

# Caspian Mud Shrimp (*Chelicorophium curvispinum*)

## Ecological Risk Screening Summary

U.S. Fish & Wildlife Service, December 2022

Revised, June 2023

Web Version, 3/7/2025

Organism Type: Crustacean

Overall Risk Assessment Category: High



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[https://commons.wikimedia.org/wiki/File:Chelicorophium\\_curvispinum\\_007756447-RMNH.5012316\\_1.jpg](https://commons.wikimedia.org/wiki/File:Chelicorophium_curvispinum_007756447-RMNH.5012316_1.jpg) (December 2022).

# 1 Native Range and Status in the United States

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## Native Range

From Baker et al. (2022):

“**Native Range:** *Chelicorophium curvispinum* originated in the Ponto-Caspian basin, and may have colonized the adjacent Black-Azov basin when they were connected in prehistory (Mordukhai-Boltovskoi 1964). It is native to large river systems discharging into the Black and Caspian Seas, including the lower reaches of the Volga, Dnieper, Dniester, and Danube (de Kluijver and Ingalsuo 1999; Bij de Vaate et al. 2002).”

## Status in the United States

No records of *Chelicorophium curvispinum* in the wild or in trade in the United States were found.

## Regulations

No species-specific regulations on possession or trade were found within the United States; however, there are water ballast regulations in place to prevent introductions of aquatic species into the Great Lakes and Hudson River.

From Baker et al. (2022):

“Ballast water regulations applicable to this species are currently in place to prevent the introduction of nonindigenous species to the Great Lakes via shipping. See Title 33: Code of Federal Regulations, Part 151, Subparts C and D (33 CFR 151 C) for the most recent federal ballast water regulations applying to the Great Lakes and Hudson River.”

## Means of Introductions within the United States

No records of *Chelicorophium curvispinum* in the wild in the United States were found.

## Remarks

From Baker et al. (2022):

“*Chelicorophium curvispinum* often occurs in conjunction with *C. sowinskyi* (Ricciardi and Rasmussen 1998; [Borza et al. 2018]), which has a similar appearance. This can potentially make it difficult to distinguish between the two (Jazdzewski 1980; Jazdzewski and Konopacka 1996).”

From Mastitsky (2009):

“The most likely region to be invaded by this amphipod in the near future is North America (Ricciardi and Rasmussen, 1998).”

## 2 Biology and Ecology

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### Taxonomic Hierarchy and Taxonomic Standing

From Horton et al. (2023):

Animalia (Kingdom) > Arthropoda (Phylum) > Crustacea (Subphylum) > Multicrustacea (Superclass) > Malacostraca (Class) > Eumalacostraca (Subclass) > Peracarida (Superorder) > Amphipoda (Order) > Senticaudata (Suborder) > Corophiida (Infraorder) > Corophiida (Parvorder) > Corophioidea (Superfamily) > Corophiidae (Family) > Corophiinae (Subfamily) > Corophiini (Tribe) > *Chelicorophium* (Genus) > *Chelicorophium curvispinum* (Species)

According to Horton et al. (2023), *Chelicorophium curvispinum* is the current valid name for this species. This species was originally named *Corophium curvispinum* prior to being placed in the genus *Chelicorophium*.

### Size, Weight, and Age Range

From Baker et al. (2022):

“Largest adults up to 8 mm long; overwintering population 2.4–4.5 mm; juveniles < 1.8 mm (van den Brink et al. 1993; Rajagopal et al. 1998).”

“The life span of *C. curvispinum* lasts no longer than 8 months (van der Velde et al. 2000).”

### Environment

From Baker et al. (2022):

“*Chelicorophium curvispinum* is found in salt, brackish, and freshwater (de Kluijver and Ingalsuo 1999). It is originally a brackish water species occurring in salinities of less than 6 ppt (Romanova 1975), with the ability to tolerate very low salinities (Taylor and Harris 1986; Bayliss and Harris 1988; Harris and Bayliss 1990; van den Brink et al. 1993). In Black Sea lagoons and estuaries, its distribution follows the 1.5 ppt isohaline (Bortkevitch 1988). This species is most successful in waters with relatively high ionic content and requires a minimum sodium ion (Na<sup>+</sup>) concentration of 0.5 mM (Harris and Aladin 1997). The lethal minimum oxygen concentration for *C. curvispinum* is 0.300 mg O<sub>2</sub>/L (Dedyu 1980). *Chelicorophium curvispinum* is able to tolerate [water] temperatures from 7.0–31.8°C (Jazdzewski and Konopacka 1990). Populations of this amphipod remain unchanged or gain biomass under conditions of moderate eutrophication (Kotta et al. 2012). It is intolerant of heavy organic pollution levels (Jazdzewski 1980; Harris and Muskó 1999).”

### Climate

From Palomares and Pauly (2023):

“Temperate”

## Distribution Outside the United States

### Native

From Baker et al. (2022):

“**Native Range:** *Chelicorophium curvispinum* originated in the Ponto-Caspian basin, and may have colonized the adjacent Black-Azov basin when they were connected in prehistory (Mordukhai-Boltovskoi 1964). It is native to large river systems discharging into the Black and Caspian Seas, including the lower reaches of the Volga, Dnieper, Dniester, and Danube (de Kluijver and Ingalsuo 1999; Bij de Vaate et al. 2002).”

### Introduced

From Baker et al. (2022):

“**Nonindigenous occurrences:** *Chelicorophium curvispinum* is widespread in Europe and has invaded the upper reaches of the Volga, Dnieper, Dniester, and Danube Rivers (Bij de Vaate et al. 2002; Paunovic et al. 2015) and the North and Baltic Seas (Ojaveer and Kotta 2015; Casties et al. 2016). It is reported in the Sava River in Croatia (Žganec et al. 2009), throughout Great Britain (Moon 1970; Gallardo and Albridge 2015), the Netherlands (van den Brink et al. 1989), France (Bachman et al. 1997), and Sweden (Leppänen et al. 2017).”

From Mastitsky (2009):

“The earliest report of this species was in the Spree-Havel system near Berlin [Germany] in 1912; in 1920s it also was found in Poland where, probably, it had been established for some time (Jazdzewski and Konopacka, 2002; [bij de] Vaate et al., 2002). [...] The most westerly current locality of the species is in Ireland where it was found in 2001 (Lucy et al., 2004). [...] United Kingdom (Crawford, 1935), a country colonised by the species in the early 1930s [...] Nowadays, *C. curvispinum* is widely distributed in Europe [...] (Ricciari and Rasmussen, 1998). In addition to the distribution table records for Belarus [Pripyat River near Mozyr], *C. curvispinum* is also found in the Belarusian section of the Neman River (S Mastitsky, Belarusian State University, Russia, personal communication, 2009).”

“The first records of *C. curvospinum* in the middle Danube River (Hungary) were made as early as the beginning of the twentieth century (Muskó, 1994; Žganec et al., 2009).”

In addition to locations already mentioned, Mastitsky (2009) reports *Chelicorophium curvispinum* as introduced, present, and invasive in Belgium, Bosnia and Herzegovina, Estonia, northern Russia, Serbia, and Ukraine; and as introduced and present in Austria, Lithuania, and Switzerland. Other countries with reported introductions are Czechia (Pergl et al. 2020), Latvia (Balalaikins and Pagad 2020), Slovakia (Pagad 2022), and Luxembourg (Ries and Pagad 2020).

## Means of Introduction Outside the United States

From Mastitsky (2009):

“Thus, there is no doubt that initially *C. curvispinum* colonized Europe through the Dnieper-Bug Canal. The second wave of its invasion in Europe occurred after re-opening of the Maine-Danube canal in 1992 ([bij de] Vaate et al., 2002; Jazdzewski and Konopacka, 2002; Karatayev et al., 2008). [...] It is likely that *C. curvispinum* was transferred to Ireland with ships coming from the United Kingdom (Crawford, 1935), a country colonised by the species in the early 1930s due to ships that were sailing from northern Germany ports ([bij de] Vaate et al., 2002). Introduction of the species into Ireland from continental Europe is also possible.”

