

Annual Operating Plan

for

Lower Snake River Fish and Wildlife Compensation Programs

Grande Ronde and Imnaha Basins, Oregon

Spring Chinook Salmon

Fall Chinook Salmon

Coho Salmon

Pacific Lamprey

For the Period of

January 1 – December 31, 2017

Prepared by:

Oregon Department of Fish and Wildlife

Confederated Tribes of the Umatilla Indian Reservation

Nez Perce Tribe

For

Lower Snake River Compensation Plan

USFWS Administration

and

Bonneville Power Administration

Final

3/17/2017

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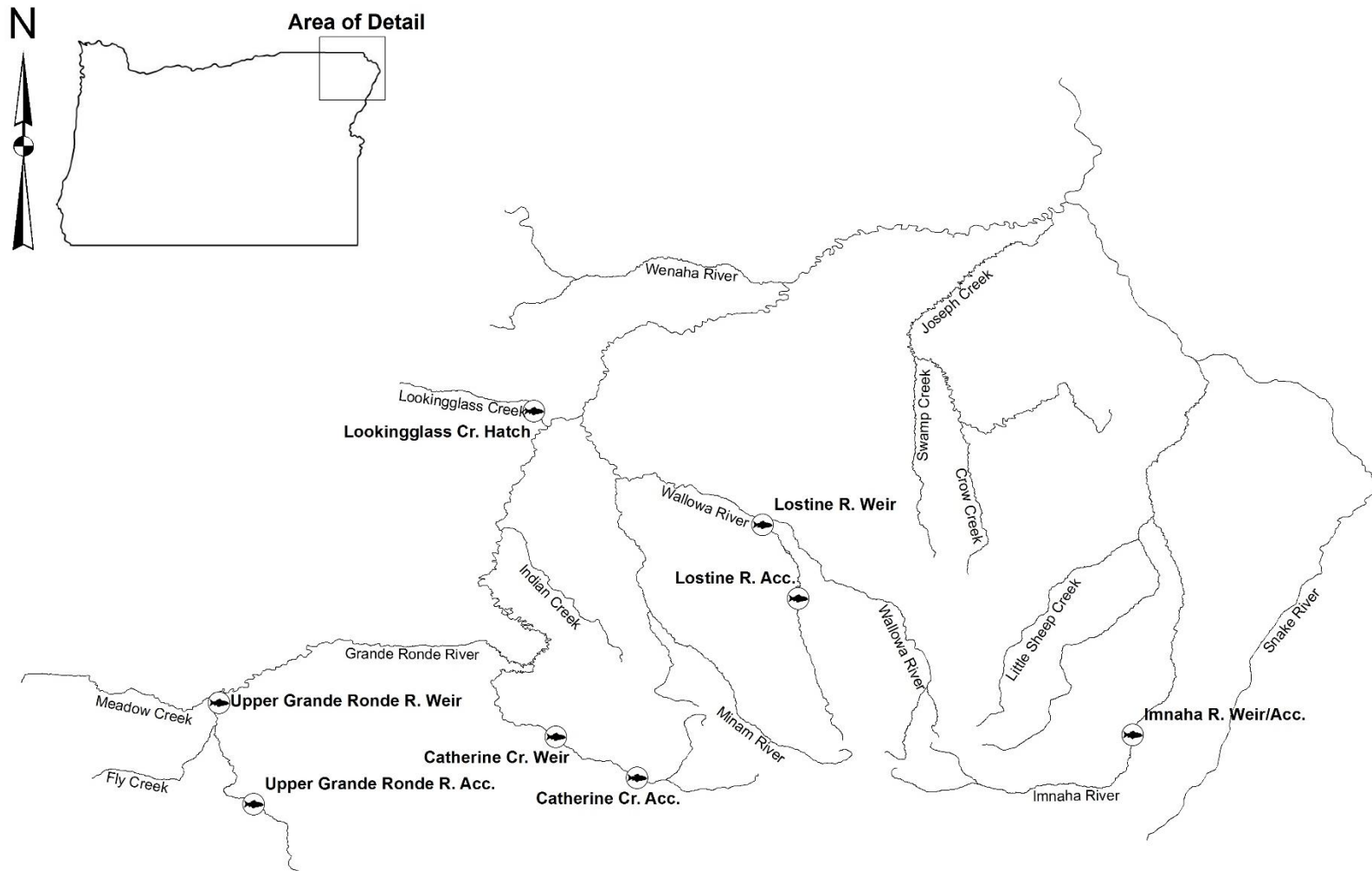


Figure 1. Grande Ronde and Imnaha basin Chinook salmon hatchery facilities and release locations.

Table 1. Grande Ronde and Imnaha basin Chinook salmon smolt release goals, including marking strategy. Source = funding source for allocation of PIT tags: LSRCP (Lower Snake River Compensation Plan), CSS (Comparative Survival Study – Fish Passage Center), and BPA (Bonneville Power Administration).

Stock	Smolt release goal	Acclimation Period		Direct Release	Ad clipped (%)	CWT (N)	CWT (%)	PIT Tags	
		First	Second					N	Source
Upper Grande Ronde R.	250,000	125,000	125,000		50	250,000	100	5,000	LSRCP
Catherine Crk.	150,000	150,000			100	100,000	67	21,000	CSS
Lookingglass Crk.	250,000	250,000	0		100	126,000	50	5,000	LSRCP
Lostine R.	250,000	125,000	125,000		100	126,000	50	6,000	BPA
Grande Ronde Basin	900,000					602,000		37,000	
Imnaha R.	490,000	280,000		210,000	100	240,000	49	21,000	CSS
Oregon Snake Basin	1,390,000	930,000	250,000	210,000		842,000		58,000	

