INVESTIGATIONS IN FISH CONTROL

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- 18. Toxicity of 22 Therapeutic Compounds—to Six Fishes
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- 20. Toxicity of Dimethyl Sulfoxide (DMSO) to Fish
- 21. Labor-Saving Devices for Bioassay Laboratories



United States Department of the Interior
Fish and Wildlife Service
Bureau of Sport Fisheries and Wildlife

18. Toxicity of 22 Therapeutic Compounds to Six Fishes

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Fish and Wildlife Service, Clarence F. Pautzke, Commissioner
Bureau of Sport Fisheries and Wildlife, John S. Gottschalk, Director
Resource Publication 35 • Washington, D.C. • December 1966

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TOXfeffy) でデーク22 THERAPEUTIC COMPOUNDS TO SIX FISHES

By Wayne A. Willford, Chemist Fish Control Laboratory, La Crosse, Wis.

ABSTRACT.--Of 22 therapeutic chemicals (18 parasiticides and 4 oral bacteriostats) tested by bioassays, 16 were toxic to fish and 6 were not. Tests were in 24- and 48-hour static bioassays on rainbow, brown, brook, and lake trout and bluegills at 12°C, and channel catfish at 17°C. The 16 toxic chemicals, in descending order, were malachite green, Trolene, CoRa1, Tiguvon, Roccal, P.M.A., Acriflavine, amopyroquin dihydrochloride, merthiolate, methylene blue, Neguvon, Ruelene, TV-1096, nickel sulfate, formalin, and quinacrine hydrochloride; the 6 that did not appear to be toxic were erythromycin thiocyanate, quinine hydrochloride, Flagyl, sulfamerazine, sulfamethazine, and sulfisoxazole.

An objective of the Fish Control Laboratories is to develop chemical tools to prevent and control fish diseases. Although efficacious concentrations of many drugs have been determined, a thorough examination of their toxicity has not been reported. Prior to clearance of drugs, the Food and Drug Administration requires data on their toxicity. The purpose of this study was to define the toxicity of 22 therapeutic chemicals to six species of fish before further research is undertaken on their efficacy and residues.

Eighteen parasiticides of known or possible value as external treatments for fish were selected for investigation upon recommendations by other investigators. Four oral bacteriostats were tested to determine whether any toxicity to fish would result through leaching, or excretion, of the compounds into water.

MATERIALS AND METHODS

Six species of fish were obtained from various fish hatcheries (table 1). All were quarantined for 10 days, and those judged acceptable for bioassays were acclimated to conditions of the tests.

The static bioassays were made in facilities described by Lennon and Walker (1964). We used 5-gallon glass jars which contained 15 liters of reconstituted, deionized water at total hardness of 42 p.p.m., and a maximum of 1 gram of fish per liter of water. Each test included 10 concentrations of a chemical. Ten fish were exposed to each concentration, and 20 fish served as controls.

The 22 therapeutic compounds were tested at $12\,^{\rm O}$ C. against five species of fish at La Crosse, Wis. (table 2). Tests against channel catfish at $17\,^{\rm O}$ were made at the Southeastern Fish Control Laboratory, Warm Springs, Ga.

A concentrated stock solution of each compound, using acetone or deionized water or both as solvents, was usually prepared for addition to the test vessels immediately before each test. When solubility of the compound prevented preparation of concentrated stocks, the compound was added directly and allowed to dissolve in the test vessel.