“Its initial spread into the Baltic Sea and North Sea drainage systems likely occurred in the second half of the nineteenth century through the canals connecting the Dnieper, Vistula, Oder and Elbe basins ([bij de] Vaate et al., 2002; Jazdzewski and Konopacka, 2002; Karatayev et al., 2008). [...] Through the Oginskiy Canal that linked the Dnieper River (Black Sea basin) with the Neman River (Baltic Sea basin), *C. curvispinum* dispersed to Lithuania, [...]”

“Natural dispersal of *C. curvispinum* occurs by active migrations ([bij de] Vaate et al., 2002; Jazdzewski and Konopacka, 2002; Josens et al., 2005). The speed of active upstream range extension of *C. curvispinum* [sic] may reach up to 15 km/year (Josens et al., 2005).”

## Short Description

From Baker et al. (2022):

“*Chelicorophium curvispinum* has a cylindrical, dorso-ventrally compressed, curled, grey-yellow body with unfused urosomal segments (with spines on uropods 1 and 2) and a small triangular rostrum. It has two pairs of antennae, the first pair slender and the second pair large and thick. The first segment of the first antennae has 3 or 4 spines along the ventral margin and 2 or 3 spines on the inner margin; antennae 1 in females is sparsely setose (hairy). The second antennae are moderately setose, with long hairs; distinguishing features of this species are that the fourth segment of these antennae bears a large curved spur and one or two smaller spurs on its ventrolateral tip, and the fifth segment bears a proximal triangular process on its ventral surface. Females and males differ in the presence of spines and spurs on the inner surface of the second antennae. This species has an evenly convex palm on its most anterior appendage (gnathopod 1), while teeth are present along the inner margin of the last segment (dactylus) of the second gnathopod (de Kluijver and Ingalsuo 1999).”

From Mastitsky (2009):

“As with other gammarids, *C. curvispinum* has a typical laterally compressed, arched, and grey-yellow body of up to 7 mm in length (Van den Brink et al., 1993). Prominent morphological features of this species include:

- very large antennae II, with one long, well developed, and 1-2 smaller spurs on the fourth segment
- 8-10 spines and 1-2 setae on the outer margin of the pedunculus of uropod I, and 4-6 spines on its inner margin

- a sharp triangular rostrum on the head (Eggers and Martens, 2001). Juvenile individuals resemble adults, but are much smaller in size. [...] Carausu (1943), Jazdzewski and Konopacka (1996), [...] Konopacka (2004).”

## Biology

From Baker et al. (2022):

“*Chelicorophium curvispinum* may aggregate on stones, wooden structures, dead and living shells (including those of Dressed mussels), sandy sediments, clay sediment, and submerged aquatic vegetation (den Hartog et al. 1992; van den Brink et al. 1993; Czarnecka et al. 2014; Kurina and Seleznev 2019). It can also be found in association with the green alga *Cladophora* (Kotta et al. 2006). In European waters, *C. curvispinum* density has been extremely variable, with some studies observing 100,000–750,000 individuals/m<sup>2</sup> (highest at 2–3 m depths) (den Hartog et al. 1992; van den Brink et al. 1993), and others observing much lower densities (13–50,000/m<sup>2</sup>) (Harris and Bayliss 1990; Schöll 1990; Kurina 2017; Barabashova et al. 2021). Average densities in more recently invaded territory (Gulf of Finland) were reported to be between 125–1425 individuals/m<sup>2</sup> (Kotta et al. 2006).”

“*Chelicorophium curvispinum* is a tube-dwelling amphipod; it collects minerals and organic particles from the water column and secretes a 1–4 cm thick layer of muddy tubes on colonization surfaces (Paffen et al. 1994). Mud used to build these tubes has ranged from 61 to 609 grams dry weight/m<sup>2</sup> stone surface, smothering previously established organisms (van der Velde et al. 1994). Oxygen consumption, and therefore metabolism, of individuals within tubes (39  $\mu\text{mol/g/h}$ ) is approximately twice as high as that by free swimming individuals (22  $\mu\text{mol/g/h}$ ) outside of tubes (Muskó et al. 1998; Harris and Muskó 1999).”

“This amphipod is a suspension feeder, filtering phytoplankton, especially diatoms, and other suspended matter from the water (van den Brink et al. 1993). This feeding strategy results in population densities being highest where current velocities are strongest (e.g., 1–1.23 m/s), allowing *C. curvispinum* to most efficiently filter the largest quantity of food and obtain the greatest amount of oxygen for metabolism (van den Brink et al. 1993; van der Velde et al. 2000). This species is an important food source for a variety of fish species, including sculpin, eels, perch, ruffe, and pike perch (van den Brink et al. 1993). Other predators include birds, crayfish, and other predatory macroinvertebrates (Biro 1974; Kelleher et al. 1998, 1999; Marguillier et al. 1998).”

“Reproduction in *C. curvispinum* occurs from May to October in the Black Sea (Bortkevitch 1988) and from April to September in the Baltic (van den Brink et al. 1993). These are the warmest periods of the year (water temperature 12–20°C). Sex ratios exhibit a female bias—females outnumber males at all times of year and at many times more than double the male population (van den Brink et al. 1993). Large females (5.00–6.30 mm) become ovigerous a few weeks before smaller individuals (3.80–4.75 mm). Three generations of offspring are produced each year, following an overwintering period—the first in April to May, the second in June to July, and the third in September to October (den Hartog et al. 1992). The progeny of the first generation of summer animals (generation 2), along with the late autumn brood (generation 3), make up the next overwintering generation (Rajagopal et al. 1998).”

“Brooded egg sizes range from 360 x 280 µm (Stage I) to 520 x 440 µm (Stage IV). The number of eggs carried by females and total female body length are correlated, ranging in the Rhine from 3 to 34 eggs (mean = 12) (van den Brink et al. 1993) and in Lake Balaton from 1 to 25 (mean = 6) (Muskó 1990). These differences in clutch size are thought to be due to differences in food availability. Both average clutch size (Rajagopal et al. 1998) and growth rate (Rajagopal et al. 1997) have been positively correlated with the availability of chlorophyll a, which leads to increased planktonic development and greater food availability. Embryonic development lasts about two weeks and larval development takes approximately four weeks. Most rapid growth rates occur from May to August, when water temperatures range from 15–20°C (van den Brink et al. 1993).”

## Human Uses

No information was found on human uses of *Chelicorophium curvispinum*.

## Diseases

**No information was found associating *Chelicorophium curvispinum* with any diseases listed by the World Organisation for Animal Health (2022).**

From Mastitsky (2009):

“In its introduced range, *C. curvispinum* has been documented to host a microsporidian parasite (Prokop et al., 2006) and an acanthocephalan parasite (Van Riel et al., 2003). The latter acanthocephalan species, *Pomphorhynchus* sp., has been suspected to be responsible for the recent decline of *C. curvispinum* in two Dutch rivers (Van Riel et al., 2003).”

Poelen et al. (2014) list *Amphilina foliacea* as a parasite of *Chelicorophium curvispinum*.

## Threat to Humans

No information was found on threats to humans from *Chelicorophium curvispinum*.

# 3 Impacts of Introductions

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From van der Velde et al. (1998):

“Monitoring data on artificial substrates over the years have shown that the macroinvertebrate species richness was reduced at the highest densities of *C. curvispinum* [...]. According to Kinzelbach (1997), *C. curvispinum* also outcompetes the freshwater isopod *Asellus aquaticus* and several species of chironomid larvae. As a result, the numbers of their predators, like leeches, have decreased.”