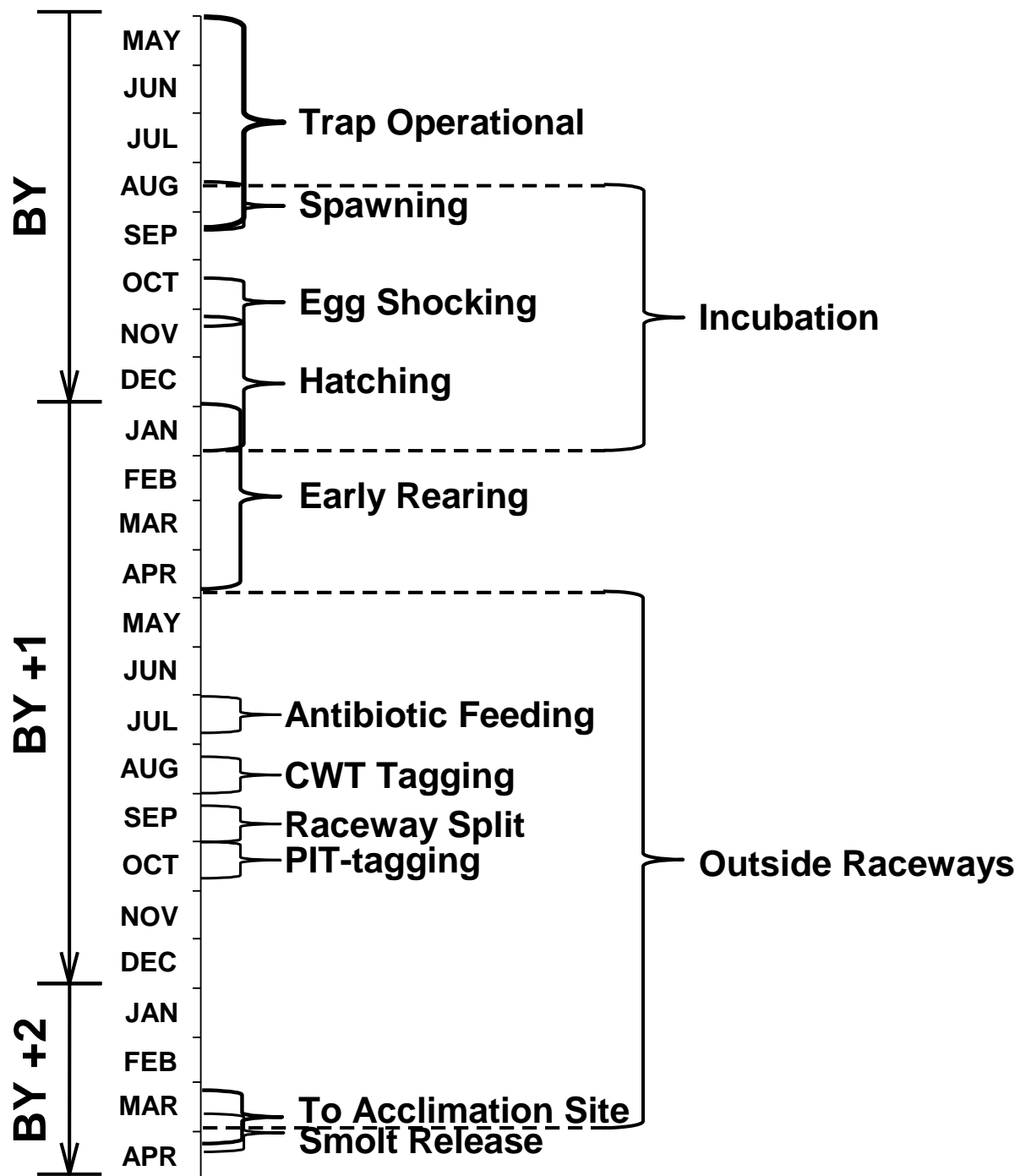


Figure 2. Grande Ronde and Imnaha basin Chinook salmon production timeline.

Table 2. Grande Ronde and Imnaha basin Chinook salmon brood year (BY) 2015 inventory, current as of 2/8/2017. Coded-wire tag (CWT) and Adipose fin clip (AD) are goals for each raceway, not actual successful marking/tagging.

Release Site	Pre-Split		Post-Split		% CWT	CWT Codes	PIT Tag (N)	AD (%)
	Container	N	Container	N				
Upper Grande Ronde R. (GQ)	RW15	62,500	RW15	65,000	100	091067	497	0
	RW18	62,500	RW18	64,500	100	091070	501	100
	RW16	62,500	RW16	63,000	100	091068	483	0
	RW17	62,500	RW17	63,000	100	091069	498	100
Upper Grande Ronde Total		250,000		255,000			1,979	
Catherine Crk. (CQ)	RW2	75,000	RW2	76,500	74	091059	10,470	100
	RW3	75,000	RW3	78,500	70	091060	10,455	100
Catherine Crk. Total		150,000		155,000			20,925	
Lookingglass Crk. (LG)	RW1	70,000	RW1	76,000	59	091053	984	100
	RW10	90,000	AHP1	41,000	56	091054	1,000	100
	RW14	90,000	AHP2	41,000	56	091054	997	100
			AHP3	41,500	56	091055	994	100
			AHP4	41,000	56	091055	1,001	100
Lookingglass Crk. Total		250,000		240,500			4,976	
Lostine River (LQ)	RW11	83,300	RW11	66,500	72	091061	595	100
	RW13	83,300	RW13	66,500	24	091061	597	100
	RW12	83,300	RW12	67,000	75	091062	599	100
			RW14	67,000	25	091062	601	100
Lostine River Total		250,000		267,000			2,392	

Table 2 Continued. Grande Ronde and Imnaha basin Chinook salmon brood year (BY) 2015 inventory, current as of 2/8/2017. Coded-wire tag (CWT) and Adipose fin clip (AD) are goals for each raceway, not actual successful marking/tagging.

Release Site	Pre-Split		Post-Split		% CWT	CWT Codes	PIT Tag (N)	AD (%)
	Container	N	Container	N				
Imnaha River (IM)	RW4	81,600	RW4	70,000	81	091063	2,982	100
	RW5	81,600	RW5	69,500	82	091064	2,974	100
	RW8	81,600	RW8	69,500	0	–	2,962	100
	RW9	81,600	RW9	69,500	29	091063, 091064	2,979	100
	RW6	81,600	RW6	74,500	84	091065	2,982	100
	RW7	81,600	RW7	70,500	82	091066	2,990	100
				RW10	69,000	29	091065, 091066	2,987
Imnaha River Total		490,000		492,500			20,856	

Table 3. Grande Ronde and Imnaha basin Chinook salmon brood year (BY) 2015 transport and release schedule in 2017.

Stock	Transfer Date	Release Type	Container	To	N	Estimated Pounds	Release Date	Force out Date
Grande Ronde R.	15-Mar	1 st Acclimation	RW 15,18	UGR Acc.	129,395	5,175	17-Mar	4-Apr
Grande Ronde R.	5-Apr	2 nd Acclimation	RW 16,17	UGR Acc.	126,492	5,059	7-Apr	14-Apr
Grande Ronde Total					255,887			
Catherine Creek	16-Mar	Acclimated	RW 2,3	Catherine Acc.	155,750	6,230	20-Mar	14-Apr
Catherine Total					155,750			
Lookingglass Creek		Volitional	RW 1, A,B,C,D	Lookingglass Crk.	241,947	9,677	1-Apr	15-Apr
Lookingglass Total					241,947			
Lostine River	13-Mar	1 st Acclimation	RW 11,13	Lostine R. Acc.	133,698	5,347	21-Mar	29-Mar
Lostine River	30-Mar	2 nd Acclimation	RW 12,14	Lostine R. Acc.	134,197	5,367	12-Apr	22-Apr
Lostine Total					267,895			
Imnaha River	23-Mar	Acclimated	RW 4,5,8,9	Imnaha R. Acc.	285,010	11,400	3-Apr	13-Apr
Imnaha River	13-Apr	Direct	RW 6,7,10	Imnaha R.	208,876	8,355	13-Apr	N/A
Imnaha Total					493,886			

Table 4. Brood year (BY) 2016 fry inventory with release goals and marking plan, current as of 2/10/2017.

