* descend to greater depths. The depth distribution of *A. pseudoharengus* within a lake can change, however, as it did in Lake Ontario following changes in the lake ecosystem (O’Gorman et al., 2000).”

“In summer, adult *A. pseudoharengus* in the Great Lakes avoid the cold hypolimnetic waters and occupy the warm epilimnetic waters although under certain conditions they [sic] some can be found in the thermocline (Wells, 1968; Olson et al., 1988; Johannsson and O’Gorman, 1991; but see Janssen and Brandt, 1980). Laboratory temperature preferences for adult *A. pseudoharengus*

were 21°C in spring, 16-19°C in summer, and 16 -20°C in autumn (Otto et al., 1976; Spotila et al., 1979). Ultimate upper lethal temperatures were 31-34°C for adult alewives acclimated to 20-27°C (Otto et al., 1976; McCauley and Binkowski, 1982).”

“*A. pseudoharengus* are severely stressed by temperatures lower than 3°C (Colby, 1973). In the Great Lakes *A. pseudoharengus* are in water at 1 to 3°C during most winters. In severe winters, as water temperature may be at or below 1°C (Mortimer, 1971; Rodgers, 1987), mass mortalities of *A. pseudoharengus* sometimes occur (O’Gorman and Schneider, 1986; Bergstedt and O’Gorman, 1989; O’Gorman et al., [2013]). [...] When held in ponds where water temperatures were < 2°C for more than six weeks, *A. pseudoharengus* experienced sublethal immunosuppression, increasing their susceptibility to disease (Lepak and Kraft, 2008).”

## **Climate**

From Froese and Pauly (2025a):

“Temperate; 52°N - 33°N, 93°W - 52°W [Page and Burr 2011]”

## **Distribution Outside the United States**

### **Native**

From Froese and Pauly (2025a):

“North America: Atlantic coast from Red Bay, Labrador in Canada to [...] USA; many landlocked populations exist. [...] Native to Lake Ontario; [...]”

Part of the native range of this species is within the United States; see section 1 for a complete description of the native range.

### **Introduced**

From Fuller et al. (2025b):

“Introduced populations have been established in [...] the Canadian province of Ontario, and throughout the Great Lakes.”

## **Means of Introduction Outside the United States**

From O’Gorman (2010):

“There is no agreement on how *A. pseudoharengus* may have invaded Lake Ontario (Daniels, 2001); some have suggested migration through navigation canals or perhaps inadvertent introduction from planned releases of other fishes (Miller, 1957; Smith, 1970).”

## Short Description

From Fuller et al. (2025b):

“The Alewife is a small herring with a dark dorsal side, bluish to greenish, and light sides with horizontal darker stripes. The head is broad and triangular and the body is relatively deep. Eyes are large with adipose eyelids. A dull black spot is located behind the operculum. Scales are easily rubbed off and form scutes on the midline of the belly. Jaw teeth are inconspicuous and tongue teeth are absent. Caudal fin is forked and lacks an adipose fin. Alewife are visually similar to Blueback Herring (*Alosa aestivalis*), but Alewife has a white peritoneal lining, larger eyes, and a greater body depth than Blueback Herring (Whitehead 1985; Page and Burr 1991; Etnier and Starnes 1993; Jenkins and Burkhead 1994; Scott and Crossman 1998).”

From Froese and Pauly (2025a):

“Dorsal spines (total): 0; Anal spines: 0. Moderately compressed, belly with a distinct keel of scutes. Lower jaw rising steeply within mouth; minute teeth present at front of jaws (disappearing with age). Lower gill rakers increasing with age. A dark spot on shoulder. Distinguished from *A. aestivalis* [blueback herring] by its silvery peritoneum; eye larger than snout length; back greyish green on capture.”

From O’Gorman (2010):

“Eyes are large. The front of the lower jaw is thick and extends past the upper jaw when the mouth is closed. The maxillary extends to below the middle of the eye. [...] There are more than 30 gill rakers on the lower angle of the first gill arch (Trautman, 1957). The single dorsal fin usually has 13-14 rays but may have 12-16. [...] The anal fin is short and wide with 15-19 rays (usually 17-18). The pelvic fins are rather small and contain 10 rays. The pectoral fins are low on the sides and they usually have 16 rays but may have as few as 14 (Scott and Crossman, 1973).”

## Biology

From Froese and Pauly (2025a):

“Occurs in open water over all bottom types [Page and Burr 2011]. Movement of schooling adults apparently restricted to coastal areas proximal to natal estuaries [Jones et al. 1978]. They migrate up rivers and even small streams to spawn in lakes and quiet stretches of rivers, then return to sea shortly after spawning [Jones et al. 1978]; landlocked populations also ascend affluent rivers and streams. Larvae remain in vicinity of spawning grounds, forming schools at sizes less than 10 mm TL [total length], within one to two weeks after hatching [Jones et al. 1978], then descend in summer and autumn or even as late as November or December. Feed on shrimps and small fishes; the young on diatoms, copepods and ostracods while in rivers.”

“Feed on planktonic and benthic invertebrates [Mills et al. 1995]. Fish measuring 5.0-7.0 cm TL feed by gulping, filtering and particulate feeding, whereas smaller fish are exclusive particulate feeders [Lazzaro 1987].”

“Spawning activity has been observed both diurnally and nocturnally, but with greatest activity at night [Edsall 1964]. Spawning activity stops above 27.8°C [Edsall 1970]. Freshwater populations mature earlier and at a smaller average size than saltwater populations [Jones et al. 1978].”

From O’Gorman (2010):

“In the Laurentian Great Lakes, *A. pseudoharengus* are found in schools during daylight hours. At night, the schools break up and some of the fish move to waters near shore. Larval *A. pseudoharengus* are pelagic and are found in bays as well as in nearshore waters around the periphery of Lake Huron and Lake Ontario (O’Gorman, 1983; Klumb et al., 2003). Young-of-year *A. pseudoharengus* in Lake Michigan prefer rocky substrate over sandy substrate when near shore (Janssen and Luebke, 2004). The young of anadromous *A. pseudoharengus* migrate to the Atlantic after spending their first summer in fresh water whereas the young of Great Lakes *A. pseudoharengus* move farther from shore as they grow, overwintering in deep water with their parents.”