From Mastitsky (2009):

“*C. curvispinum* is also known as an ecological engineer due to its ability to build mud tubes on hard substrates. For example, in 1989, the population density of this amphipod in the middle and

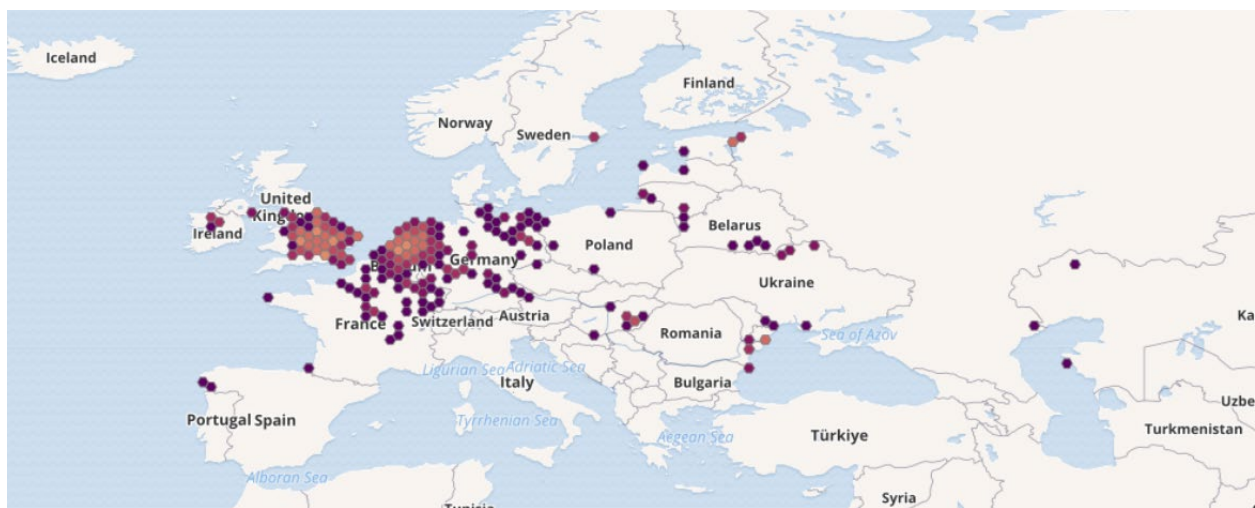
lower sections of the River Rhine was so high that its silty tubes covered all available hard surfaces. In addition, these surfaces became totally covered by fine matter removed by the animals from the water column as a result of their filtering activity. This chain of events affected other epilithic species, which became devoid of a substrate to colonize. The zebra mussel, *Dreissena polymorpha*, [also introduced in this location] was one of the most affected species as the mud tubes of *C. curvispinum* impaired larval settlement of this mollusc (Van den Brink et al., 1991; Van der Velde et al., 1998; Haas et al., 2002). Nevertheless, some epifaunal species may benefit from the presence of the mud tubes of *C. curvispinum*. In the Dnieper River reservoirs, the inter-tube spaces are particularly suitable for the development of communities composed of gammarids (including invasive *Dikerogammarus haemobaphes* and *Dikerogammarus villosus*), oligochaetes, leeches, molluscs, and chironomids (Lubyanov, 1967).”

No species-specific regulations on possession or trade were found within the United States; however, there are water ballast regulations in place to prevent introductions of aquatic species into the Great Lakes and Hudson River.

## 4 History of Invasiveness

*Chelicorophium curvispinum* has established nonnative populations through much of Europe. *C. curvispinum* has been found to outcompete native macroinvertebrate species and reduce macroinvertebrate species richness. This has led to changes in native species abundance at other trophic levels. It has also been documented to alter the substrate and therefore the development of the community of organisms using the substrate. The history of invasiveness for *C. curvispinum* is classified as High due to the impacts on native macroinvertebrate species.

## 5 Global Distribution



**Figure 1.** Reported global distribution of *Chelicorophium curvispinum*. Map from GBIF Secretariat (2022). Observations are reported from Eurasia, spanning from Ireland to southern Russia. The points in Spain and the western coast of France were not used to select source points for the climate matching analysis as they do not represent established populations.



Mastitsky (2009) gave specific locations of known established populations in Belarus, Croatia, Estonia, Ireland, and Ukraine.

Georeferenced observations representing populations across much of the described native range of the “large river systems discharging into the Black and Caspian Seas” (Baker et al. 2022) were not found.

## 6 Distribution Within the United States

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No records of *Chelicorophium curvispinum* in the wild in the United States were found.

## 7 Climate Matching

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### Summary of Climate Matching Analysis

*Chelicorophium curvispinum* had a medium climate match across much of the contiguous United States. Areas of high match occurred around the Great Lakes, Appalachian Mountain range, throughout the Midwest, as well as scattered areas throughout the Great Basin and Colorado Plateau. Areas of low match were found along the West Coast, including the Sierra-Nevada Range, and along the Gulf Coast and Florida. The overall Climate 6 score (Sanders et al. 2023; 16 climate variables; Euclidean distance) for the contiguous United States was 0.772, indicating that Yes, there is establishment concern for this species. The Climate 6 score is calculated as: (count of target points with scores  $\geq 6$ )/(count of all target points). Establishment concern is warranted for Climate 6 scores greater than or equal to 0.002 based on an analysis of the establishment success of 356 nonnative aquatic species introduced to the United States (USFWS 2024).

Projected climate matches in the contiguous United States under future climate scenarios are available for *Chelicorophium curvispinum* (see Appendix). These projected climate matches are provided as additional context for the reader; future climate scenarios are not factored into the Overall Risk Assessment Category.