Stock	Stock-BY ID	Estimated Fry	Release Goal	% Ad Clip	% CWT	PIT Tag (N)
Upper Grande Ronde	GQ 80-16	229,869	250,000	50	100	5,000
Catherine Creek	CQ 201-16	127,601	150,000	100	72	21,000
Lookingglass Creek	LG 81-16	233,711	250,000	100	57	5,000
Lostine River	LQ 200-16	262,588	250,000	100	50	6,000
Imnaha River	IM 29-16	495,793	490,000	100	55	21,000
Total Fry to Pond		1,349,562	1,390,000			58,000

Table 5. Pre-season return estimates for adult (age 4-5) Chinook salmon returning to Oregon Snake Basin tributaries in 2017.

Stock	Origin	BON	Conversion to LGD	LGD	Conversion to Tributary	Tributary Mouth	Conversion from BON to Tributary
Upper Grande Ronde	Natural	212	85%	180	90%	162	77%
	Hatchery	912	70%	639	90%	575	63%
Catherine Creek	Natural	214	85%	182	90%	164	77%
	Hatchery	305	70%	214	90%	193	63%
Lookingglass Creek	Natural	146	85%	124	90%	111	77%
	Hatchery	1,016	70%	711	90%	640	63%
Lostine River	Natural	315	85%	267	90%	240	77%
	Hatchery	930	75%	698	90%	629	68%
Grande Ronde Total	Natural	887		753		677	
	Hatchery	3,163		2,262		2,037	
Imnaha River	Natural	550	85%	467	90%	421	77%
	Hatchery	1,232	75%	925	90%	833	68%
Oregon Snake Basin	Natural	1,437		1,220		1,098	
	Hatchery	4,395		3,187		2,870	

Table 6. Grande Ronde and Imnaha basin Chinook salmon brood year (BY) 2017 brood stock collection goals.

Stock	Goals			Fecundity Estimate (eggs per female)	Natural-origin in brood (%) ^a
	Female Collection	Females Spawned	Green Eggs Collected		
Upper Grande Ronde (GQ)	85	73	287,000	3,937	50
Catherine Creek (CQ)	49	44	177,100	4,025	50
Lookingglass Creek (LG)	85	76	288,040	3,790	30
Lostine River (LQ)	83	73	315,871	4,327	25
Grande Ronde Total	290	259	1,068,011		
Imnaha (IM)	141	134	607,154	4,531	25
Oregon Snake Basin Tributaries	427	382	1,625,324		

^a Subject to change based on in-season run projections.

Table 7. BY 2017 Upper Grande Ronde River weekly brood stock collection strategy, by sex, during years when ‘normal’ water temperatures are observed and when ‘high’ temperatures require early removal of the weir and trapping facilities.

Week	Est. NOR Timing (%)	Females		Males		Jack	
		Normal Temp	High Temp	Normal Temp	High Temp	Normal Temp	High Temp
6-May	0.0	0	0	0	0	0	0
13-May	0.0	0	0	0	0	0	0
20-May	0.0	0	0	0	0	0	0
27-May	0.0	0	0	0	2	0	0
3-Jun	3.7	3	7	3	4	2	2
10-Jun	28.0	24	26	21	23	2	2
17-Jun	40.2	34	36	30	32	2	2
24-Jun	17.1	15	16	13	14	2	2
1-Jul	7.3	6	<i>Missed</i>	5	<i>Missed</i>	0	<i>Missed</i>
8-Jul	0.0	0		0		0	
15-Jul	2.4	2		2		0	
22-Jul	1.2	1		1		0	
29-Jul	0.0	0		0		0	
5-Aug	0.0						
Total Goal		85	85	75	75	8	8

Table 8. BY 2017 Catherine Creek weekly brood stock collection strategy, by sex, and origin (natural-origin, NOR, and hatchery-origin, HOR) for Chinook salmon.

Week	Est. NOR Timing (%)	Females		Males		Jack	
		NOR	HOR	NOR	HOR	NOR	HOR
6-May	0.0	0	0	0	0	0	0
13-May	0.8	0	0	0	0	0	0
20-May	1.6	0	0	0	0	0	0
27-May	6.5	0	0	0	0	0	0
3-Jun	14.6	1	2	1	2	0	0
10-Jun	14.6	4	5	3	5	0	0
17-Jun	27.6	3	5	3	5	0	0
24-Jun	17.9	5	9	4	8	1	2
1-Jul	8.1	3	6	3	5	0	1
8-Jul	4.1	1	2	1	2	0	0
15-Jul	2.4	1	1	1	1	0	0
22-Jul	0.8	0	1	0	1	0	0
29-Jul	0.8	0	0	0	0	0	0
5-Aug	0.0	0	0	0	0	0	0
Total Goal		18	31	16	29	1	3

Table 9. BY 2017 Lookingglass Creek weekly brood stock collection strategy, by sex and origin (natural-origin, NOR, and hatchery-origin, HOR) for Chinook salmon.

Week	Est. NOR Timing (%)	Females		Males		Jack	
		NOR	HOR	NOR	HOR	NOR	HOR
6-May	0.0	0	0	0	0	0	0
13-May	0.0	0	0	0	0	0	0
20-May	0.0	0	0	0	0	0	0
27-May	1.6	0	1	0	1	0	0
3-Jun	11.5	3	7	3	6	0	1
10-Jun	16.4	4	10	4	9	0	1
17-Jun	21.3	8	12	5	11	0	2
24-Jun	13.1	3	8	3	7	0	2
1-Jul	11.5	3	7	3	6	0	1
8-Jul	4.9	1	3	1	3	0	1
15-Jul	3.3	1	2	1	2	0	0
22-Jul	1.6	0	1	0	1	0	0
29-Jul	1.6	0	1	0	1	0	0
5-Aug	1.6	0	1	0	1	0	0
12-Aug	0.0	0	0	0	0	0	0
19-Aug	3.3	1	2	1	2	0	0
26-Aug	4.9	1	3	1	3	0	0
2-Sept	3.3	1	2	1	2	0	0
Total Goal		26	60	23	55	0	8

Table 10. BY 2017 Lostine River weekly adult brood stock collection strategy, by sex and origin (natural-origin, NOR, and hatchery-origin, HOR) for Chinook salmon.

Week	Est. NOR Timing (%)	Females		Males		Jacks	
		NOR	HOR	NOR	HOR	NOR	HOR
< 8-Jun	0.0	0	0	0	0	0	0
8-Jun	0.0	0	0	0	0	0	0
15-Jun	1.0	0	1	0	1	0	0
22-Jun	3.0	1	2	1	2	0	0
29-Jun	6.0	1	4	1	4	0	0
6-Jul	6.0	1	4	1	4	0	0
13-Jul	20.0	4	12	4	12	0	1
20-Jul	12.0	2	7	2	7	0	1
27-Jul	9.0	2	6	2	6	0	1
3-Aug	3.0	1	2	1	2	0	0
10-Aug	2.0	1	2	1	2	0	0
17-Aug	1.0	0	1	0	1	0	0
24-Aug	12.0	3	8	3	8	0	1
31-Aug	15.0	3	9	3	9	0	1
7-Sep	8.0	2	4	2	4	0	1
Total Goal		21	62	21	62	0	6

Table 11. BY 2017 Imnaha River weekly brood stock collection strategy, by sex and origin (natural-origin, NOR, and hatchery-origin, HOR) for Chinook salmon.