Observations on survival and mortality were recorded at 24 and 48 hours. The data were then analyzed by plotting concentration against mortality on logarithmic normal

Species	Lot	Average length (inches)	Average weight (grams)	Grading date	Source
Rainbow trout, Salmo gairdneri Do Brown trout, Salmo trutta Do Brook trout, Salvelinus fontinallis Do Lake trout, Salvelinus namaycush Do Channel catfish, Ictalurus punctatus Do Bluegill, Lepomis macrochirus Do Do Do	159 159 177 177 161 161 78 78 78 W-70 W-74 115 131	1.5 1.8 1.7 1.9 1.5 1.6 4.0 4.1 2.1 2.1 2.2 1.6 1.4	0.5 0.9 0.8 1.2 0.4 0.6 2.5 2.8 3.2 1.2 1.5 0.8 0.7	1-21-65 2-15-65 3-16-65 4- 1-65 1-21-65 2-15-65 8-14-64 8-28-64 10- 7-64 7-21-65 8- 4-65 11- 5-64 11-17-64 12- 1-64	National Fish Hatchery, Manchester, Iowa National Fish Hatchery, Lake Mills, Wis. State Fish Hatchery, St. Croix Falls, Wis. National Fish Hatchery, Jordan River, Mich. National Fish Hatchery, Marion, Ala. National Fish Hatchery, Lake Mills, Wis.

TABLE 2. -- Common names and active ingredients of compounds tested

Wildlife Component Component Component Season					
Common name	Grade or formulation	Active ingredient			
Acriflavine (neutral)	technical	3,6-diamino-10-methyl acridinium chloride and 3,6-diaminoacridine 4-(7-chloro-4-quinolylamino)-a-l-pyrrolidyl-o-cresol dihydrochloride			
CoRal Erythromycin thiocyanate	technical	0,0-diethyl 0-3-chloro-4-methyl-2-oxo-2H-1-benzopyran-7-yl-phosphorothioate erythromycin thiocyanate			
Flagyl	technical.	1-(2-hydroxyethy1)-2-methy1-5-nitroimidazole 37-percent formaldehyde gas in water			
Malachite green	technical	bis-(p-dimethylaminophenyl) phenylmethane treated with HCL sodium cthylmercurithiosalicylate			
Methylene blue	technical 50-percent soluble powder.	3,7-bis(dimethylamino) phenazathionium chloride			
Nickel sulfete	analytical reagent.	0,0-dimethyl 2,2,2-trichloro-1-hydroxyethyl phosphonate MISO. 6M,0 pyridylmercuric scetate			
Quinacrine hydrochloride (Atabrine)	technical	3-chloro-7-methoxy-9- (1-methyl-4-diethylaminobutylamino) acridine dihydrochloride			
Quinine hydrochloride	technical	quinine hydrochloride			
Roccal	50-percent concentrate	alkyl dimethylbenzylammonium chlorides			
Ruelene	227 mg/cc	4-tert-butyl-2-chlorophenyl methyl methylphosphoramidate			
Sulfamerazine. Sulfamethazine	U.S.P. U.S.P.	\mathcal{N}^{-} (4-methyl-2-pyrimidyl) sulfanilamide \mathcal{N}^{2} -(4,6-dimethyl-2-pyrimidinyl) sulfanilamids			
Sulfisoxazole Tiguvon	U.S.F. 300 mg/cc	N ² -(3,4-dimethyl-5-isoxazolyl) sulfanilamide 0,0-dimethyl 0-[4-(methylthio)-m-tolyl] phosphorothioate			
Trolene	technicai	0,0-dimethyl 0-2,4,5-trichlorophenyl phosphorothicate			
TV-1096 (Parke, Davis & Company)	technical	Lg-threo-2-(5-nitro-2-furyl)-5-(p-nitrophenyl)-2-oxazoline-4-methanol			

(probability) graph paper to define the concentration that produced 50-percent mortality (LC 50) as described by Litchfield and Wilcoxon (1949). Variance and the 95-percent confidence interval (C.I.) were also determined.

Most of the compounds tested were technical or U.S.P. materials, and the rest were formulated materials. To eliminate confusion, all results are reported in terms of p.p.m. of

total material (formulated or technical) instead of active ingredient.

RESULTS

Of the 22 compounds, 16 were toxic to the six species of fish, and the LC $_{50}$ values were determined (tables 3 to 8).

The most toxic compound, malachite green, is relatively uniform in toxicity to the six

TABLE 3.--Toxicity of 16 compounds to rainbow trout at $12^{\rm O}$ C.