**Species:** *Chelicorophium curvispinum*

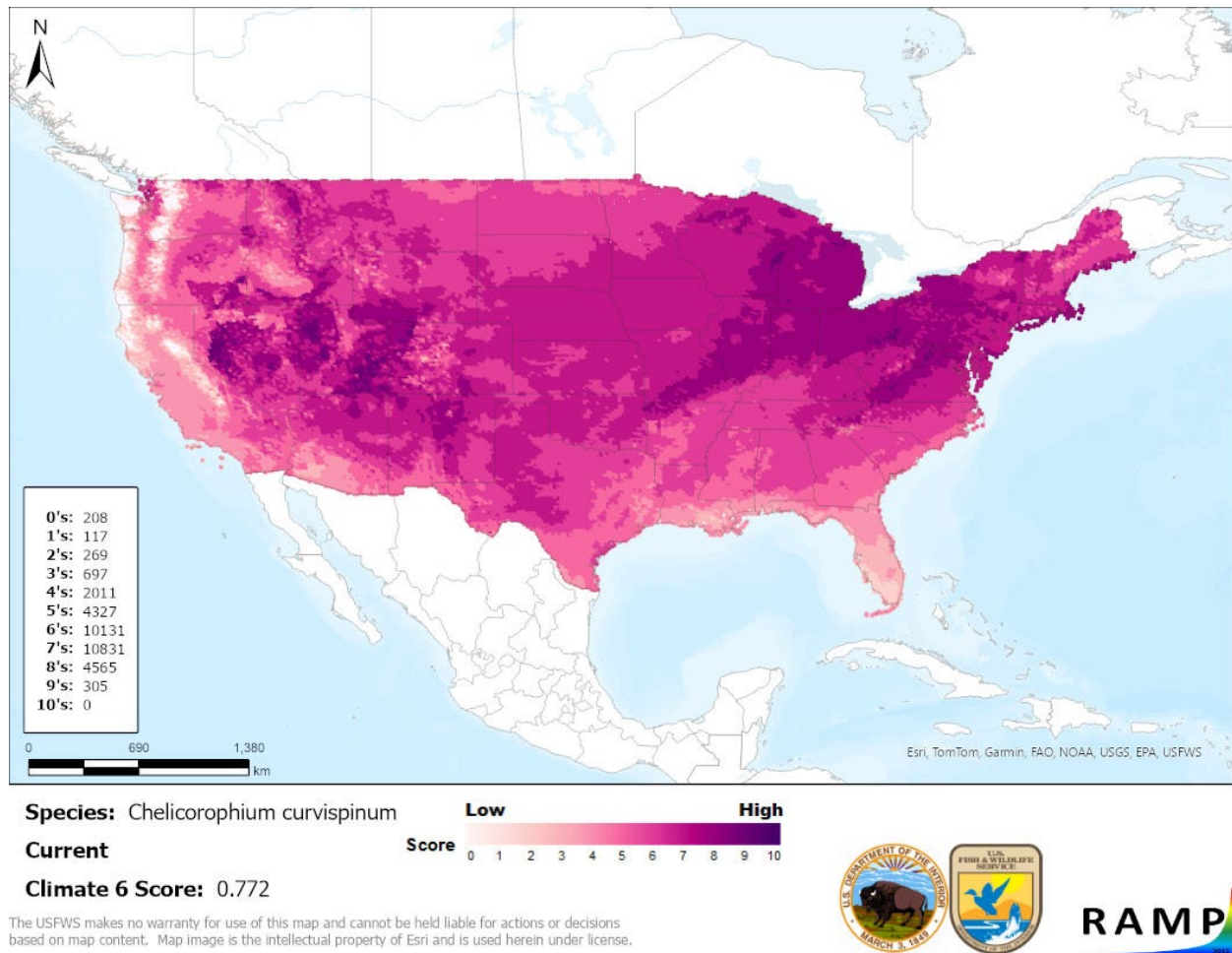
**Selected Climate Stations** ●



**RAMP**

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**Figure 2.** RAMP (Sanders et al. 2023) source map showing weather stations in Eurasia selected as source locations (red; Russia, Kazakhstan, Ukraine, Estonia, Belarus, Romania, Poland, Germany, Czechia, Denmark, Sweden, Netherlands, Belgium, France, Switzerland, United Kingdom, Ireland, Croatia, Bosnia Herzegovina, Hungary, Lithuania, Austria) and non-source locations (gray) for *Chelicorophium curvispinum* climate matching. Source locations from Mastitsky (2009) and GBIF Secretariat (2022). Selected source locations are within 100 km of one or more species occurrences, and do not necessarily represent the locations of occurrences themselves.



**Figure 3.** Map of RAMP (Sanders et al. 2023) climate matches for *Chelicorophium curvispinum* in the contiguous United States based on source locations reported by GBIF Secretariat (2022). Counts of climate match scores are tabulated on the left. 0/Pale Pink = Lowest match, 10/Dark Purple = Highest match.

## 8 Certainty of Assessment

The Certainty of Assessment for *Chelicorophium curvispinum* is classified as High. Records of introduction and establishment were available. Information on impacts from introduced established populations was found; however, much of that information focused on impacts to another introduced species. The distribution of the species is well represented in the source points of the climate match except for some areas potentially within the native range of the species. However, it is not thought that this is enough to impact the result of the climate matching analysis that *C. curvispinum* is a concern for establishment in the contiguous United States.

## 9 Risk Assessment

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### Summary of Risk to the Contiguous United States

*Chelicorophium curvispinum*, Caspian Mud Shrimp, is an amphipod crustacean that is native to lower reaches of the large rivers that drain into the Black and Caspian Seas. This shrimp can be found in salt, brackish, and fresh water. *C. curvispinum* has been introduced throughout Europe from the release of ballast water and sediment as well as natural dispersal through European canal systems. *C. curvispinum* has been found to outcompete native macroinvertebrate species and reduce macroinvertebrate species richness. This has led to changes in abundance of native species at other trophic levels. The History of Invasiveness for *Chelicorophium curvispinum* is classified as High due to the impacts to native macroinvertebrates. The climate matching analysis for the contiguous United States indicates establishment concern for this species. Areas of high match were found around the Great Lakes region and areas of the Great Basin and Colorado Plateau. Much of the remainder of the contiguous United States was found to have a medium match. The Certainty of Assessment is classified as High. The Overall Risk Assessment Category for *Chelicorophium curvispinum* in the contiguous United States is High.

### Assessment Elements

- **History of Invasiveness (see section 4): High**
- **Establishment Concern (see section 7): Yes**
- **Certainty of Assessment (see section 8): High**
- **Remarks, Important additional information: No additional remarks.**
- **Overall Risk Assessment Category: High**

## 10 Literature Cited

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**Note: The following references were accessed for this ERSS. References cited within quoted text but not accessed are included below in section 11.**

- Baker E, Dettloff L, Fusaro A, Bartos A. 2022. *Chelicorophium curvispinum* (G.O. Sars, 1895). Gainesville, Florida: U.S. Geological Survey, Nonindigenous Aquatic Species Database, and Ann Arbor, Michigan: NOAA Great Lakes Aquatic Nonindigenous Species Information System. Available: [https://nas.er.usgs.gov/queries/greatlakes/FactSheet.aspx?Species\\_ID=3612&Potential=Y&Type=2&HUCNumber](https://nas.er.usgs.gov/queries/greatlakes/FactSheet.aspx?Species_ID=3612&Potential=Y&Type=2&HUCNumber) (December 2022).
- Balalaikins M, Pagad S. 2020. Global Register of Introduced and Invasive Species - Latvia. Version 1.3. Invasive Species Specialist Group ISSG. Checklist dataset accessed via Gbif.org. Available: <https://doi.org/10.15468/hb3hza> (October 2024).
- GBIF Secretariat. 2023. GBIF backbone taxonomy: *Chelicorophium curvispinum* (G.O.Sars, 1895). Copenhagen: Global Biodiversity Information Facility. Available: <https://www.gbif.org/species/4314683> (October 2024).

- Horton T, Lowry J, De Broyer C, Bellan-Santini D, Coleman C.O, Corbari L, Costello MJ, Daneliya M, Dauvin JC, Fišer C, Gasca R, Grabowski M, Guerra-García JM, Hendrycks E, Hughes L, Jaume D, Jazdzewski K, Kim YH, King R, Krapp-Schickel T, LeCroy S, Lörz AN, Mamos T, Senna AR, Serejo C, Sket B, Souza-Filho JF, Tandberg AH, Thomas JD, Thurston M, Vader W, Väinölä R, Vonk R, White K, Zeidler W. 2023. *Chelicorophium curvispinum* (G.O. Sars, 1895). World Register of Marine Species. Available: <https://www.marinespecies.org/aphia.php?p=taxdetails&id=148582> (June 2023).
- Mastitsky S. 2009. *Chelicorophium curvispinum* (Caspian mud shrimp). In CABI Compendium. Wallingford, United Kingdom: CAB International. Available: <https://www.cabidigitallibrary.org/doi/10.1079/cabicompendium.108307> (December 2022).
- Pagad S. 2022. Global Register of Introduced and Invasive Species - Slovakia. Version 1.1. Invasive Species Specialist Group ISSG. Checklist dataset accessed via Gbif.org. Available: <https://doi.org/10.15468/9aomvv> (October 2014).
- Palomares MLD, Pauly D, editors. 2023. *Chelicorophium curvispinum*. SeaLifeBase. Available: <https://www.sealifebase.ca/summary/Chelicorophium-curvispinum.html> (June 2023).
- Pergl J, Wong LJ, Pagad S. 2020. Global Register of Introduced and Invasive Species - Czechia. Version 1.2. Invasive Species Specialist Group ISSG. Checklist dataset accessed via Gbif.org. Available: <https://doi.org/10.15468/c5x6ar> (October 2014).
- Poelen JH, Simons JD, Mungall CJ. 2014. Global Biotic Interactions: an open infrastructure to share and analyze species-interaction datasets. *Ecological Informatics* 24:148–159.
- Ries C, Pagad S. 2020. Global Register of Introduced and Invasive Species - Luxembourg. Version 1.2. Invasive Species Specialist Group ISSG. Checklist dataset accessed via Gbif.org. Available: <https://doi.org/10.15468/tvi3gf> (October 2014).
- Sanders S, Castiglione C, Hoff M. 2023. Risk Assessment Mapping Program: RAMP. Version 5.0. U.S. Fish and Wildlife Service.
- van der Velde G, Rajagopal S, Kelleher B, Muskó IB. 1998. Ecological impact of crustacean invaders: General considerations and examples from the Rhine River. In von Vaupel Klein JC, Schram FR, editors. *The biodiversity crisis and Crustacea: Proceedings of the fourth International Crustacean Congress*. Amsterdam, The Netherlands.
- World Organisation for Animal Health. 2022. Animal diseases. Paris: World Organisation for Animal Health. Available: <https://www.woah.org/en/what-we-do/animal-health-and-welfare/animal-diseases/> (December 2022).