Week	Est. NOR Timing (%)	Females		Males		Jacks	
		NOR	HOR	NOR	HOR	NOR	HOR
< 16-Jun	4.0	1	4	1	4	0	0
17-Jun	12.0	4	13	4	13	1	1
24-Jun	28.0	10	29	10	29	2	2
1-Jul	24.0	10	25	10	25	2	2
8-Jul	12.0	4	13	4	13	1	1
15-Jul	10.0	4	11	4	11	0	0
22-Jul	4.0	1	4	1	4	0	0
29-Jul	3.0	1	3	1	3	0	0
5-Aug	0.0	0	0	0	0	0	0
12-Aug	3.0	1	3	1	3	0	0
19-Aug	0.0	0	0	0	0	0	0
Total Goal		36	105	36	105	6	6

Table 12. Grande Ronde and Snake River Fall Chinook salmon brood year 2016 (BY 16) inventory from Lyon’s Ferry Hatchery (LFH), current as of 2/21/17.

Release Site	Scheduled Release	Release Type	Stock	Expected at Release	CWT (%)	PIT (N)	AD (%)
Snake River	May 15,17,19	Dir.	LFH	200,000	100	1,500	100
		Dir.	LFH	900,000	0	1,500	100
Snake River Total				1,100,000		3,000	
Grande Ronde River	June 1	Dir.	LFH	200,000	100	1,500	100
		Dir.	LFH	240,000	0	1,500	0
Grande Ronde River Total				440,000		3,000	

Table 13. Grande Ronde basin Coho salmon brood year (BY) 2015 inventory, current as of 2/17/2017.

Release Site	Cascade Raceway	Release Type	N	Fish Size (fpp) ^a	Marks			Transfer Date
					CWT (%)	PIT (N)	AD (%) ^b	
Lostine River	24	Direct	72,755	20.5	0	–	0	March 9
	25	Direct	72,748	20.5	0	–	0	March 9
	26	Direct	72,556	20.5	0	–	0	March 9
	27	Direct	72,721	20.5	0	–	0	March 9
	28	Direct	72,717	20.5	0	–	0	March 9
	29	Direct	72,782	20.5	0	1,800	0	March 9
	30	Direct	66,395	20.5	100	3,182	100	March 9
Lostine River Total			502,674			4,982		

^a Fish are expected to be 19.0 fish/lb. by transfer on 3/9/17

^b Brood year 2015 was not ad-clipped due to expected shortages, future brood years will be 100% ad clipped.

Table 14. Tentative release numbers and locations of lamprey in the Grande Ronde River basin in 2017.

Program / Location	Life Stage	Estimated Release	Release Timing
Confederated Tribes of the Umatilla Indian Reservation (CTUIR)			
Upper Grande Ronde River	Adult	400	May-June 2017
Catherine Creek	Adult	150	Late May 2017
Lookingglass Creek	Adult	150	Late May 2017
CTUIR Total		700	
Nez Perce Tribe (NPT)			
Wallowa River	Adult	38	Late May 2017
Minam River	Adult	37	Late May 2017
Joseph Creek ^a	Adult	75	Late May 2017
NPT Total		150	

^a Lamprey will be released at two locations: Chesnimnus Creek on the Yost property (37 lamprey), and at the confluence of Chesnimnus and Devil's Run Creeks (38 lamprey)

Standard Operating Procedures

for

Lower Snake River Fish and Wildlife Compensation Programs

Grande Ronde and Imnaha Basins, Oregon

Spring Chinook Salmon

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Final

3/16/2017

1. Grande Ronde and Imnaha Spring/Summer Chinook

A. Production Goals – Consistent with production tables specified in the *U.S. vs. Oregon* fish management agreement, production objectives for each tributary is described below. All production is conventional. Release target size is 20-25 fpp.

- i. Upper Grande Ronde = 250,000
- ii. Catherine Creek = 150,000
- iii. Lookingglass Creek = 250,000
- iv. Lostine River = 250,000
- v. Imnaha River = 490,000

B. Smolt Releases

i. Key Contacts

1. Grande Ronde - Hatcheries: CTUIR (McLean), ODFW (Montgomery), NPT (Zollman). Fish Research: CTUIR (Naylor, Crump) ODFW (Hoffnagle, Feldhaus, Jonasson), NPT (Hesse, Vogel, Vatland, Young), and NOAA (Farman).
2. Imnaha - NPT (B. Johnson, Hesse, Olson, Vatland, Young, Vogel), ODFW (D. Eddy, Feldhaus, Hoffnagle, Yanke), CTUIR (McLean, Zimmerman), LSRCP (Collins, Starr, Engle, Robertson) and NOAA (Farman).

ii. Acclimation facilities - All facilities should be set-up and operational at least 2 days prior to scheduled delivery of smolts.

1. Release numbers = Last physical inventory – mortalities.
2. Operators report final numbers to the ODFW LGH staff or Shari Beals.
3. Scan all acclimation mortalities for PIT tags. PIT tag data provided to NPT (Vatland) and ODFW (Keniry). Mortalities should be provided to fish health for examination.

iii. Contingencies – Under extreme conditions, smolts may be released earlier than scheduled. In those cases, notify downstream rotary trap operators immediately.

1. Grande Ronde Traps - Brian Jonasson, Scott Favrot, Brad Garner, Pat Keniry, Mike McLean, Julie Collins, Les Naylor, Carrie Crump.
2. Imnaha Traps - Brian Jonasson, Julie Collins, Jim Harbeck, Jason Vogel, and Devin Olsen
3. Notify co-managers, LSRCP, and NOAA within 24 hours.

iv. Site-specific scheduling

1. Upper Grande Ronde River Acclimation
 - a. Volitional release begins within one week after delivery.
 - b. Smolts are forced out after 10 to 21 days of acclimation.
 - c. Fish are split equally into 4 containers.
 - d. Two acclimation phases – early and late (half of production each, half of CWT only group in each acclimation).

2. Catherine Creek Acclimation
 - a. Volitional release begins within one week after delivery.
 - b. Smolts are forced out after 25 days of acclimation (around mid-April).
 - c. Fish are split equally into 4 containers.
 - d. One acclimation phase.
3. Lookingglass Creek Direct Release
 - a. Screens are pulled, releasing fish, around the first of April.
 - b. Remaining fish are forced out mid-April.
4. Lostine River Acclimation
 - a. Smolts are volitionally released after 7-10 days of acclimation.
 - b. Fish are split evenly into 4 containers.
 - c. Two acclimation phases – early and late (half of production each).
5. Imnaha River Acclimation and Direct
 - a. Four raceways (280,000 smolts) are acclimated and released, and three raceways (210,000 smolts) are direct-stream released near the vicinity of the acclimation site.
 - b. Acclimate for seven days and screens will be pulled in early-April, remaining fish will be forced out mid-April.