TABLE 6.--Toxicity of 16 compounds to lake trout at 12° C.

	At	24 hours	At 48 hours	
Compound	LC ₅₀ (p.p.m.)	95-percent C.I.	(p.p.m.)	95-percent C.I.
Acriflavine	30.1	26.2-34.6	19.9	17.0-23.3
	47.0	43.5-50.8	35.3	33.3-37.4
Coffal	2.60	2.28-2.96	0.55	0.51-0.59
Formalin	207	182-236	168	154-183
Malachite green	0.50	0.36-0.69	0.39	0.33-0.46
Merthiolate	60.5	53.5-68.4	21.2	18.6-24.2
Methylene blue	25.0	20.5-30.5	16.0	13.8-18.7
	32.5	29.3-36.1	12.2	10.6-14.0
Nickel sulfate	320	302~339	160	150-171
P.M.A	5.00	4.35~5.75	3.75	3.02-4.65
Quinacrine HCL	3.24	2.92-3.60	172 2.57	159-1 <i>8</i> 6 2.16-3.06
Ruelene	35.0	31.5-38.8	32.0	30.5-33.6
Liguvon	5.30	4.82-5.83	4.35	3.62-5.22
TroleneTV-1096	1.17 24.2	0.89-1.53 22.8-25.7	0.74 16.1	0.64-0.86

	At 24	hours	At 48 hours		
Compound	LC ₅₀	95-percent	LC ₅₀	95-percent	
	(p.p.m.)	C.I.	(p.p.m.)	C.I.	
Acriflavine	37.5	34.7-40.5	28.0	24.4-32.2	
	15.5	12.1-19.8	14.0	10.8-18.1	
Collal	6.80	4.GO-11.56	4.00	1.25-12.80	
	220	2GO-242	167	160-174	
Malachite green	0.57	0.49-0.66	0.40	0.33-0.49	
Marthiolate	13.0	9.6-17.6	2.13	1.06-4.26	
Mathylane blue	35.0	29.4-41.6	34.0	29.3-39.4	
	41.0	38.7-46.5	9.00	7.20-11.25	
Nickel sulfate	170	139-209	75.0	55.6-101.2	
	12.5	11.8-13,2	7.60	6.33-9.12	
Quinacrine HCL	28.0	18.8-42.0	21.0	12.4-35.7	
	2.70	2.41-3.02	1.95	1.68-2.26	
RueleneTiguvon	27.0	25.0-29.2	27.0	23.9-30.5	
	6.50	6.08-6.96	5.30	4.91-5.72	
TroleneTV-1096	0.73	0.62-0.86	0.62	0.53-0.72	
	32.0	28.6-35.8	16.5	13.2-20.6	

TABLE 4.--Toxicity of 16 compounds to brown trout at $12^{\rm O}$ C.

TABLE 7.-- Toxicity of 16 compounds to channel catfish at 17° C.

NI CONTRACTOR OF THE CONTRACTO	At 24 hours		At 48	hours
Compound	LC50	95-percent	LG50	95-percent
	(p.p.m.)	C.I.	(p.p.m.)	C.I.
Acriflavine	40.0	36.4-44.0	27.0	25.0-29.2
	42.0	37.5-47.0	36.0	33.3-38.9
CoRalFormelin	0,92	0.84-1.00	0.73	0.62-0.86
	325	304-348	185	165-208
Malachite green	0.45	0.42-0.49	0.34	0.30-0.38
Merthiolate	11.0	75-160	54.0	47.8-61.0
Mathylene blue	54.0	46.2-63.2	32.8	28.8-37.4
Meguvon	54.0	48.2-60.5	16.5	11.8-23.1
Mickel sylfate	400	345-464	270	241-302
P.M.A	9,30	6.30-10.42	6.22	5.71-6.78
Quinacrine HCL	390	361-421	230	1.84-288
	2.95	2.46-3.54	2.05	1.74-2.42
Ruelene	26.2	24.7-27.8	25.7	24.2-27.2
Tiguvon	4.50	4.09-4.95	3.62	2.78-4.71
TrolenaTV-1096	0.53 	0.38-0.74	0.39	0.30-0.51