## 11 Literature Cited in Quoted Material

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**Note: The following references are cited within quoted text within this ERSS, but were not accessed for its preparation. They are included here to provide the reader with more information.**

- Bachmann V, Usseglio-Polatera P, Cegielka E, Wagner P, Poinsaint JF, Moreteau JC. 1997. Preliminary observations about the coexistence of *Dreissena polymorpha*, *Corophium curvispinum* and *Corbicula* spp. in the river Moselle. Bulletin Français de la Pêche et de la Pisciculture 334-335:373–384.
- Barbashova MA, Trifonova MS, Kurashov EA. 2021. Features of the spatial distribution of invasive amphipod species in the littoral of Lake Ladoga. Russian Journal of Biological Invasions 12(2):136–147.
- Bayliss D, Harris RR. 1988. Chloride ion regulation in the freshwater amphipod *Corophium curvispinum* and acclimatory effects of external Cl<sup>-</sup>? Journal of Comparative Physiology B 158:81–90.
- bij de Vaate A, Jazdzewski K, Ketelaars HAM, Gollash S, van der Velde G. 2002. Geographical patterns in range extension of Ponto-Caspian macroinvertebrate species in Europe. Canadian Journal of Fisheries and Aquatic Sciences 59:1159–1174.
- Biró P. 1974. Observations on the food of eel (*Anguilla anguilla* L.) in Lake Balaton. Tihany, Hungary: Annales Instituti Biologici 41:133–152.
- Bortkevitch LV. 1988. Ecology and production of *Corophium curvispinum* in the estuarine sections of rivers of the northwest Black Sea coast. Hydrobiological Journal 23(6):91–97.
- Borza P, Huber T, Leitner P, Remund N, Graf W. 2018. Niche differentiation among invasive Ponto-Caspian *Chelicorophium* species (Crustacea, Amphipoda, Corophiidae) by food particle size. Aquatic Ecology 52:179–190.
- Casties I, Seebens H, Briski E. 2016. Importance of geographic origin for invasion success: A case study of the North and Baltic Seas versus the Great Lakes–St. Lawrence River region. Ecology and Evolution 6(22):12.
- Carausu S. 1943. Amphipodes de Roumanie 1. Gammarids de type Caspien. Monogr. Inst. Cere. pise. Roumaniei 1:1–293.
- Crawford GI. 1935. *Corophium curvispinum* G. Sars var. *devium* Wundsch, in England. Nature 136:685.
- Czarnecka M, Pilotto F, Pusch MT. 2014. Is coarse woody debris in lakes a refuge or a trap for benthic invertebrates exposed to fish predation? Freshwater Biology 59(11):2400–2412.

- de Kluijver MJ, Ingalsuo SS. 1999. Macrobenthos of the North Sea-Crustacea. *Corophium curvispinum*. Available:  
[http://speciesidentification.org/species.php?species\\_group=crustacea&menuentry=soorten&id=298&tb=beschrijving](http://speciesidentification.org/species.php?species_group=crustacea&menuentry=soorten&id=298&tb=beschrijving) (June 2012).
- Dedyu II. 1980. Amphipods of fresh and salt waters of the south-west part of the USSR. Kishinev, Moldova: Shtiintsa Publishers.
- den Hartog C, van den Brink F, van der Velde G. 1992. Why was the invasion of the River Rhine by *Corophium curvispinum* and *Corbicula* species so successful? Journal of Natural History 26:1121–1129.
- Eggers TO, Martens A. 2001. Identification key for the freshwater Amphipoda (Crustacea) of Germany. Bestimmungsschlüssel der Süßwasser-Amphipoda (Crustacea) Deutschlands. Lauterbornia 42:1–68.
- Gallardo B, Aldridge DC. 2015. Is Great Britain heading for a Ponto–Caspian invasional meltdown? Journal of Applied Ecology 52:41–49.
- Haas G, Brunke M, Strei B. 2002. Fast turnover in dominance of exotic species in the Rhine River determines biodiversity and ecosystem function: an affair between amphipods and mussels. Pages 426–432 in Leppäkoski E, Gollasch S, Olenin S, editors. Invasive aquatic species of Europe: distribution, impacts and management. Dordrecht, The Netherlands: Kluwer Academic Publishers.
- Harris RR, Aladin NV. 1997. The ecophysiology of osmoregulation in Crustacea. Pages 1–25 in Hazon N, Eddy BF, Flik G, editors. Ionic regulation in animals: a tribute to professor W.T.W. Potts. Heidelberg, Germany: Springer Berlin.
- Harris RR, Bayliss D. 1990. Osmoregulation in *Corophium curvispinum* (Crustacea: Amphipoda), a recent coloniser of freshwater. Journal of Comparative Physiology B 160:85–92.
- Harris RR, Musko I. 1999. Oxygen consumption, hypoxia, and tube-dwelling in the invasive amphipod *Corophium curvispinum*. Journal of Crustacean Biology 19(2):224–234.
- Jazdzewski K. 1980. Range extensions of some Gammaridean species in European inland waters caused by human activity. Crustaceana 6:84–107.
- Jazdzewski K, Konopacka A. 1990. New, interesting locality of the Ponto-caspian gammarid *Echinogammarus ischnus* (Stebbing, 1898) (Crustacea, Amphipoda) in Poland. Przegląd Zoologiczny 34:101–112.
- Jazdzewski K, Konopacka A. 1996. Remarks on the morphology, taxonomy and distribution of *Corophium curvispinum* G. Sars, 1885 and *Corophium sowinskyi* Martynov, 1924

- (Crustacea, Amphipoda, Corophiidae). Bollettino del Museo Civico di Storia Naturale di Verona 20:487–501.
- Jazdzewski K, Konopacka A. 2002. Invasive Ponto-Caspian species in waters of the Vistula and Oder basins and the southern Baltic Sea. Pages 384–398 in Leppäkoski E, Gollasch S, Olenin. Invasive aquatic species of Europe: distribution, impacts and management. Dordrecht, The Netherlands: Kluwer Academic Publishers.
- Josens G, Bij de Vaate A, Usseglio-Polatera P, Cammaerts R, Cherot F, Grisez F, Verboonen P, Van den Bossche JP. 2005. Native and exotic Amphipoda and other Peracarida in the River Meuse: new assemblages emerge from a fast changing fauna. Hydrobiologia 542:203–220.
- Karatayev AY, Mastitsky SE, Burlakova LE, Olenin S. 2008. Past, current, and future of the central European corridor for aquatic invasions in Belarus. Biological Invasions 10:215–232.
- Kelleher B, Bergers PJM, van den Brink FWB, Giller FWB, Van der Velde G, Bij de Vaate A. 1998. Effects of exotic amphipod invasions on fish diet in the Lower Rhine. Archiv für Hydrobiologie 143:363–382.
- Kelleher B, Van der Velde G, Wittmann KJ, Faasse MA, Bij de Vaate A. 1999. Current status of the freshwater Mysidae in the Netherlands, with records of *Limnomysis benedeni* Czerniavsky, 1882, a Pontocaspian species in Dutch Rhine branches. Universiteit van Amsterdam: Bulletin Zoologisch Museum 16(13):89–94.
- Kinzelbrach R. 1997. Aquatische Neozoen in Europa. Universität Rostock, Allgemeine und Spezielle Zoologie, Newsletter der Arbeitsgruppe Neozoen.
- Konopacka A. 2004. Invasive amphipods (Crustacea, Amphipoda) in Polish waters. Przegląd Zoologiczny XLVIII:141–162.
- Kotta J, Herkül K, Kotta I, Orav-Kotta H. 2006. Invasion history and distribution of the key benthic alien invertebrate species in the Estonian coastal sea. Estonian Marine Institute Report Series Number 14. Pages 12–18 in Ülikool T, Ojaveer H, Kotta J, editors. Alien invasive species in the north-eastern Baltic Sea: population dynamics and assessment of ecological impacts. Tallinn: Estonian Marine Institute.
- Kotta J, Lauringson V, Kaasik A, Kotta I. 2012. Defining the coastal water quality in Estonia based on benthic invertebrate communities. Estonian Journal of Ecology 61(2):86–105.
- Kurina EM. 2017. Alien species of amphipods (Amphipoda, Gammaridea) in the bottom communities of the Kuybyshev and Saratov reservoirs: Features of distribution and life cycle strategies. Russian Journal of Biological Invasions 8(3):251–260.