C. Adult Trapping

- i. Key Contacts – Adult Trapping
 1. Transportation - Facility operators (NPT and CTUIR) coordinates all hauling and notifies LGH (Montgomery) of the stock, number being hauled, and estimated arrival time.
 2. Communications – Operators should distribute summaries of collections and operations to co-manager regularly (weekly or bi-monthly). Wallowa hatchery provides a summary of fish provided for distribution/food bank (Lostine and Imnaha stocks).
- ii. Grande Ronde Basin trapping guidelines
 1. While in operation, trapping facilities are checked daily.
 2. Captured fish will be collected for transport or released at the time they are first handled. If more than 15 adults are in the trap then fish will be collected and transported or passed upstream that day. If less than 15 fish are in the trap, fish may be held up to 72 hours.
 3. Monitor water temperatures, and adjust schedule to best coincide their work with the coolest water temperatures.
 4. Operators may conduct walking surveys, making visual observations of fish below weir sites to anticipate trapping numbers. Surveys may include snorkeling.
 5. Broodstock fish are injected with tulathromycin and oxytetracycline during processing.

iii. Imnaha trapping guidelines

1. The Imnaha satellite facility is staffed 24 hours per day, 7 days per week, while operational.
2. After the first Chinook is captured, trap is processed daily through the last week of July.
3. After July, trap is processed during weekdays. Exception is if 10 or more natural Chinook salmon, or 30 or more total fish or one bull trout are estimated in the trap on Saturday or Sunday, the trap will be processed on that day.
4. Broodstock is collected on Monday and Thursday of each week, or as needed. Extra broodstock can be collected if there is a deficit in brood numbers the prior week.
5. On all other days collected fish are passed above, held for broodstock in pen, recycled, or transported to Wallowa Hatchery.
6. Refer to Appendix E; Standard Operating Procedures for Imnaha River Weir, for further details.

iv. Trap installation and scheduling

1. Upper Grande Ronde and Catherine Creek
 - a. Lead agency CTUIR.
 - b. Trapping begins in March to monitor steelhead abundance, and continues through late-July if river conditions allow. The traps are typically staffed overnight occurs after mid-April.
 - i. The Upper Grande Ronde weir is removed when the daily maximum water temperature exceeds 68° F (20° C).
2. Lookingglass Creek
 - a. Lead agency ODFW.
 - b. The intake trap at Lookingglass Hatchery is operated from early March (or as conditions allow) until no fish are captured for 10 days after September 1. Operators should note when pickets are removed.
3. Lostine River
 - a. Lead agency NPT.

Trapping begins mid-February for steelhead monitoring. Chinook are typically trapped beginning in May. The trap is operating until no Chinook are captured for 10 days after September 1. During the pilot phase of the Grande Ronde Coho Salmon program, trapping will continue until December 1.
4. Imnaha River
 - a. Lead agency ODFW.
 - b. Trap is installed in early June, or as soon as river conditions allow, and operated until mid-September or the last scheduled spawning ground survey.
 - c. Staffing guidelines:
 - i. Weekdays - Two (2) ODFW staff each day (one Lookingglass Hatchery staff will stay at the weir and one Wallowa Hatchery staff travel on site). NPT provides one (1) staff that will travel on site each day.
 - ii. Weekends - ODFW provide two (2) staff daily (one Lookingglass Hatchery staff stay on-site, another travels on-site on Friday to stay the weekend). NPT provides

one (1) staff that will travel on site each day. NPT haul recycles and/or outplants. Any surplus fish beyond the capacity of the NPT tanker can be placed in a holding pen. Alternatively, one hatchery staff may haul fish.

v. Site-specific weir management guidelines

1. Upper Grande Ronde River - The Grande Ronde conventional program calls for collection of up to 50% of natural fish and up to 100% of conventional return to reach the broodstock goal (Table 1).
 - a. Fish are handled without anesthesia.
2. Catherine Creek – Pre-season run projections and the sliding scale (Table 2) are used to determine brood collection and natural spawning objectives, and may be reassessed with PIT tag data during the run.
 - a. Surplus - If surplus hatchery fish are available, the first 50 should be outplanted to Indian Creek, then 100 to Lookingglass Creek below the weir, then 50 more to Indian Creek, and remainder to Lookingglass below the weir. Surplus fish are differentially marked with an opercle punch.
 - b. Fish are handled using electro narcosis.
3. Lookingglass Creek – Pre-season run projections and the sliding scale (Table 3) are used to determine brood collection and natural spawning objectives, and may be reassessed with PIT tag data during the run. If the projected run is under 300 total fish, refer to Lookingglass Creek Management plan. Otherwise apply the following guidelines:
 - a. Fish are handled using electro narcosis, with MS-222 as a backup.
 - b. Mark all hatchery and natural adults released upstream of the weir with a right opercle punch (1-ROP). Provide punches to CTUIR.
 - c. Collect scale samples from all natural origin fish passed upstream.
 - d. Inject broodstock with antibiotics and transport to the adult holding ponds (*see* vii. Adult holding Ponds).
 - e. Release all natural origin jacks upstream.
 - f. Do not release hatchery jacks upstream. Sacrifice all hatchery origin jacks for tag recoveries and provide carcasses to foodbank, or bury. Hatchery jacks without CWT can be food banked, buried, or recycled into lower Lookingglass Creek.
 - g. Do not release identifiable out-of-basin (within or outside Grande Ronde basin) stray salmon upstream.
 - i. If needed, strays from the Upper Grande Ronde River trapped at Lookingglass may be added to the Grande Ronde broodstock or held for return to the river.
 - ii. Catherine Creek strays can be recycled to lower Lookingglass Creek, used for Catherine Creek brood or, if needed Lookingglass brood.
 - h. Additional guidance can be found in the Lookingglass Creek Spring Chinook Management Plan.

4. Lostine River - Pre-season run projections and the sliding scale (Table 4) are used to determine brood collection and natural spawning objectives, and may be reassessed with PIT tag data during the run. The basic collection guidelines are as follows:
 - i. Fish are handled using electro narcosis, with MS-222 as a backup.
 - ii. Release all natural origin jacks upstream. Hatchery jacks should represent a maximum of ten percent (10%) of the total male hatchery fish released upstream. Natural- and hatchery-origin fish may be radio tagged and released at the weir.
 1. Radio-tagged fish are included in estimates of the hatchery/natural composition of fish passed above the weir.
 - iii. Fish that are injected with antibiotics are individually marked.
 - iv. Hatchery-origin fish may be transferred to Wallowa Hatchery for distribution or recycled for harvest.
 - v. Surplus hatchery may be outplanted according to these guidelines:
 1. Hatchery fish can be outplanted to the Wallowa River and tributaries (Bear Creek, Hurricane Creek, and Prairie Creek).
 2. If outplants occur in new reaches, subsequent spawning success will be monitored in target reaches by NPT and/or ODFW staff.