“The minimum temperature at which Atlantic *A. pseudoharengus* spawn is 10°C (Cianci, 1965) and spawning peaks at 13-16°C (Richkus, 1974; Tyus, 1974). In landlocked populations, the spawning period is protracted, lasting more than a month (Odell, 1934; Norden, 1967; Hlavek and Norden, 1978). Mean fecundity of Lake Michigan *A. pseudoharengus* was 11,150 for fish averaging 160 mm total length, 16,140 for fish averaging 176 mm, and 22,400 for fish averaging 192 mm (Norden, 1967; Hlavek and Norden, 1978). The non-adhesive eggs are demersal and are broadcast at random over any type of bottom (Odell, 1934; Mansueti, 1956). Optimum temperature for egg incubation is 17.8°C and incubation time varies from 15 days at 7.2°C to 3.7 days at 21.1°C (Edsall, 1970). Alewife larvae average 3.8 mm at hatching, 5.1 mm at yolk sac absorption, and, when held at 20°C, they begin feeding two days after hatching (Norden, 1967; Heinrich, 1981). The larvae are positively phototropic and pelagic (Odell, 1934).”

“*A. pseudoharengus* in Lake Michigan begin spawning when water temperatures reach about 15.6°C; spawning is interrupted when water temperatures exceed 27.8°C (Edsall, 1970). Eggs hatch when incubated at 6.9 to 29.4°C but not at lower or higher temperatures. About 69% of the larvae from eggs incubated at less than 10.6°C are deformed and not likely to survive. Alewife young-of-year (YOY) have been found at 16 to 29°C in Lake Michigan (Brandt, 1980; Dufour et al., 2008). Laboratory studies show a YOY temperature preference range of 21 to 31°C (Otto et al., 1976; Spotila et al., 1979) and critical thermal maximum range of 32 to 34°C for YOY acclimated to 20-25°C (Otto et al., 1976). In the Laurentian Great Lakes, *A. pseudoharengus* must grow to a total length of 60 mm or longer if they are to survive the winter (O’Gorman and Stewart, 1999). *A. pseudoharengus* are rare in Lake Superior, the northern-most Great Lake, presumably because the short growing season does not allow most young *A. pseudoharengus* sufficient time to grow to a size that would allow successful overwintering (O’Gorman et al., 1997; O’Gorman et al., [2013]).”

“Land-locked *A. pseudoharengus* eat zooplankton throughout their life (Morsell and Norden, 1968; Mills et al., 1992, 1995; Stewart et al., 2009). They are size-selective feeders, preferentially eating the largest available zooplankters (Kohler, 1980). Indeed, the movements of



*A. pseudoharengus* around a large lake or the magnitude of an alewife population can sometimes be tracked by the size and species composition of zooplankton (Wells, 1970; Warshaw, 1972; O’Gorman et al., 1991). The first food of larval alewife is cyclopoid copepodites and as the larvae grow they incorporate larger zooplankton in their diet (Heinrich, 1981). When the fish grow to about 110-119 mm they begin to feed on larger invertebrates (Morsell and Norden, 1968) - amphipods, insects, and, in the Laurentian Great Lakes, the opossum shrimp *Mysis diluviana* and the bloody-red shrimp *Hemimysis anomola* (Walsh et al., 2008; Stewart et al., 2009; Lantry et al., 2010). *A. pseudoharengus* are adept at capturing prey in mid-water but have difficulty capturing prey located on or near the lake bottom (Janssen, 1978a). They can feed in the dark (Janssen, 1978b; Janssen et al., 1995; Kelso and Ney, 1983) and they have three modes of feeding: particulate, filtering, and gulping (Janssen, 1976). Nocturnal movement of *A. pseudoharengus* to nearshore areas of lakes to feed has been recorded (Kelso and Ney, 1983). Conversely, *A. pseudoharengus* have been found feeding at night far offshore in the Great Lakes on vertically migrating *Mysis* (Boscarino et al., 2009). *A. pseudoharengus* also eat their own larvae (Odell, 1934; Rhodes et al., 1974) as well as the larvae of other fishes and small young-of-year fishes (Kohler and Ney, 1980; Brandt et al., 1987; Kreuger et al., 1995).”

## Human Uses

From Froese and Pauly (2025a):

“Utilized fresh, dried or salted, smoked and frozen; eaten fried [Frimodt 1995]. Also used for crab and lobster bait and sometimes for pet food [Frimodt 1995].”

From O’Gorman (2010):

“Commercial harvest of the low-value alewife was not economically viable [...]”

## Diseases

**No information was found associating *Alosa pseudoharengus* with any diseases listed by the World Organization for Animal Health (2024).**

From Froese and Pauly (2025a):

“Parasites found are Acanthocephala, cestodes, trematodes and copepods.”

Froese and Pauly (2025b) list *Alosa pseudoharengus* as a host for *Argulus alosae*, *Brachyphallus crenatus*, *Clavellisa cordata*, *Derogenes varicus*, *Echinorhynchus salmonis*, *Hemiurus appendiculatus*, *H. levinseni*, *Kudoa clupeiidae*, *Lechithophyllum botryophoron*, *Lecithaster confusus*, *Lernaeenicus radiatus*, *Mazocraeoides georgei*, *Opecoeloides vitellosus*, and *Siphodera vinaledwardsii*.

Poelen et al. (2014) list *Alosa pseudoharengus* as a host for these additional parasites and pathogens: *Acanthocephalus dirus*, *Anisakis simplex*, *Bothriocephalus scorpii*, *Camallanus oxycephalus*, *Coccidia*, *Contracaecum* sp., *Cyathocephalus truncatus*, *Diphyllbothrium* sp., *Diplostomum spathaceum*, *Eubothrium fragile*, *E. salvelini*, *Goussia ameliae*, *Grillotia erinaceus*, *Hysterothylacium aduncum*, *Ichthyobronema hamulatum*, *Ichthyocotylurus erraticus*,

*Ichthyocphonus hoferi*, *Monostomum* sp., *Mazocraes* sp., *Paenibacillus thiaminolyticus*, *Paratenuisentis ambiguus*, *Posthodiplostomum minimum*, *Pseudoterranova decipiens*, *Scolex pleuronectis*, *S. polymorphus*, and *Tetracotyle* sp.

## Threat to Humans

From Fuller et al. (2025a):

“Periodic large-scale die-offs littered the beaches of the Great Lakes with rotting fish in the 1960's. Such die-offs can pose both a nuisance and a health hazard (Becker 1983).”