- Kurina EM, Seleznev DG. 2019. Analysis of the patterns of organization of species complexes of Ponto-Caspian and Ponto-Azovian macrozoobenthos in the Middle and Lower Volga Reservoirs. *Russian Journal of Ecology* 50:65–74.
- Leppänen JJ, Kotta J, Daneliya M, Salo E. 2017. First record of *Chelicorophium curvispinum* (G.O. Sars, 1895) from Lake Mälaren, SE Sweden. *BioInvasions Records* 6(4):345–349.
- Lubyanov IP, Buzakova AM, Gaydash YK. 1967. Changes in composition of macro- and microzoobenthos of the Dneprovskoe Reservoir after establishment of regulated run-off in the middle course of the Dnieper River. Pages 167–175 in Vladimirova KS, Zerov KK, Melnichuk GL, Olivari GA, Tseeb YY, editors. *Hydrobiological regime of the Dnieper River under conditions of regulated run-off*. Kiev, Ukraine: Naukova Dumka Press.
- Lubyanov IP, Fatovenko MA. 1967. First stages of formation of the bottom fauna of the Dneprodzerzhinskoe Reservoir. Pages 147–158 in Vladimirova KS, Zerov KK, Melnichuk GL, Olivari GA, Tseeb YY, editors. *Hydrobiological regime of the Dnieper River under conditions of regulated run-off*. Kiev, Ukraine: Naukova Dumka Press.
- Lucy F, Minchin D, Holmes JMC, Sullivan M. 2004. First records of the Ponto-Caspian amphipod *Chelicorophium curispinum* (Sars, 1895) in Ireland. *Irish Naturalists Journal* 27:461–464.
- Marguillier S, Dehairs F, Van der Velde G, Kelleher B, Rajagopal S. 1998. Initial results on the trophic relationships based on *Corophium curvispinum* in the Rhine traced by stable isotopes. Pages 171–177 in Nienhuis PH, Leuven RSEW, Ragas AMJ, editors. *New concepts for sustainable management of river basins*. Leiden, The Netherlands: Backhuys Publishers.
- Moon HP. 1970. *Corophium curvispinum* (Amphipoda) recorded again in the British Isles. *Nature* 226(5249):976–976.
- Mordukhai-Boltovskoi. 1964. Caspian fauna beyond the Caspian Sea. *Internationale Revue der gesamten Hydrobiologie und Hydrographie* 49:139–176.
- Muskó IB. 1990. Qualitative and quantitative relationships of Amphipoda (Crustacea) living on macrophytes in Lake Balaton (Hungary). *Hydrobiologia* 191:269–274.
- Muskó IB. 1994. Occurrence of Amphipoda in Hungary since 1853. *Crustaceana* 66:144–152.
- Muskó IB, Tóth LG, Szabó E. 1998. Respiratory energy loss of *Corophium curvispinum* (Crustacea: Amphipoda) in Lake Balaton (Hungary) during the vegetation period. *International Association for Theoretical and Applied Limnology: Negotiations* 26:2107–2114.

- Ojaveer H, Kotta J. 2015. Ecosystem impacts of the widespread non-indigenous species in the Baltic Sea: literature survey evidences major limitations in knowledge. *Hydrobiologia* 750:171–185.
- Paffen B, van den Brink F, van der Velde G, Bij de Vaate A. 1994. The population explosion of the Amphipod *Corophium curvispinum* in the Dutch Lower Rhine. *Water Science and Technology* 29(3):53–55.
- Paunovic M, Csányi B, Simonovic P, Zoric K. 2015. Invasive alien species in the Danube. Pages 389–410 in Liska I, editor. *The Danube River Basin*. Volume 39. Heidelberg, Germany: Springer Berlin.
- Prokop ZM, Dunn AM, Smith JE, Grabowski M. 2006. Genetic diversity and microsporidian parasites in a Ponto-Caspian invader *Chelicorophium curvispinum*. [Pages not provided by source] in Abstracts of the Symposium "An Evolutionary Aspect of Biological Invasions". Fribourg, Switzerland.
- Rajagopal S, van der Velde G, Paffen B, Bij de Vaate A. 1997. Ecology and impact of the exotic amphipod, *Corophium curvispinum* Sars, 1895 (Crustacea: Amphipoda), in the River Rhine and Meuse. Lelystad, The Netherlands: Institute for Inland Water Management and Waste Water Treatment (RIZA).
- Rajagopal S, van der Velde G, Paffen B, Bij de Vaate A. 1998. Crustaceans and the biodiversity crisis: proceedings of the Fourth International Crustacean Congress. Page 458–472 in *Growth and production of Corophium curvispinum* GO Sars, 1895 (Amphipoda), an invader in the lower Rhine. Amsterdam, The Netherlands.
- Ricciardi A, Rasmussen JB. 1998. Predicting the identity and impact of future biological invaders: a priority for aquatic resource management. *Canadian Journal of Aquatic and Fisheries Science* 55:1759–1765.
- Romanova N. 1975. Quantitative distribution and ecology of corophiids (Crustacea, Amphipoda, Corophium). *Bulletin of the Moscow Society of Naturalists. Biological Department* 80(3):51–63.
- Schöll F. 1990. The occurrence of *Corophium curvispinum* SARS in the river Rhine. *Lauterbornia* 58:67–70.
- Taylor PM, Harris RR. 1986. Osmoregulation in *Corophium curvispinum* (Crustacea: Amphipoda), a recent coloniser of freshwater. *Journal of Comparative Physiology B* 156(3):323–329.
- van den Brink FWB, van der Velde G, Bij de Vaate A. 1989. A note on the immigration of *Corophium curvispinum* Sars, 1895 (Crustacea: Amphipoda) into the Netherlands via the River Rhine. *Bulletin of the Zoological Museum of the University of Amsterdam* 11(26):211–213.