5. Imnaha River - Pre-season run projections and the sliding scale (Table 5) are used to determine brood collection and natural spawning objectives, and may be reassessed with PIT tag data during the run.
 - i. See Appendix E. Standard Operating Procedures for the Imnaha River weir for handling guidelines.
 - ii. ODFW staff will determine fish disposition on-site. Priorities for hatchery fish will be: 1) broodstock, 2) natural spawning above the weir, 3) recycle to fishery, 4) transport to Wallowa Hatchery for CWT recovery, subsistence and/or foodbank use, and 5) outplants. Priorities for natural fish will be: 1) broodstock, and 2) natural spawning above the weir.
 - iii. *Chinook salmon* - Fish retained for broodstock are injected with tulathromycin and oxytetracycline. After broodstock and upstream passage, surplus hatchery jacks and adults can be utilized as described above.
 1. If fish are exposed to MS-222, a 21-day period is required before they are used for consumption. Fish held for distribution are sampled according to Appendix A.
 2. A total of 300 live hatchery adults can be outplanted to Big Sheep Cr. and Lick Cr. If more adult out plants are anticipated, notification should be provided to NOAA Fisheries.
 3. Surplus carcass jacks can be disposed of in Big Sheep Cr. and Lick Cr. Surplus live jacks can be released in Big Sheep after the last redd count survey.
 - iv. *Steelhead* – See Imnaha weir SOP (Appendix E).
 - v. *Bull Trout* – See Imnaha weir SOP (Appendix E).
 - vi. Marking - Fish captured at the Imnaha weir, and not retained for broodstock, will receive an opercle punch that indicates their intended disposition:
 1. Pass above weir = 1 ROP

2. Outplant = 1 LOP
3. Recycle = 2 LOP
4. Double recycle = 3 LOP (previously recycled fish will have another LOP applied).

vii. **Mortalities and carcasses**

1. Before redd surveys: Imnaha staff samples weir mortalities for length, sex, pre-spawn status, scales (natural fish only), recapture (opercle punch), and origin).
2. After redd surveys: Fish research will sample mortalities as described above, then carcasses should be clearly identified as sampled (tails removed) and returned to the river below the weir.
3. Biological data will be sent to ODFW Fish Research (Feldhaus).

vi. **Data collection**

1. Chinook salmon - Enumerate, measure, sex, record origin, and scan for CWT and PIT tags before fish is sent to final disposition.
2. Steelhead - Enumerate, measure, scan for tags and record sex, origin, and fin marks; release upstream. Known hatchery-origin fish are euthanized on Lookingglass Creek. Data and reports sent to ODFW (ODFW District and Regional offices), and LSRCP (Robertson).
3. Bull trout - Enumerate and estimate length (minimize handling). Data and reports sent to ODFW (ODFW District offices), and LSRCP (Robertson).
4. Trap mortalities
 - a. Label, freeze, and provide the first 10 site mortalities to Fish Health for examination. Following examination, carcasses should be disposed of in the landfill.
 - b. Freeze fish dead more than 24 hours.
 - c. Weir mortalities or other pre-spawning mortalities discovered during stream bank surveys or unusual loss will be coordinated with fish health.
 - d. Send mortality data to ODFW (Feldhaus) and CTUIR (Naylor, Crump)
 - e. Following examination, carcasses may be disposed of in the landfill.

vii. **Broodstock transportation**

1. CTUIR and NPT will transport fish from Upper Grande Ronde, Catherine Creek, and Lostine River sites. ODFW or NPT will provide transportation of fish from the Imnaha weir to Lookingglass Hatchery.
2. Broodstock should be hauled daily, and not held more than 72 hours. Imnaha broodstock will be hauled on Monday and Thursday, or as needed to make broodstock.
3. Drivers should complete a transfer data sheet, provide to Lookingglass Hatchery staff for data entry in the HMIS system.
4. Minimize shock by hauling during morning hours to take advantage of cooler stream temperatures. Temperature differences between transport container and facility water should not exceed 10° F (5.6° C). Temper if needed.
5. Handling - Fish will be netted from the transport tank and placed in holding tanks at Lookingglass Hatchery. Lookingglass Hatchery personnel will record all observations on data sheets and report to Fish Health at the end of the season.

viii. Adult holding at Lookingglass Hatchery

1. Upper Grande Ronde – One (1) adult holding pond
2. Catherine Creek – Endemic building, circular tank 20
3. Lookingglass Creek – One (1) adult holding pond, food bank fish held in circular tank 19
4. Lostine River – One (1) adult holding pond, food fish held at Wallowa Hatchery
5. Imnaha River – One (1) adult holding pond, food fish held at Wallowa Hatchery

D. Spawning, Incubation, and Rearing

- i. Key Contacts – Carrie, Deb, Mike, Rick, Deb, Sally, Rick
- ii. Schedule - The first sort will occur in early to mid-August, with an expected first spawn during mid-August. Fish are spawned weekly as described below:
 1. Tuesday – Imnaha and Lostine.
 2. Wednesday – Lookingglass Creek.
 3. Thursday – Upper Grande Ronde and Catherine Creek.
- iii. Anesthetic – Electro-anesthesia for all stocks.
- iv. Spawning – Broodstock are spawned at Lookingglass Hatchery. Sorting and spawning for each stock takes place on the same day.
 1. Hatchery and co-manager staffs will determine fertilization matrices.
 2. All Tyvek tag numbers are recorded on the spawning matrix sheets.
- v. Pairing
 1. Fertilize a maximum of 10% of the eggs with three year old males.
 2. Fertilize a minimum of 30% of the eggs with five year old males (UGR, LGC, and CC males >80cm, and LR and IM males >85cm).
 3. Large males may be spawned up to 3 times.
 4. Conduct jack spawning with 1 female to 1 jack matrix. Most adult spawning matrices will be 2 females x 2 males, but matrices of 1 x 1, 1 x 2, 2 x 1, or 3 x 2 can be used if necessary.
 5. Incubate fertilized eggs at Lookingglass hatchery.
 6. Determine fecundity at eye-up.
- vi. General fertilization procedures
 1. Sort and euthanize ripe females.
 2. Collect eggs preventing addition of outside containments (other body parts).
 3. Store eggs separately for each individual female.
 4. Drain ovarian fluid from eggs.
 5. Sort males, spawn in dry cup.

6. Mix sperm with eggs, activate with pathogen free water (~100 ml).
 7. Wait 60 seconds, rinse eggs.
 8. Treat fertilized and rinsed eggs in 100 ppm iodophore solution for minimum of 45 minutes.
 9. Tray eggs, 1 tray per individual female.
- vii. Surplus - Fish that are surplus to broodstock needs may be returned to stream. Fish injected with antibiotics will not be used for human consumption or released where legal harvest is possible.
- viii. Staffing support - CTUIR will provide fish culture support for Upper Grande Ronde, Catherine, and Lookingglass stocks. The Nez Perce Tribe will provide fish culture support for spawning of the Imnaha and Lostine River stocks.

E. Incubation

- i. All eggs will be incubated to eyed stage at Lookingglass Hatchery. Until eye-up, segregate individual females. After eye-up, eggs will be enumerated and segregated by disease profile. If possible, only low risk BKD eggs will be reared (<0.200 OD units).
- ii. Eggs are incubated using chilled and un-chilled well water and UV treated Lookingglass Creek water in Heath incubators. Untreated creek water may have to be used in the event of power outage or other emergency.