	At 24	hours	At 48 hours		
Сопроила.	LC ₅₀	95-percent C.I.	LC50 (p.p.m.)	95-percent C.I.	
Acriflavina	43.5	39.9-47.4	33.2	31.0-35.5	
	19.8	17.7-22.2	12.5	11.8-13.2	
CoRalFormalin	6.60 137	5.81-7.96 129-145	96.0	90.6-101.8	
Malachite green	0.21	0.17-0.27	0.20	0.16-0.26	
	7.50	6.41-8.78	5.65	4.79-6.67	
Methylene blue	120	110-131	104	93-116	
	80.0	72.7-88.0	32.0	24.8-41.3	
Nickel sulfate	368	334~405	165	129-211	
P.M.A	3.22	2.66~3.90	2.89	2.60-3.21	
Quinacrine HGL	198	169-232	70.0	59.3-82.6	
	1.28	1.16-1.41	1.12	1.03-1.22	
RueleneTiguvon	39.5	37.6-41.5	34.8	32.5-37.2	
	5.90	4.50-7.73	5.90	4.50-7.73	
Trolene	1.76	1.54-2.01	1.26	1.09-1.46	
	27.0	24.8-29.4	20.3	19.3-21.3	

TABLE 5.---Toxicity of 16 compounds to brook trout at 120 C.

TABLE 8.--Toxicity of 16 compounds to bluegills at 12° C.

	At 24	hours	At 48 hours	
Compound	LC50	95-percent	LC50	95-percent
	(p.p.m.)	C.I.	(p.p.m.)	C.I.
Acriflavine	48.0	43.2-53.3	14.6	14.0-15.7
	52.0	44.8-60.3	40.0	38.1-42.0
CoRal	1.06	0.87-1.29	0.50	0.70-0.91
	196	187-206	157	143-173
Malachite green	0.30	0.22-0.40	0.26	0.22-0.31
Merthiolate	89.5	85.2-94.0	74.5	71.0-78.2
Methylene blue	49.8	41.2-60.3	22.9	17.2~30.5
	34.0	23.4-49.3	16.8	14.1~20.0
Nickel sulfate	15.5	12.9-18.6	242 10.7	224-261 9.8-11.7
Quinacrine HCL	4.10	3.79-4.50	230 3.40	177-299 3.09-3.74
Ruelene.	36.8	34.4-39.4	35.0	31.5-38.8
Tiguvon.	6.15	5.21-7.26	5.50	5.14-5.88
TroleneTV-1096	0.59	0.44-0.78	0.39	0.26-0.59
	29.3	26.4-32.5	19.0	16.8-21.5

	At 24	hours	At 48 hours		
Compound	LC ₅₀ (p.p.m.)	95-percent C.I.	LC ₅₀ (p.p.m.)	95-percent C.I.	
Acriflavine	18.0	16.5-19.3	13.5	12.6-14.4	
	33.0	23.6-42.2	18.5	16.7-20.5	
CoftalPormalin	10.5	8.1-13.6	8.00	6.11-10.48	
	185	156-220	140	127-154	
Malachite green	0.26	0.22-0.31	0.11	0.09-0.14	
Merthiolate	110	87-139	64.5	57.6-72.2	
Wethylene blue	51.0	40.2-64.8	33.0	26.2-41.6	
	78.0	64.5-94.4	71.0	55.9-90.2	
Nickel sulfate	20.0	18.0-22.2	495 16.0	450-544 13.4-19.0	
Quinacrine HCL	120	73-1.98	79.0	54.1-115.3	
	2.10	1.94-2.27	1.68	1.56-1.81	
Ruelene	36.0	34.3-37.8	35.0	33.0-37.1	
	15.7	13.2-18.7	8.90	7.67-10.32	
Trolene	2.50	2.25-2.78	1.00	0.67-1.50	
	37.0	33.3-41.1	28.2	25.9-30.7	

range.