- van den Brink FWB, van der Velde G, Bij de Vaate A. 1991. Amphipod invasion on the Rhine. *Nature* 352:576.
- van den Brink FWB, van der Velde G, Bij de Vaate A. 1993. Ecological aspects, explosive range extension and impact of a mass invader, *Corophium curvispinum* Sars, 1895 (Crustacea: Amphipoda), in the Lower Rhine (The Netherlands). *Oecologia* 93:224–232.
- van Riel MC, van der Velde G, Bij de Vaate A. 2003. *Pomphorynchus spec.* (Acanthocephala) uses the invasive amphipod *Chelicorophium curvispinum* (G. Sars, 1895) as an intermediate host in the River Rhine. *Crustaceana* 76:241–246.
- van der Velde G, Paffen BGP, van den Brink FWB. 1994. Decline of zebra mussel populations in the Rhine—Competition between two mass invaders (*Dreissena polymorpha* and *Corophium curvispinum*). *Naturwissenschaften* 81:32–34.
- van der Velde G, Rajagopal S, van den Brink FWB, Kelleher B, Paffen BGP, Kempers AJ, Bij de Vaate A. 1998. Ecological impact of an exotic invasion in the River Rhine. Pages 159–169 in Nienhuis PH, Leuven RSEW, Ragas AMJ, editors. *New concepts for sustainable management of river basins*. Leiden, Netherlands: Backhuys Publishing.
- van der Velde G, Rajagopal S, Kelleher B, Musko IB, Bij de Vaate A. 2000. Ecological impacts of crustacean invaders: general considerations and examples from the Rhine River. *Crustacean Issues* 12:3–33.
- Žganec K, Gottstein S, Hudina S. 2009. Ponto-Caspian amphipods in Croatian large rivers. *Aquatic Invasions* 4:327–335.

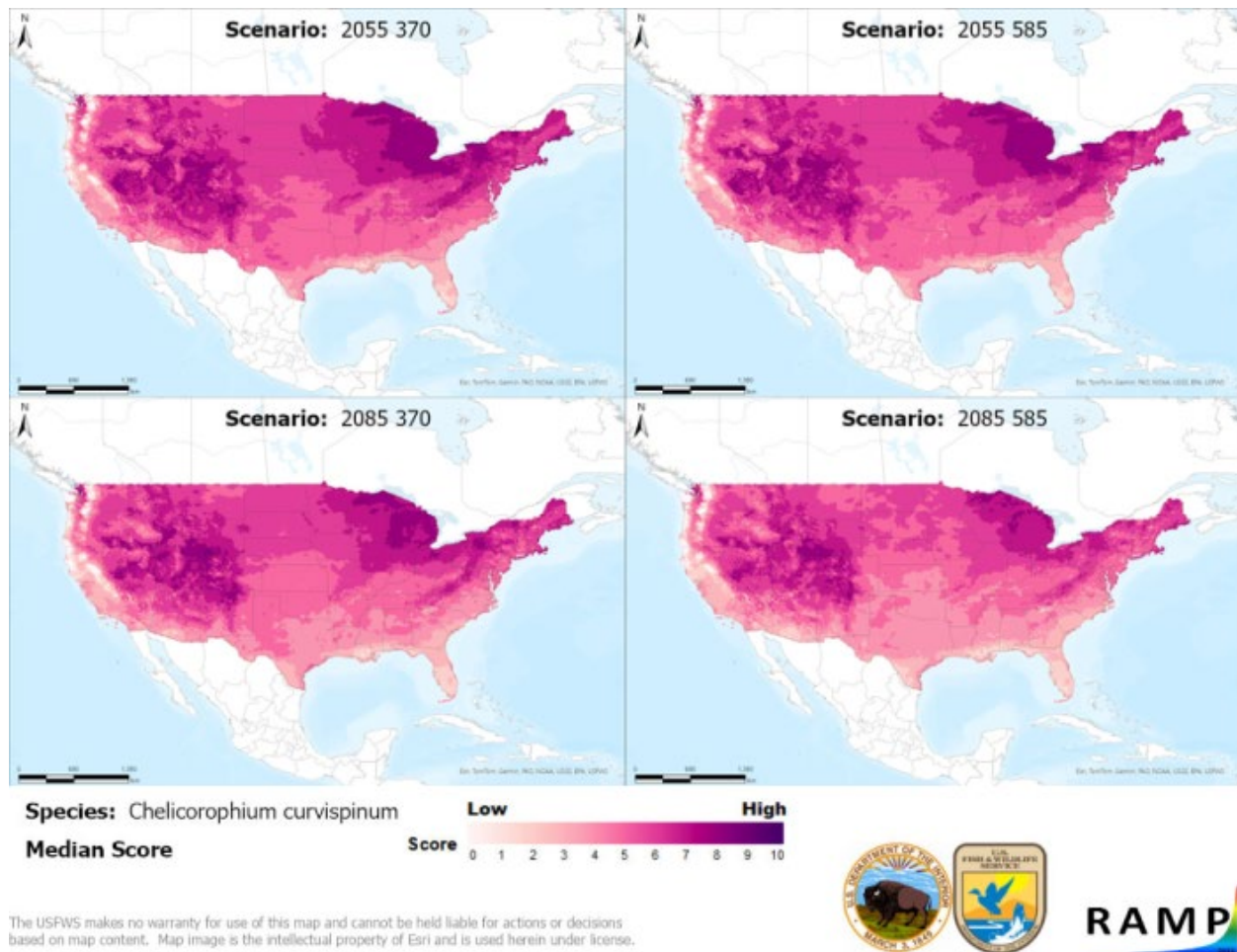
# Appendix

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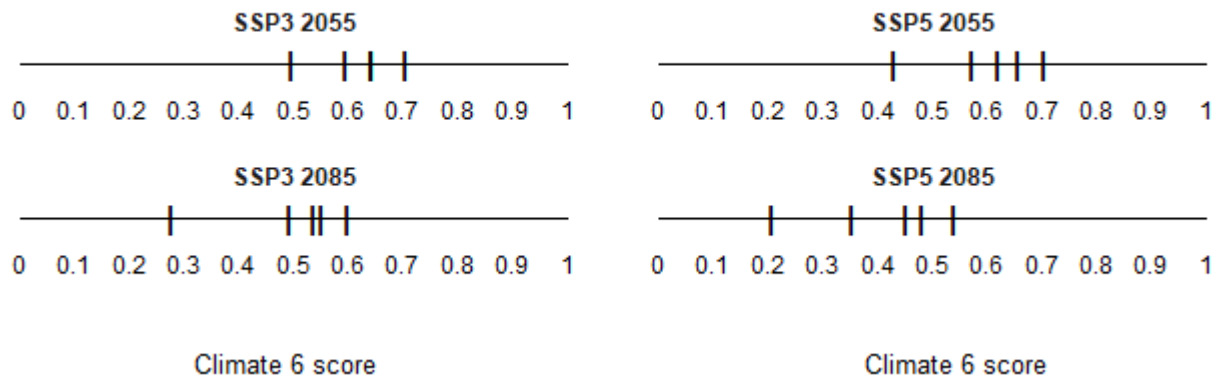
## Summary of Future Climate Matching Analysis

Future climate projections represent two Shared Socioeconomic Pathways (SSP) developed by the Intergovernmental Panel on Climate Change (IPCC 2021): SSP5, in which emissions triple by the end of the century; and SSP3, in which emissions double by the end of the century. Future climate matches were based on source locations reported by Mastitsky (2009) and GBIF Secretariat (2022).