F. Smolt Production

- i. Size at release - Smolts are produced at a target size of 20-25 fpp at release.
- ii. Early Rearing – Fry are reared in double deep troughs at Lookingglass Hatchery on UV-treated Lookingglass Creek water. Fish are transferred outside to Lookingglass Creek water in April or May. Trough loading will be range from 30,000-50,000 per trough.
- iii. Final Rearing – In May, fish are moved to outdoor raceways. After marking and spawning, fish will be placed in final rearing containers. Refer to AOP for final ponding plan.
- iv. Lookingglass Hatchery raceway allocation - Eleven raceways are allocated for Grande Ronde tributary production, seven raceways for Imnaha production. Lookingglass Creek smolts are reared in the adult holding ponds (AHP). Current allocation by stock is:
 1. Upper Grande Ronde River – 4 raceways.
 2. Catherine Creek – 2 raceways.
 3. Lookingglass Creek – 1 raceway and 4 adult holding ponds.
 4. Lostine River – 4 raceways.
 5. Imnaha River – 7 raceways.

G. Marking

- i. Key Contacts - ODFW (Haugen, Feldhaus, LaPoint, Hoffnagle and Keniry), NPT (Vatland, Vogel)

- ii. Mark Type
 - 1. Adipose clipping –occurs during the last two weeks of August.
 - 2. Coded wire tagging (CWT) - occurs during the last two weeks of August.
 - 3. PIT tagging –occurs during the first two weeks of October.
 - 4. See AOP for marking, CWT and PIT tag number information.

H. Monitoring and Evaluation

- i. Key Contacts: ODFW (Hoffnagle, Feldhaus), CTUIR (Naylor, Crump), NPT (Vogel, Vatland)
- ii. Data is collected on behalf of several ongoing monitoring efforts, including: program evaluations, monitoring differences between natural and hatchery production, growth and survival of natural-origin fish (in collaboration with ODFW Early Life History Project).
- iii. PIT tags are used to detect fish at main stem dams, in-stream arrays, and weirs to assess downstream and upstream survival and timing.
 - 1. Catherine Creek and Imnaha stocks are part of the Comparative Survival Study (CSS), and therefore receives PIT tags from the Fish Passage Center.
- iv. Data collection efforts
 - 1. During pre-liberation sampling the following data is collected per raceway:
 - a. Collect 50 weights
 - b. Collect 250 lengths
 - c. Check 500 fish per tag code for tag retention and fin clip quality
 - 2. While processing adult fish (live or mortalities) at trapping facilities, operators should collect the following data:
 - a. Data collection – count, length, marks/tags
 - b. Tissue collection – snout/scales, kidney sample, genetic samples
 - 3. During broodstock spawning and early incubation, collect the following:
 - a. Length (all fish), weight (females), marks/tags, eyed egg weights, individual fecundity
 - b. Eyed egg weights (separately for natural origin and hatchery origin)
 - c. Tissue collection – snout/scales, kidney sample, genetic sample
 - 4. During spawning ground surveys, the following data is collected:
 - a. Redds (count, GPS)
 - b. Live fish (count)
 - c. Carcasses (count, length, marks/tags, snout/scales, kidney sample, genetic sample)
 - 5. Genetic tissue collection for monitoring and pedigree analysis
 - a. 50 samples/stock

2. Grande Ronde and Snake River Fall Chinook Program

A. Smolt Release

- i. Grande Ronde River - Priorities 13 and 15 in the *U.S. vs. Oregon* production tables target a total production of 400,000 sub-yearlings scheduled for release in the Grande Ronde River around May 29 at 50 fpp. Marks for this release are as follows:
 1. 200,000 AdCWT
 2. 200,000 no marks

- ii. Snake River - Priorities 14 and 16 in the *U.S. vs. Oregon* production tables target a total production of 1 million sub-yearlings, and scheduled for release in the Snake River at Hells Canyon Dam in late May or early June at 50 fpp. Marks for this release are as follows:
 1. 200,000 AdCWT
 2. 800,000 Ad only

B. Adult collections and spawning – refer to Lyons Ferry AOP.

C. Incubation and Rearing

- i. Incubation at Lyons Ferry - After eye-up, inventory, and disease profiles, Lyons Ferry staff will combine eggs and ship to Irrigon Hatchery in December. Only eggs from females below BKD titers levels 0.2 are transferred.
- ii. Fish are reared and tagged at Irrigon Hatchery prior to release.

D. Key Contacts

- i. Lyons Ferry Hatchery (Bumgarner), ODFW (Deal, Keniry, Yanke), CTUIR (Zimmerman), NPT (Johnson), IDFG (Putnam), IPC (Rosenburger)

3. Grande Ronde Coho Salmon Program

A. Overview

- i. Consistent with production tables specified in the *U.S. vs. Oregon* fish management agreement, the production objective for this program is 500,000 smolts released in the Lostine River.
- ii. Management objective – To reintroduce coho salmon to Northeast Oregon, continue to provide measurable harvest benefits that these fish provide for treaty and non-treaty fisheries in the mainstem Columbia, and develop fisheries in the Snake and Grande Ronde rivers. Concurrently this program could reestablish natural production of coho salmon in the Wallowa/Lostine River and provide the opportunity for natural recolonization of coho in Grande Ronde tributaries (e.g., Minam and Wenaha rivers) that historically produced coho salmon.
- iii. The coho program described in this document is the first step of a phased approach to reintroduce coho salmon to the Grande Ronde basin. This first ‘pilot’ phase will direct stream release smolts during 2017-2024 and evaluate subsequent adult returns to determine if this Grande Ronde program can be self-sustaining.

B. Smolt release

- i. Final rearing - Fish are reared to smolt at Cascade Hatchery.
- ii. Transportation – Coho smolts will be hauled from Cascade Hatchery to the Lostine River for direct stream release in during March. Transportation is coordinated between NPT and ODFW.
- iii. Release – Fish are direct-stream released into the Lostine River from Cascade Hatchery. The release site and timing will be in the vicinity of the Lostine River acclimation site during March or April, and will be coordinated annually between NPT and ODFW.
 1. Rotary screw trap operators in the Lostine River should be notified prior to release. Key contacts are: Garner and Jonasson (ODFW), and Vatland (NPT). Screw trap is removed from operation (cone raised) for 48 hours after release. After that, NPT provides support to the Lostine River trap until trap resumes normal operations.

C. Adult collections and spawning

- i. Trap installation and scheduling
 1. The first returns of adult coho salmon will be trapped at the Lostine River weir beginning in 2017 (jacks only), adults in 2018.
 2. The Lostine River weir will remain in operation following trapping for Chinook salmon, and all applicable guidelines described above for spring Chinook salmon will be followed (see Grande Ronde and Imnaha Spring Chinook Salmon, Adult Trapping).
 3. Broodstock is not collected during this pilot phase of the program. Coho salmon are enumerated and sampled for marks (see monitoring and evaluation below).
 - a. During the pilot phase, disposition of coho will be coordinated annually with NPT and ODFW.

- ii. Spawning - Tanner Creek stock from the Bonneville Complex or other Tanner Creek sources will be used for the program until a local broodstock can be developed.

D. Incubation, and rearing

- i. Incubation and rearing – Rearing from eyed egg to smolt will occur at Cascade Hatchery.