Nickel sulfate, formalin, and quinacrine hy-

drochloride are the least toxic of the com-

pounds analyzed. Formalin exhibits a fairly

hours. The other two have a much wider

Clemens and Sneed (1958a) reported the

LC₅₀ values of formalin on channel catfish to be 87 and 69 p.p.m. in 24 and 48 hours, re-

spectively, at 250 whereas we found them to

be 137 and 96 p.p.m. in 24 and 48 hours, re-

spectively, at 17°C. This variation in results

seems to indicate that the toxicity of formalin

may be increased by an increase in tempera-

ture. The observation is supported by our re-

sults which show that formalin is more toxic

Erythromycin thiocyanate and quinine hy-

drochloride were tested at an arbitrary level

of 100 p.p.m. Their solubility would have per-

mitted higher concentrations but preliminary

The poor solubility of Flagyl, sulfamera-

zine, sulfamethazine, and sulfisoxazole pre-

Solutions were saturated before lethal levels

could be reached. The arbitrary concentration

of 100 p.p.m. was selected then for tests. This

vented the determination of LC 50 values.

resulted in saturated solutions with excess

chemical remaining on the bottom of the bio-

assay vessels. None of them was toxic to the

DISCUSSION

Woodbury, 1936). Recently, Amlacher (1961)

six species of fish.

long-term treatments.

tests indicated little toxicity. At 100 p.p.m.,

the substances were not toxic to the fish.

to channel catfish at 170 than it is to four

species of trout and to bluegills at 12°.

uniform LC 50 range of 96 to 185 p.p.m. at 48

species, and LC $_{50}$ values range from 0.11 to

0.40 p.p.m. at 48 hours. Clemens and Sneed

(1958a) reported its LC 50 to channel catfish

Our results show the LC 50 values to be 0.21

as 0.14 p.p.m. in 24 and 48 hours at 25 $^{\circ}$ C.

and 0.20 p.p.m. in 24 and 48 hours respec-

may be due to differences in test tempera-

tively at 17°. This variation between results

Following malachite green in decreasing

Tiguvon, all of which have the basic structure

range of toxicity are Roccal and P.M.A., with

Roccal the more toxic of the two. Roccal, like

malachite green, exhibits relatively uniform

toxicity, and LC 50 values range from 1.12 to

P.M.A. exhibits a much wider range of tox-

icity with LC so values of 2.9 to 16.0 p.p.m. at

its LC50 to channel catfish as 3.8 p.p.m. in

authors (1958b) reported the LC 50 of P.M.A.

to channel catfish as 2.96 and 2.81 p.p.m. in 24 and 48 hours, respectively, at 16.5°. Both

values of 3.22 and 2.89 p.p.m. for 24 and 48

merthiolate, methylene blue, Neguvon, Rue-

lene, and TV-1096 fall into an intermediate

toxicity range with LC $_{50s}$ of 10 to 100 p.p.m.

Only Ruelene exhibits a uniform LC₅₀ range

TV-1096 is soluble only to approximately

30 p.p.m. in water. Amounts above this level

produce a saturated solution with a precipi-

tate on the bottom of the test vessel. Brown

30 p.p.m. and for this reason, LC50 values

could not be derived for the species. Also,

amounts of TV-1096 in excess of a saturated

solution are nontoxic to brown trout. In con-

trast, LC₅₀ values of TV-1096 for the other

species range from 16.1 to 28.2 p.p.m. at 48

trout fail to succumb to concentrations below

of 25.7 to 35.0 p.p.m. for six species in 48

hours. The other compounds of this group demonstrate a relatively wide range of tox-

icity to the different species.