## 3 Impacts of Introductions

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From Simonin et al. (2018):

“The observed change in Walleye  $\delta^{15}\text{N}$  values since 1997 is consistent with a change in Walleye diets to more Alewives in 2011. In 1997, nearly all Walleyes sampled in the main lake had consumed only Rainbow Smelt, mostly of lengths 50–175 mm [...]. Our calculations of Walleye trophic levels (based  $\delta^{15}\text{N}$  and a 3.4 unit fractionation per trophic level; Post 2002) resulted in a decline of about one trophic level for Walleyes from 1997 to 2011. This decline is likely caused by consumption of Alewives in 2011, a prey fish feeding at lower trophic levels than adult Rainbow Smelt [...]. Yellow Perch *Perca flavescens* are consumed by Walleyes in other lakes (e.g., Lantry et al. 2008), but our previous research in Lake Champlain found that Yellow Perch were consumed by Walleyes only in the southern region of the lake (Overman and Parrish 2001). This section of the lake does not support Rainbow Smelt so was not included in this study. The change in Walleye diets to prey that feed at lower trophic levels (such as Alewives) will decrease the biomagnification of contaminants and increase the efficiency of energy flow through the ecosystem to Walleyes.”

“Alewives have altered the trophic structure of the fish community in Lake Champlain and are likely to remain as influential as they have been in several of the Laurentian Great Lakes since the mid-20th century (e.g., Crowder 1980; Bunnell et al. 2006; Madenjian et al. 2008).”

From Fuller et al. (2025a):

“The diet of Alewife is diverse and impactful, as it is a proficient feeder of eggs, insects, zooplankton, and larval fish. It selects for the largest zooplankton in invaded water bodies and subsequently has altered the species composition of zooplankton in the Great Lakes where Alewife is abundant (Hutchinson 1971; Johannsson et al. 1991). The degree to which Alewife reduces large zooplankton populations is so severe that the seasonal movements of the fish in Lake Ontario have been tracked by the size of zooplankton (O’Gorman et al. 1991). Alewife also preys on larval fish, benefited by its wide-ranging seasonal movements and ability to feed in midwater and low light levels (O’Gorman et al. 2013).”

“Disappearance of native planktivorous salmonids, such as whitefish, in the Great Lakes has been attributed in part to the introduction of Alewife, which reduced zooplankton populations (Crowder and Binkowski 1983; Todd 1986; Page and Laird 1993). Crowder (1984) speculated

that a cisco native to Lake Michigan, the bloater *Coregonus hoyi*, evolved fewer and shorter gill rakers, and shifted to benthic habitat and diet as a result of competition with Alewives. Smith (1970) attributed the extermination of the lake herring and decline of chub species in the Great Lakes to the Alewife. Smith also talks about the various interrelated changes that took place in each of the Great Lakes as Alewife abundance increased. Christie (1972), on the other hand, argues that the Alewife was not responsible for these changes. The Alewife is the dominant fish in Lake Michigan. It accounts for 70–90% of the fish weight (Becker 1983). Alewife has recently become the dominant prey item for double-crested cormorants in Lake Champlain (DeBruyne et al. 2012). [...] Pothoven et al. (2013) documented an increased abundance of age-0 yellow perch (*Perca flavescens*) and changes in the zooplankton community structure in Saginaw Bay, Lake Huron following the disappearance of alewife, including increased abundance of cladocerans (e.g., *Daphnia* spp., *Bythotrephes*, *Leptodora*) and decreased abundance of cyclopoid copepods.”

“Alewife is a very important species in the history of biological invasions in the Great Lakes. Periodic large-scale die-offs littered the beaches of the Great Lakes with rotting fish in the 1960's. Such die-offs can pose both a nuisance and a health hazard (Becker 1983). Prompted by calls for Alewife management, Pacific salmonids were introduced to both control Alewife populations and utilize alewife as a food source for sport fisheries.”

From Fuller et al. (2025b):

“As the abundance of alewife continued to increase in the absence of predators, massive annual die-offs of alewife began in Lake Ontario, Lake Huron, and Lake Michigan. Beaches and nearshore regions were littered with “huge windrows” of fish (Brown 1968), reportedly removed by bulldozer (Alewife explosion 1967).”

“It was estimated that Alewife populations were responsible for 28% of the total consumption (by wet weight) in Lake Michigan in 1987, and 96% of the total predation on invertebrates in Lake Ontario in 1990 (Rand et al. 1995). The abundance of Alewife combined with a diet preference of zooplankton and larval fishes has been shown to affect both the zooplankton community and certain native fish populations over time. Preference for macrozooplankton and microcrustaceans has shifted the zooplankton community structure towards a prevalence of small species. Following an Alewife decline in Lake Michigan in the mid 1970s, Evans (1990) noted a significant increase in abundance of *Limnocalanus macrurus* and *Diaptomus sicilis*, two of the largest copepods. Similarly, a 1987-1995 study of Lake Ontario found that abundances of cyclopoids and other larger species of zooplankton increased during this period of Alewife decline (Johannsson et al. 1998). Changes in zooplankton abundance and structure caused by Alewife can lead to changes in the phytoplankton community (Shapiro et al. 1975).”

“Disappearance of native planktivorous salmonids, such as lake whitefish (*Coregonus clupeaformis*), in the Great Lakes has been attributed in part to the introduction of Alewife because of reduced zooplankton populations (Crowder and Binkowski 1983, Page and Laird 1993, Todd 1986).”

“In a review of the adverse effects of Alewife on Great Lake fish communities, Madenjian et al. (2008) presented evidence that agreed with Eck and Wells (1987), who stated that Alewife likely has a larger effect on native fish populations through predation of larvae than competition for food resources. Using time-series data for various fish populations along with change point regression analysis, they concluded that predation of larvae by Alewife likely contributed to the decline of yellow perch (*Perca flavescens*), deepwater sculpin (*Myoxocephalus thompsonii*), burbot (*Lota lota*), Atlantic salmon (*Salmo salar*), lake trout (*Salvelinus namaycush*), and emerald shiner (*Notropis atherinoides*) (Madenjian et al. 2008).”