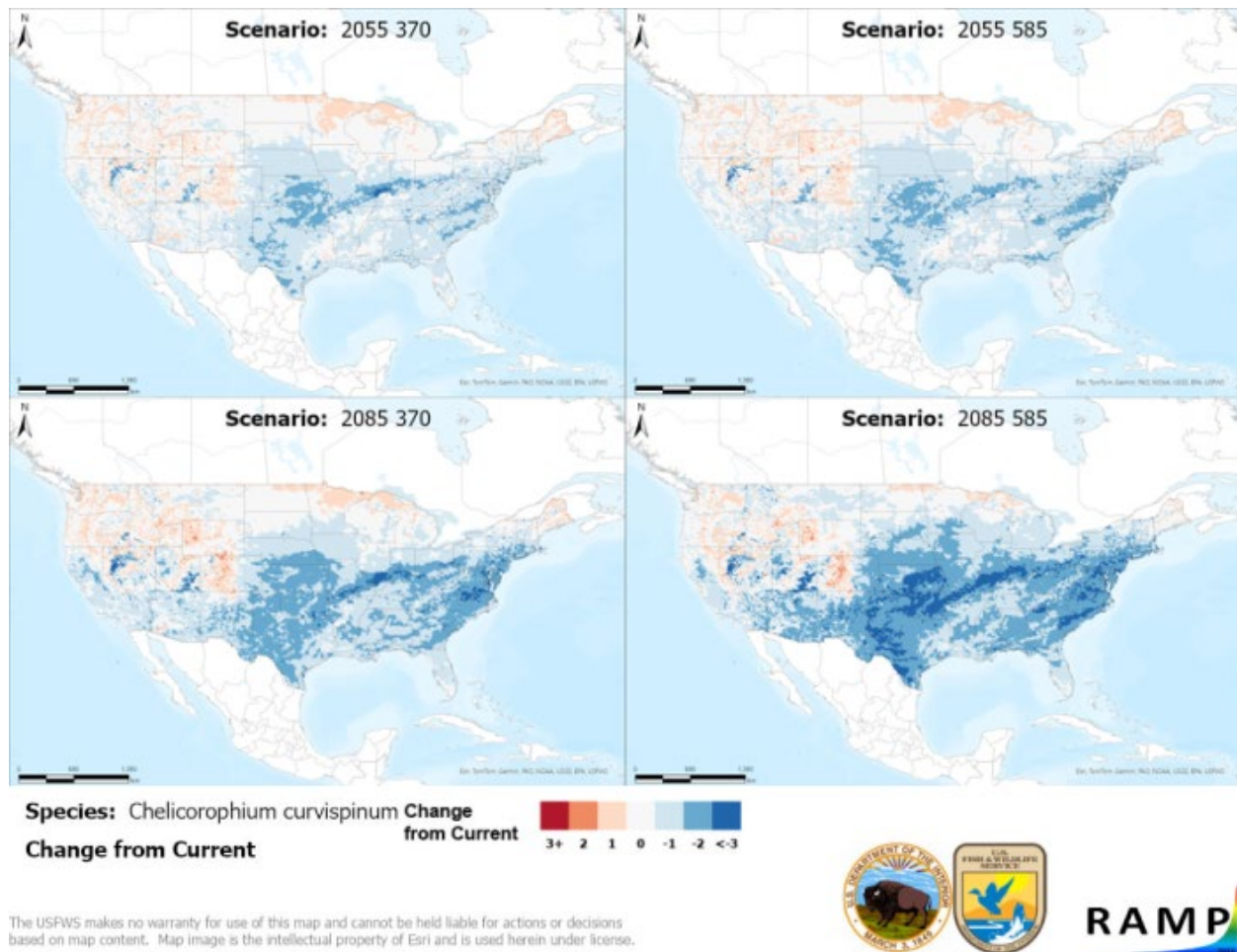
Under the future climate scenarios (figure A1), on average, high climate match for *Chelicorophium curvispinum* was projected to occur in the Great Lakes region of the contiguous United States. There were also areas of high match in the Great Basin and Colorado Plateau under most scenarios. Areas of low climate match were projected to occur in the Northern Pacific Coast region and Southern Florida as well as the Sierra Nevada range. In time step 2085 areas of low match were also found along the Gulf Coast. The Climate 6 scores for the individual future scenario models (figure A2) ranged from a low of 0.204 (model: UKESM1-0-LL, SSP5, 2085) to a high of 0.703 (model: MPI-ESM1-2-HR, SSP3, 2055). All future scenario Climate 6 scores were above the Establishment Concern threshold, indicating that Yes, there is establishment concern for this species under future scenarios. The Climate 6 score for the current climate match (0.772, figure 3) falls above the range of scores for future projections. The time step and climate scenario with the most change relative to current conditions was SSP5, 2085, the most extreme climate change scenario. Under one or more time step and climate scenarios, areas within the Colorado Plateau saw a moderate increase in the climate match relative to current conditions. No large increases were observed regardless of time step and climate scenarios. Under one or more time step and climate scenarios, areas within the Appalachian Range, Colorado Plateau, Great Basin, Great Lakes, Gulf Coast, Mid-Atlantic, Southeast, Southern Atlantic Coast, Southern Plains, and Southwest saw a large decrease in the climate match relative to current conditions. Additionally, areas within California, The Northeast, and Northern Plains saw a moderate decrease in the climate match relative to current conditions. Additionally, very small areas of large or moderate change may be visible on the maps (figure A3).



**Figure A1.** Maps of median RAMP (Sanders et al. 2023) climate matches projected under potential future climate conditions using five global climate models for *Chelicerophium curvispinum* in the contiguous United States. Climate matching is based on source locations reported by Mastitsky (2009) and GBIF Secretariat (2022). Shared Socioeconomic Pathways (SSPs) used (from left to right): SSP3, SSP5 (IPCC 2021). Time steps: 2055 (top row) and 2085 (bottom row). Climate source data from CHELSA (Karger et al. 2017, 2018); global climate models used: GFDL-ESM4, UKESM1-0-LL, MPI-ESM1-2-HR, IPSL-CM6A-LR, and MRI-ESM2-0. 0/Pale Pink = Lowest match, 10/Dark Purple = Highest match.



**Figure A2.** Comparison of projected future Climate 6 scores for *Chelicorophium curvispinum* in the contiguous United States for each of five global climate models under four combinations of Shared Socioeconomic Pathway (SSP) and time step. SSPs used (from left to right): SSP3, SSP5 (Karger et al. 2017, 2018; IPCC 2021). Time steps: 2055 (top row) and 2085 (bottom row). Climate source data from CHELSA (Karger et al. 2017, 2018); global climate models used: GFDL-ESM4, UKESM1-0-LL, MPI-ESM1-2-HR, IPSL-CM6A-LR, and MRI-ESM2-0.



**Figure A3.** RAMP (Sanders et al. 2023) maps of the contiguous United States showing the difference between the current climate match target point score (figure 3) and the median target point score for future climate scenarios (figure A1) for *Chelicorophium curvispinum* based on source locations reported by Mastitsky (2009) and GBIF Secretariat (2022). Shared Socioeconomic Pathways (SSPs) used (from left to right): SSP3, SSP5 (IPCC 2021). Time steps: 2055 (top row) and 2085 (bottom row). Climate source data from CHELSA (Karger et al. 2017, 2018); global models used: GFDL-ESM4, UKESM1-0-LL, MPI-ESM1-2-HR, IPSL-CM6A-LR, and MRI-ESM2-0. Shades of blue indicate a lower target point score under future scenarios than under current conditions. Shades of red indicate a higher target point score under future scenarios than under current conditions. Darker shades indicate greater change.

## Literature Cited

- GBIF Secretariat. 2023. GBIF backbone taxonomy: *Chelicorophium curvispinum* (G.O.Sars, 1895). Copenhagen: Global Biodiversity Information Facility. Available: <https://www.gbif.org/species/4314683> (October 2024).
- [IPCC] Intergovernmental Panel on Climate Change. 2021. Climate change 2021: the physical science basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press.
- Karger DN, Conrad O, Böhner J, Kawohl T, Kreft H, Soria-Auza RW, Zimmermann NE, Linder P, Kessler M. 2017. Climatologies at high resolution for the Earth land surface areas. *Scientific Data* 4:170122.
- Karger DN, Conrad O, Böhner J, Kawohl T, Kreft H, Soria-Auza RW, Zimmermann NE, Linder HP, Kessler M. 2018. Data from: Climatologies at high resolution for the earth's land surface areas. *EnviDat*. Available: <https://doi.org/10.16904/envidat.228.v2.1>.
- Mastitsky S. 2009. *Chelicorophium curvispinum* (Caspian mud shrimp). In *CABI Invasive Species Compendium*. Wallingford, United Kingdom: CAB International. Available: <https://www.cabidigitallibrary.org/doi/10.1079/cabicompendium.108307> (December 2022).
- Sanders S, Castiglione C, Hoff M. 2023. Risk Assessment Mapping Program: RAMP. Version 5.0. U.S. Fish and Wildlife Service.