Acriflavine, amopyroquin dihydrochloride.

reports compare favorably with our LC50

48 hours. Clemens and Sneed (1958a) reported

24 hours at 24 °C. In a later publication, these

order of toxicity are Trolene, CoRal, and

of phosphorothioate. In the same general

3.40 p.p.m. at 48 hours for all species.

tures.

hours at 170.

hours.

sic

recommended it for prolonged treatment of fish in ponds to combat Ichthyophthirius, Chilodenella, and Costia. He applied 0.15

p.p.m. and allowed it to dissipate in the water. Concentrations of 0.11 to 0.40 p.p.m. in our

bioassays proved toxic within 48 hours to the

Malachite green has been in use for many years as a fungicidal dip for fish (Foster and

six species of fish tested. Thus, there is a risk with concentrations over 0.11 p.p.m. in

Trolene, CoRal, and Tiguvon are under consideration as prolonged treatments for control of Ichthyophthirius. Tiguvon is the least toxic of these organophosphates to fish. This indicates that it may prove the most valuable of the group if minimum concentrations required for control of "Ich" are approximately the same for all three.

Roccal has been in use as a bactericide for many years (Fish, 1947). Putz (1964) reported its possible value in prolonged, indefinite treatments at 0.25 to 0.50 p.p.m. for Ichthyophthirius. In treatments such as this, the chemical attacks the free-living stages of "Ich". He did not say which formulation of the chemical he used, but 10-percent active is the formulation commonly used in hatcheries (Davis, 1956). We used 50-percent active, and upon converting from 10-percent active to 50-percent active, the treatment levels could be reduced to 0.05 and 0.10 p.p.m. This permits a comparison between treatment levels and toxicity which shows a 10-fold difference in concentrations.

P.M.A. has been of considerable value in combating bacterial and protozoan diseases (Davis, 1956). Evidence of its greater toxicity to rainbow trout than other trouts has been reported over the years (Foster and Olson, 1951; Rodgers et al., 1951; Wolf, 1951; Hammer, 1960). Allison (1957) reported variations in the toxicity of P.M.A. from lot to lot of chemical. We used only one lot of P.M.A. in this study, and the results support the earlier findings that it has greater toxicity to rainbow trout. For example, it was up to three times as toxic to rainbow trout as to brook trout. Channel catfish appear to be sensitive to the compound at 17°C.

Snieszko and Friddle (1948) used merthiolate (sulfo) as a disinfectant for rainbow trout eggs. Van Duijn (1956) cautioned against use of merthiolate as a fish bath since the compound is a mercurial and is certain to be toxic to fish in contact with it for some period. We find an extreme variation in its toxicity to different species. This is especially true at 24 hours where LC $_{50}$ values range from 7.5 p.p.m. for channel catfish to 110.0 p.p.m. for brown trout. This variation diminishes some-

what at 48 hours, and lake trout become the most sensitive to the chemical. The LC $_{50}$ at 48 hours for lake trout is 2.1 p.p.m. in contrast with 74.5 p.p.m. for brown trout. Variations in resistance such as this may make merthical extremely difficult to work with in routine treatments of several species.

Acriflavine, amopyroquin dihydrochloride, methylene blue, Neguvon, Ruelene, and TV-1096 are under consideration as prolonged, indefinite treatments for control of Ichthyophthirius. Putz (1964) reported that 3 p.p.m. of acriflavine shows promise against the parasite. Our results indicate that bluegills are the most sensitive to the compound with LC 50 values of 18.0 and 13.5 p.p.m. at 24 and 48 hours respectively. Channel catfish are the most resistant with LC 50 values of 43.5 and 33.2 p.p.m. at 24 and 48 hours.

Clemens and Sneed (1958a) reported the LC_{50} values of acriflavine on channel catfish at 24 and 48 hours to be 11.5 and 6.8 p.p.m., respectively, at 20 °C. Our finding, is that it is only about one-fourth as toxic as that. Possible causes for the discrepancy are many. Among them are differences in water quality and temperature, differences in the condition of fish, and purity of the compound used. In addition to this unexplained variation in the toxicity of acriflavine, another factor warrants serious consideration in its use. Van Duijn (1956) reported sterility in both egglaying and live-bearing aquarium fish. This is a temporary situation and normal fertility is restored after several months.