“Furthermore, Alewife has an elevated level of thiaminase, an enzyme that can degrade thiamine in those species that prey on Alewife (Tillitt et al. 2005). Alewife has thus been shown to cause thiamine deficiency and, consequently, early mortality syndrome (EMS) in populations of Alewife predators. EMS and its adverse effects on recruitment and fish populations is well-documented for [...] lake trout, and Atlantic salmon (in which it is also referred to as Cayuga syndrome), among other fishes (Fitzsimons et al. 1999, Ketola et al. 2000, Madenjian et al. 2008). In a spawning reef in Lake Ontario, 50–75% of newly hatched lake trout fry were estimated to suffer from EMS from 1992-1999 (Mills et al. 2005).”

From O’Gorman (2010):

“Establishment of alewife populations in land-locked waterbodies has had both positive and negative economic impacts. The negative impact most noticeable to the general public has been the periodic die-offs of large numbers of fish. As early as 1892, residents of Burlington, Canada, on Lake Ontario, were complaining of the costs of removing the smelly, dead *A. pseudoharengus* that washed ashore in summer (Pritchard, 1929). In 1967, a massive die-off of *A. pseudoharengus* in Lake Michigan (the fifth largest lake in the world) resulted in an estimated loss in excess of 100 million dollars to industry, municipalities, and businesses dependent on recreation (Greenwood, 1970). Other less visible but no less important negative economic impacts include harmful effects on other fishes important to recreational and commercial fisheries.”

“The largest positive economic impact was the development of multi-million dollar recreational fisheries for hatchery-raised salmon and trout in the Laurentian Great Lakes (Bence and Smith, 1999; Connelly and Brown, 2009) -- *A. pseudoharengus* provides an important food source for these fish. These new fisheries are hatchery-dependent, however, and thus require large capital investments to start and continuing investments of capital to maintain.”

*Alosa pseudoharengus* is regulated in at least 10 states, see section 1.

## 4 History of Invasiveness

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The History of Invasiveness for *Alosa pseudoharengus* is classified as High. The established nonnative populations of *A. pseudoharengus* in the Laurentian Great Lakes are well-documented. This species has also been introduced and established outside its native range in several States throughout the eastern United States. *A. pseudoharengus* populations cause large scale changes to zooplankton communities and impact trophic levels and recruitment of native fish species. Consumption of *A. pseudoharengus*, and its elevated level of thiaminase enzyme, by native

salmonids can lead to Early Mortality Syndrome, significantly impacting native salmonid populations.

## 5 Global Distribution

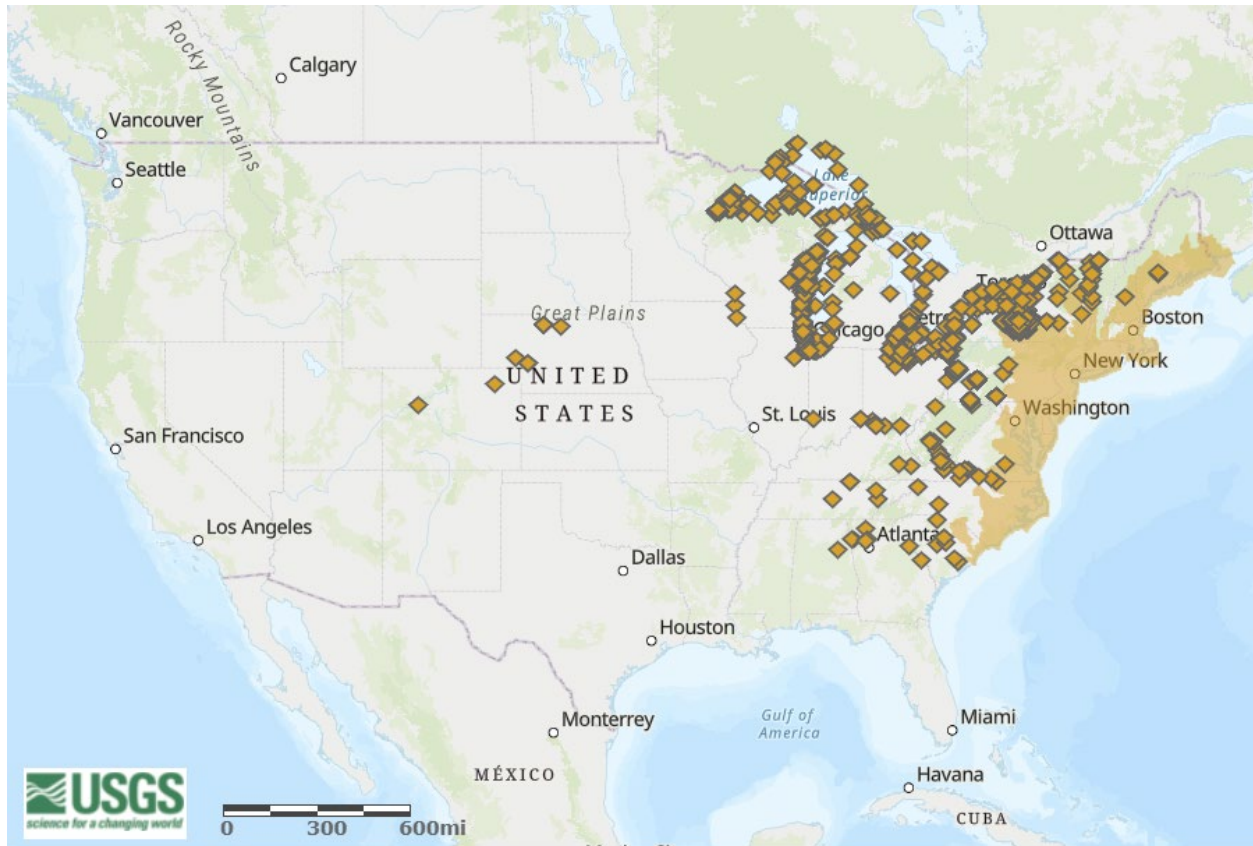
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**Figure 1.** Reported global distribution of *Alosa pseudoharengus*. Map from GBIF Secretariat (2024). Observations are reported from North and South America. The location in Brazil was not used to select source points for the climate matching analysis; the record information indicates that the collection location is incorrect. The locations in Florida, Louisiana, Mississippi, and Mexico were not used to select source locations for the climate matching analysis; occurrence of the species in those locations was not supported in the literature and the observations are not thought to represent established populations. The location in Colorado was not used to select source points for the climate matching analysis; establishment status at this location was unknown.

Marine locations in figure 1 were not used to select source points for the climate matching analysis. The climate matching analysis is only valid for freshwater and brackish water locations.

## 6 Distribution Within the United States



**Figure 2.** Known distribution of *Alosa pseudoharengus* in the United States. Yellow shading indicates the native range of the species. Map from Fuller et al. (2025a). The locations in Alabama and Colorado were not used to select source points for the climate matching analysis; establishment status at these locations is unknown.

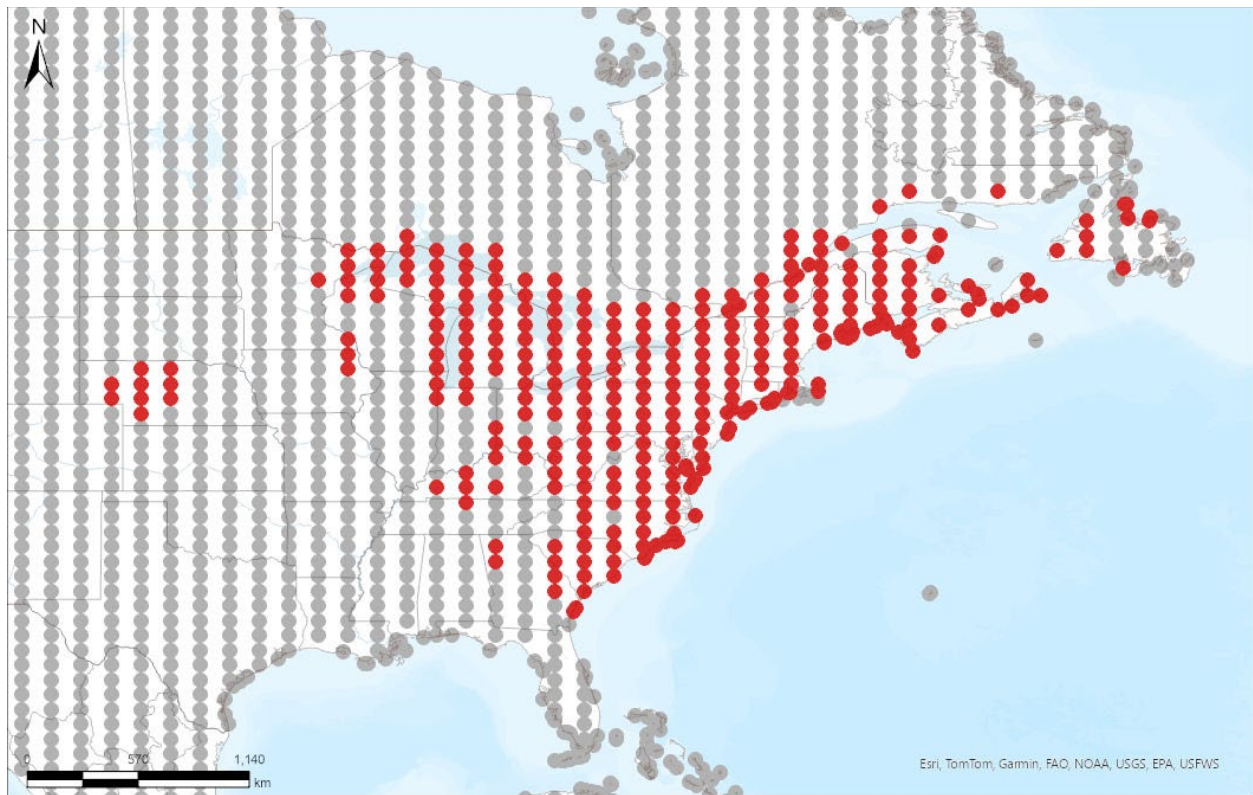
## 7 Climate Matching

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### Summary of Climate Matching Analysis

The climate match for *Alosa pseudoharengus* was generally high in the eastern two thirds of the contiguous United States and low along the Pacific Coast. The climate match was high along the Atlantic Coast from Maine to mid-Florida, along the Gulf Coast from mid-Florida to eastern Texas, and west to the northern and central Great Plains. This area includes the native range of the species along the Atlantic Coast. Southern Florida had a medium climate match. The climate match was also medium in central Texas and low along the Mexican border and the entire Pacific Coast, inland into the Great Basin. The overall Climate 6 score (Sanders et al. 2023; 16 climate variables; Euclidean distance) for the contiguous United States was 0.786, indicating that yes, there is establishment concern for this species outside its native range. The Climate 6 score is calculated as: (count of target points with scores  $\geq 6$ )/(count of all target points). Establishment concern is warranted for Climate 6 scores greater than or equal to 0.002 based on an analysis of the establishment success of 356 nonnative aquatic species introduced to the United States (USFWS 2024).

Projected climate matches in the contiguous United States under future climate scenarios are available for *Alosa pseudoharengus* (see Appendix). These projected climate matches are provided as additional context for the reader; future climate scenarios are not factored into the Overall Risk Assessment Category.