Amopyroquin dihydrochloride also shows promise as a prolonged, indefinite treatment at 0.05 to 0.10 p.p.m. for control of Ichthyophthirius (Putz, 1964). Our results show that its toxicity to all trout tested, with the exception of lake trout, is between 35 and 40 p.p.m. for 48 hours. Also, bluegills at 12°C. and channel catfish at 17°0 are approximately as sensitive as lake trout. A treatment level of 0.1 p.p.m. would include a safety margin in use of more than a hundredfold even against these three species.

Van Duijn (1956) recommended methylene blue as a satisfactory control for Ichthyophthirius in aquariums. He used 2 to 4 p.p.m.

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of it in a permanent bath at temperatures between 21 and 27°C. We found that rainbow trout are the most sensitive to the dye, and the LC $_{50}$ is 16 p.p.m. at 48 hours. The most resistant species is channel catfish with an LC $_{50}$ of 104 p.p.m. at 48 hours. The remaining species are intermediate in sensitivity with a 48-hour LC $_{50}$ range of 22.9 to 34.0 p.p.m. Comparison of the use levels with toxicity levels indicates a good safety margin.

Neguvon and Ruelene are of approximately the same toxicity except in one very important respect. Neguvon has a marked difference between the 24-hour and 48-hour LC $_{50}$. The most striking example of this involves lake trout with LC $_{50}$ values of 41 p.p.m. at 24 hours and 9 p.p.m. at 48 hours. Differences between the 24- and 48-hour LC $_{50}$ values by a factor of at least two are common except with bluegills. For some unknown reason the difference with bluegills is only 78 to 71 p.p.m.

Ruelene provides a contrast with Neguvon because it exhibits approximately the same toxicity at 24 and 48 hours. In the case of lake trout, the 24- and 48-hour LC $_{50}$ values are identical at 27 p.p.m. It is possible that Ruelene degrades very rapidly in the test vessel to a nontoxic level.

TV-1096 has toxicity comparable to that of Neguvon and Ruelene. Like Neguvon, it does not appear to degrade as rapidly as Ruelene.

Nickel sulfate is under consideration as a prolonged, indefinite treatment for control of Ichthyophthirius. Our results show that it is relatively low in toxicity when compared with the other compounds tested, but it has a fairly wide range of toxicity among the species tested. The LC 50 values at 48 hours range from 75 p.p.m. for lake trout to 495 p.p.m. for bluegills. Twenty-four-hour tests of 50 to 275 p.p.m. on brook trout and 200 to 500 p.p.m. on bluegills did not cause death.

Allison (1957) reported use of formalin as a parasiticide in long-term treatments in ponds. He suggested 5 p.p.m. for Gyrodactylus and 15 p.p.m. for Trichodina and Ichthyophthirius. Our results show that formalin is relatively and uniformly low in toxicity when compared

with the other compounds tested. It does appear to increase in toxicity as temperatures rise from 17° to 25° C. Even with this increase in toxicity, the compound retains a safety margin of at least sixfold at recommended use levels.

Van Duijn (1956) recommended use of quinacrine hydrochloride in treatment of stubborn cases of "Ich" in aquarium fish. The treatment consists of three applications of 1 p.p.m. at 48-hour intervals. This totals 3 p.p.m. if no degradation of the compound occurs. He also stated that this treatment should not be extended over long periods and that 8 to 10 days should be sufficient.

Our results show that lake trout are approximately 10 times as sensitive to quinacrine hydrochloride as the other trout and 3 or 4 times as sensitive as bluegills and channel catfish. The sensitivity is complicated by the fact that the toxicity to lake trout is quite erratic and some deaths occur over a wide range of concentrations. The 48-hour LC, of quinacrine hydrochloride for lake trout is approximately 10 p.p.m., the LC $_{\rm 50}$ is 21 p.p.m., and the LC $_{\rm 100}$ $\,$ is 110 p.p.m. Some fish succumb to the chemical quickly and at comparatively low concentrations whereas the rest survive for long periods. Further evidence of this lingering is shown by the slight difference between the 24-hour LC 50 of 28 p.p.m. and the 48-hour LC₅₀ of 21 p.p.m. In contrast, there is a considerable difference between the 24- and 48-hour LC 50 values obtained for the other species. A possible explanation is that there is considerable variation in resistance among lake trout individuals.