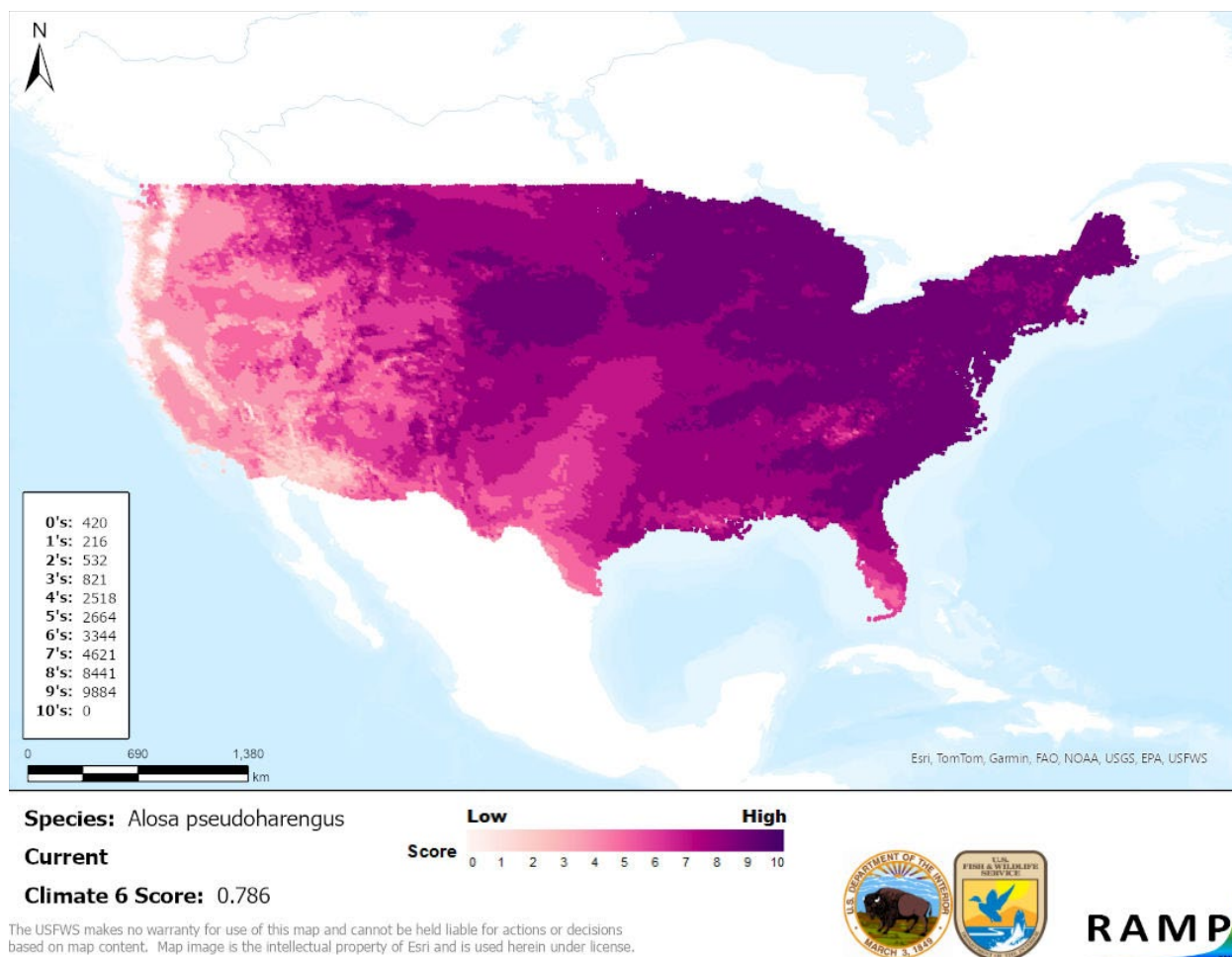


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**Figure 3.** RAMP (Sanders et al. 2023) source map showing weather stations in North America selected as source locations (red; United States and Canada) and non-source locations (gray) for *Alosa pseudoharengus* climate matching. Source locations from GBIF Secretariat (2024). Selected source locations are within 100 km of one or more species occurrences, and do not necessarily represent the locations of occurrences themselves.





**Figure 4.** Map of RAMP (Sanders et al. 2023) climate matches for *Alosa pseudoharengus* in the contiguous United States based on source locations reported by GBIF Secretariat (2024). Counts of climate match scores are tabulated on the left. 0/Pale Pink = Lowest match, 10/Dark Purple = Highest match.

## 8 Certainty of Assessment

The Certainty of Assessment for *Alosa pseudoharengus* is High. Information on the biology, distribution, and history of invasiveness of this species is well-documented in peer-reviewed sources. The literature documents some positive economic impact in addition to negative economic and ecological impacts. The climate matching analysis did not cover the marine portion of the range, but landlocked populations are known.

## 9 Risk Assessment

### Summary of Risk to the Contiguous United States

Alewife, *Alosa pseudoharengus*, is an anadromous marine and freshwater fish native to the Atlantic Coast of North America and associated streams and rivers. Landlocked populations also exist. This species generally consumes plankton and is a prey species in both the native and introduced ranges. There is some human consumption, however it is more commonly used as

bait and for pet food. The History of Invasiveness is High. *A. pseudoharengus* has spread to the Great Lakes most likely through man-made channels, such as the Erie and Welland canals. It was also intentionally stocked or was a possible contaminant of other stocked species elsewhere. It has become established in much of the eastern contiguous United States. *A. pseudoharengus* is an efficient zooplanktivore and causes changes in the zooplankton community. Those changes and direct competition have caused declines in native fishes, especially planktivorous coregonids. Salmonids that consume *A. pseudoharengus* are at risk of a thiamine deficiency, resulting in Early Mortality Syndrome in their offspring which has severely impacted populations. *A. pseudoharengus* is prone to mass die-offs, which are a nuisance for lakeshore property owners and have a negative economic impact on the recreation industry. There may be some beneficial impacts through a reduction in biomagnification of contaminants in Walleye as *A. pseudoharengus* are on a lower trophic level than the Walleye's native prey. *A. pseudoharengus* is regulated in 11 States. The climate matching analysis for the contiguous United States indicates establishment concern outside its native range. Almost the entire eastern two-thirds of the contiguous United States (including the native range of the species) had a high climate match. Low climate matches were found along the Pacific Coast and the Desert Southwest. The climate match is valid for freshwater and brackish water areas only. The Certainty of Assessment is High. The biology of the species, distribution, and history of invasiveness is well documented. The Overall Risk Assessment Category for *Alosa pseudoharengus* is High.

## Assessment Elements

- **History of Invasiveness (see Section 4): High**
- **Establishment Concern (see Section 7): Yes**
- **Certainty of Assessment (see Section 8): High**
- **Remarks, Important additional information: None**
- **Overall Risk Assessment Category: High**

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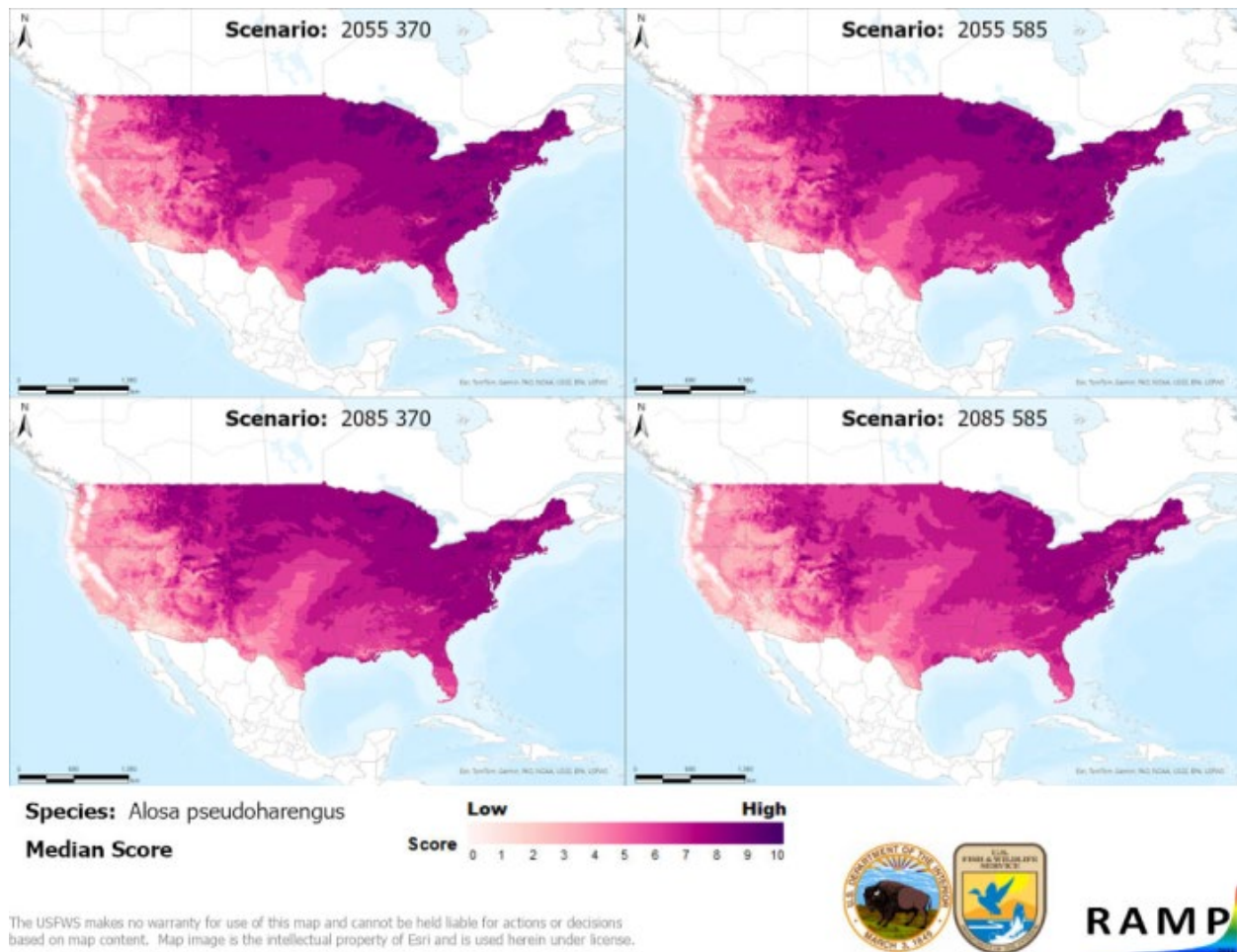
# Appendix

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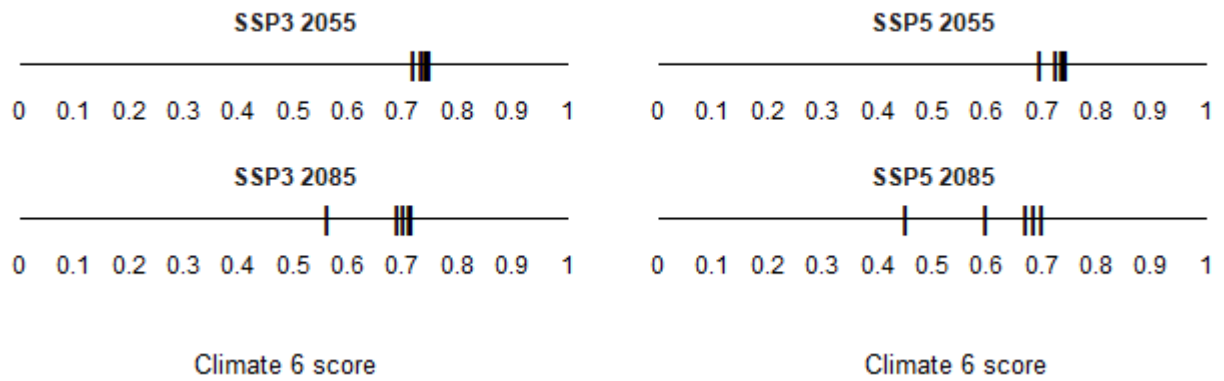
## Summary of Future Climate Matching Analysis

Future climate projections represent two Shared Socioeconomic Pathways (SSP) developed by the Intergovernmental Panel on Climate Change (IPCC 2021): SSP5, in which emissions triple by the end of the century; and SSP3, in which emissions double by the end of the century. Future climate matches were based on source locations reported by GBIF Secretariat (2024).

Under the future climate scenarios (figure A1), on average, high climate match for *Alosa pseudoharengus* was projected to occur in the Appalachian Range, Great Lakes, Mid-Atlantic, Northeast, Northern Plains, and Southern Atlantic Coast regions of the contiguous United States. In general, the areas of high climate match contracted northward between time step 2055 and time step 2085. Areas of low climate match were projected to occur in California and the Northern Pacific Coast regions. The Climate 6 scores for the individual future scenario models (figure A2) ranged from a low of 0.449 (model: UKESM1-0-LL, SSP5, 2085) to a high of 0.746 (model: IPSL-CM6A-LR, SSP3, 2055). All future scenario Climate 6 scores were above the Establishment Concern threshold, indicating that Yes, there was an Establishment Concern for this species. The Climate 6 score for the current climate match (0.786, figure 4) falls above the range of scores for future projections. The time step and climate scenario with the most change relative to current conditions was SSP5, 2085 (figure A3). Under all time step and climate scenarios only minor or no increases in the climate match relative to the current match were observed. Under one or more time step and climate scenarios, areas within the Appalachian Range, Gulf Coast, Northern Plains, Southeast, and Southern Plains saw a large decrease in the climate match relative to current conditions. Additionally, areas within California, the Colorado Plateau, Great Basin, Great Lakes, Mid-Atlantic, Northeast, Southern Atlantic Coast, Southern Florida, Southwest, and Western Mountains saw a moderate decrease in the climate match relative to current conditions. The magnitude of change from current conditions was more pronounced in time step 2085 than in time step 2055 under both scenarios, SSP3 and SSP5.