Van Duijn (1956) recommended use of quinine hydrochloride for treatment of Ichthyophthirius in aquarium fish. The treatment consists in adding 1 p.p.m. on 3 successive days, a final treatment level of 3 p.p.m. He cautioned against use of the treatment for long periods because of possible fertility problems. Our results show that 100 p.p.m. of the chemical in water are not toxic within 48 hours to the species tested.

Erythromycin thiocyanate has been used as a food additive for control of kidney disease in rainbow trout at 4.5 grams per 100 pounds of fish per day for 21 days (Piper, 1961). Warren (1963) reported that it is toxic to rainbow trout at 500 mg. per kg. Our results show that 100 p.p.m. of the antibiotic in water is not toxic to the species tested within 48 hours.

Flagyl has been used in medicine as an antiprotozoal agent (Cutting, 1962). Putz (1964) reported experimental use of it at 1.5 p.p.m. for control of Ichthyophthirius. Our results show that Flagyl is nontoxic at 100 p.p.m. The finding is qualified somewhat since the compound is not immediately soluble at 100 p.p.m. It dissolves slowly, however, and is completely in solution within 48 hours.

Snieszko and Bullock (1957) reported use of sulfamerazine, sulfamethazine, and sulfisoxazole as food additives in the treatment of furunculosis at 8 to 10 grams per 100 pounds of fish per day for 10 to 20 days. Van Duijn (1956) recommended use of the sodium salt of sulfamerazine at 1 part per thousand as an effective cure for worm cataract in aquarium fish. In our water, sulfamerazine, sulfamethazine, and sulfisoxazole are not soluble at 100 p.p.m., and saturated solutions are not toxic to the six species within 48 hours.

All of our results were obtained with fish which were, to the best of our knowledge, healthy. They showed no signs of disease or physical injuries. The toxicity of these compounds to fish which are sick or in poor condition might be significantly different.

None of the compounds reported herein are cleared by the Food and Drug Administration and the Department of Agriculture for use on fish destined for human consumption. The data and discussion presented in this paper should not be construed as recommendations for use.

CONCLUSIONS AND SUMMARY

The toxicities of 22 therapeutic compounds to rainbow trout, brown trout, brook trout, lake trout, and bluegills at $12^{\,0}$ C. and channel catfish at $17^{\,0}$ were determined in 24- and 48-hour static bioassays.

LC $_{50}$ values for malachite green, the most toxic compound tested, range from 0.1 to 0.4 p.p.m. for all species tested. CoRal, P.M.A., Roccal, Tiguvon, and Trolene are less toxic than malachite green, but still rank relatively high in toxicity. Their LC $_{50}$ values range from approximately 0.5 to 10 p.p.m. for all species.

Acriflavine, amopyroquin dihydrochloride, merthiolate, methylene blue, Neguvon, Ruelene, and TV-1096 are intermediate in toxicity. The LC $_{50}$ values range from approximately 10 to 100 p.p.m. for all species. Merthiolate has wide variations in toxicity to various species.

Formalin, nickel sulfate, and quinacrine hydrochloride have relatively low toxicities. The LC $_{50}$ values are usually above 100 p.p.m. Quinacrine hydrochloride is substantially more toxic to lake trout than to the other species.

No tests of erythromycin thiocyanate and quinine hydrochloride were made at concentrations above 100 p.p.m. They are not toxic within 48 hours at this concentration. Saturated solutions of Flagyl, sulfamerazine, sulfamethazine, and sulfisoxazole are also not toxic within 48 hours.

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