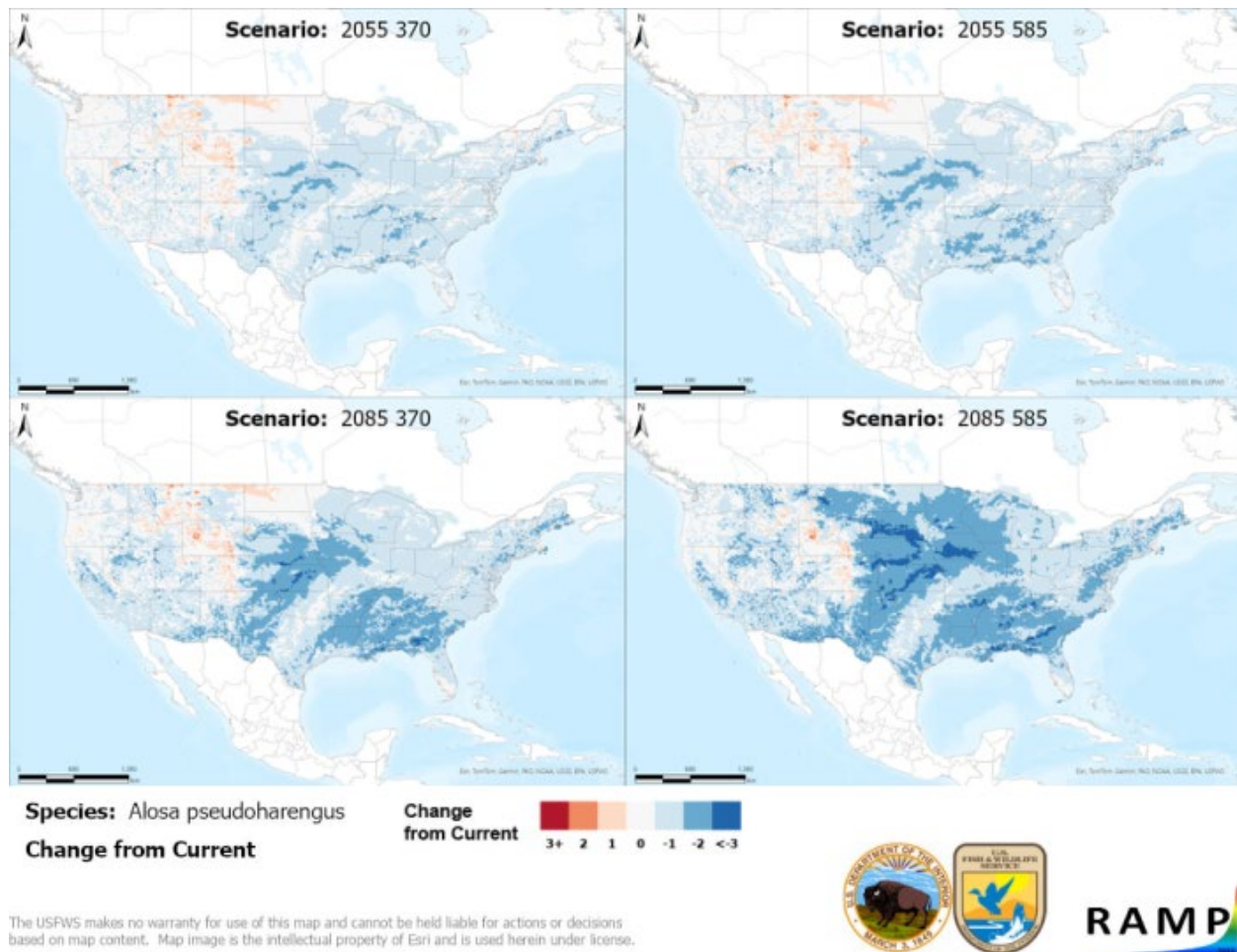


**Figure A1.** Maps of median RAMP (Sanders et al. 2023) climate matches projected under potential future climate conditions using five global climate models for *Alosa pseudoharengus* in the contiguous United States. Climate matching is based on source locations reported by GBIF Secretariat (2024). Shared Socioeconomic Pathways (SSPs) used (from left to right): SSP3, SSP5 (IPCC 2021). Time steps: 2055 (top row) and 2085 (bottom row). Climate source data from CHELSA (Karger et al. 2017, 2018); global climate models used: GFDL-ESM4, UKESM1-0-LL, MPI-ESM1-2-HR, IPSL-CM6A-LR, and MRI-ESM2-0. 0/Pale Pink = Lowest match, 10/Dark Purple = Highest match.



**Figure A2.** Comparison of projected future Climate 6 scores for *Alosa pseudoharengus* in the contiguous United States for each of five global climate models under four combinations of Shared Socioeconomic Pathway (SSP) and time step. SSPs used (from left to right): SSP3, SSP5 (Karger et al. 2017, 2018; IPCC 2021). Time steps: 2055 (top row) and 2085 (bottom row). Climate source data from CHELSA (Karger et al. 2017, 2018); global climate models used: GFDL-ESM4, UKESM1-0-LL, MPI-ESM1-2-HR, IPSL-CM6A-LR, and MRI-ESM2-0.





**Figure A3.** RAMP (Sanders et al. 2023) maps of the contiguous United States showing the difference between the current climate match target point score (figure 4) and the median target point score for future climate scenarios (figure A1) for *Alosa pseudoharengus* based on source locations reported by GBIF Secretariat (2024). Shared Socioeconomic Pathways (SSPs) used (from left to right): SSP3, SSP5 (IPCC 2021). Time steps: 2055 (top row) and 2085 (bottom row). Climate source data from CHELSA (Karger et al. 2017, 2018); global models used: GFDL-ESM4, UKESM1-0-LL, MPI-ESM1-2-HR, IPSL-CM6A-LR, and MRI-ESM2-0. Shades of blue indicate a lower target point score under future scenarios than under current conditions. Shades of red indicate a higher target point score under future scenarios than under current conditions. Darker shades indicate greater change.

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