E. Marking

- i. Smolts are marked consistent with the following guidelines, and revisions to marked schemes may be coordinated in the *U.S. vs. Oregon* forum:
 - 1. Adipose clipping – 100%
 - a. For BY15 only (release 2017) these fish are not adipose fin clipped.
 - 2. Coded Wire Tagging (CWT) - Appropriate level of CWT to evaluate harvest contribution (~60,000 AdCWT).
 - a. For BY15 only (release 2017) this group is approximately 66,000.
 - 3. PIT Tags - A portion of juveniles will be implanted with PIT tags to assess downstream survival (initial goal of 5,000 AdPIT, depending on funding).
 - a. For BY15, Mitchell Act funds were made available for 5,000 PIT tags, but because of litigation they were not procured prior to juvenile release.

F. Monitoring and evaluation

- i. Objectives - During the pilot phase of this program, monitoring objectives will be limited to the following:
 - 1. Enumerate adult returns to weir.
 - 2. Estimate smolt-to-adult survival of release, contribution to fisheries, and straying using Coded Wire Tags (CWT).
 - 3. Determine if natural spawning is occurring in the Lostine River.
 - a. Other Grande Ronde tributaries may be surveyed (Minam, Wenaha, Wallowa Rivers).
- ii. Data Collection – Data will be limited to adult returns at the Lostine River weir, but may be expanded if resources allow. At the Lostine weir, operators collect the following data:
 - 1. Enumerate coho catch.
 - 2. Disposition, as decided by co-managers annually.
 - 3. Examine for external marks.
 - 4. Estimate sex and length of individual fish.
 - 5. Scan for CWT and PIT Tags, retain and bag snouts.

G. Key Contacts

- i. Key Contacts – B. Johnson, Zollman, Hesse, Vatland (NPT); Requa, Harrod, Woods, Myatt, Yanke, Ruzycki (ODFW); Zimmerman (CTUIR); Mark Traynor (ODFW Cascade Hatchery).

4. Grande Ronde Lamprey

A. Overview

- i. Objectives - The purpose of this stop gap effort is to avoid local extirpation in the Snake River Basin and maintain a population of ammocoetes that serve as a source of pheromone attractants drawing adults upstream to spawn in the abundant habitat in this region, thereby continuing a presence in the Snake River Basin until upstream adult and downstream juvenile passage problems are identified and corrected, and healthy, harvestable populations are restored.
- ii. CTUIR program
 1. Adult collection – Adult lamprey are trapped and collected by CTUIR at main stem dams on the Columbia River.
 2. Adult holding – Once trapped, lamprey are held at Minthorn Springs facility in the Umatilla basin.
 3. Adult releases – Lamprey are released to spawn naturally in the upper Grande Ronde at River Campground (~RM 163) and into Catherine Creek at Catherine Creek State Park (~RM 49) and Lookingglass Creek upstream of river mile 5. Release locations will be determined on an annual basis, and as availability of lamprey allows.
- iii. NPT Program
 1. Adult collection – Adult lamprey are trapped and collected by NPT Fisheries staff at Bonneville, The Dalles, and John Day dams.
 2. Adult Holding – Once trapped, lamprey are held at Nez Perce Tribal hatchery on the Clearwater River through winter months.
 3. Adult releases – Adult Lamprey are released to spawn naturally in tributaries of the Clearwater and Grande Ronde Rivers, including Asotin Creek in Washington. In Oregon, lamprey have been released in the Wallowa and Minam Rivers, and Joseph Creek. Release locations will be determined on an annual basis, and as availability of lamprey allows.
- iv. Fish Health
 1. Fish Health recommends an examination (up to 5 grab-sampled) be conducted prior to lamprey being transferred to Oregon waters. At a minimum, all moribund and dead lamprey should be examined during rearing in Oregon and Idaho to develop a pathogen history. If unable to lethally sample due to tribal policy, then develop a pathogen history as best as possible with moribund and dead lamprey. For lamprey releases in Oregon, Fish Health recommends source lamprey for holding in Idaho come from direct transfer from the dam collection site or the CTUIR holding site near Pendleton, OR rather than Yakima Indian Nation site near Prosser.

B. Key Contacts

- i. Key Contacts – Harbeck, Vatland (NPT); Yanke, Bailey (ODFW).

Table 1. Upper Grande Ronde River weir management guidelines for broodstock collection and passage.

Total adult escapement to the mouth (HOR+NOR)^a	HOR at mouth (%)	Maximum NOR in Broodstock (%)	HOR to retain for Broodstock (%)^b	HOR adults released above weir (%)	Minimum NOR in broodstock (%)	Strays allowed above weir (%)^c
Any	Any	Up to 50	Up to 100	Up to 100	– ^d	≤5

^a Pre-season estimate of total escapement

^b Conventional hatchery adults only, all captive brood adults released to spawn naturally or outplanted

^c For hatchery adults originating from different gene conservation groups (Rapid River stock or strays from outside the Grande Ronde basin)

^d Not decision factor at this level of escapement, percentage determined by other criteria

Table 2. Catherine Creek Spring Chinook broodstock/upstream passage management guidelines.

Total adult escapement to the mouth (HOR+NOR)^a	HOR at mouth (%)	Max of NOR Run in Broodstock (%)	HOR to retain for Broodstock (%)^b	HOR adults released above weir (%)	Minimum NOR in broodstock (%)	Strays allowed above weir (%)^c
< 250	Any	40	40	– ^d	– ^d	≤5
251-500	Any	20 ^d	20	≤70	≥20	≤5
>500	Any	≤20	– ^e	≤50	≥30	≤5

^a Pre-season estimate of total escapement

^b Conventional hatchery adults only, all captive brood adults released to spawn naturally or outplanted

^c For hatchery adults originating from different gene conservation groups (Rapid River stock or strays from outside the Grande Ronde basin)

^d Not to exceed 150,000 smolt production

^e Not decision factor at this level of escapement, percentage determined by other criteria

Table 3. Lookingglass Creek weir management guidelines for broodstock collection and passage.

Escapement Level	Passed Above (%)	Retain for Broodstock (%)
150	67	33
200	60	40
250	55	45
300	50	50

>300 – adjustments will be made based on brood needs. If brood need has been met remainder to be released upstream

Table 4. Lostine River weir management guidelines for broodstock collection and passage. Assumes program goal of 250,000 smolts (166 adults for broodstock).

Estimated Natural Run of ADULTS to River Mouth (proportion of minimum abundance threshold recommended by ICTRT¹)	Number of ADULT Natural Fish to River Mouth	Max % Natural ADULTS for broodstock	Number of ADULT Natural Fish Retained for Broodstock (Proportion of Natural Brood)	Max Proportion of ADULT hatchery fish released above weir	Minimum % natural ADULTS in Broodstock
< 0.05 of Critical	< 8	0	0	NA	NA
0.05 – 0.5 of Critical	8 – 74	50%	4 – 37	NA	NA
0.5 of Critical - Critical	75 – 149	40%	30 – 60	70%	20%
Critical – 0.5 of Viable	150 – 249	40%	60 – 100	60%	25%
0.5 Viable – Viable	250 – 499	30%	75 – 150	50%	30%
Viable – 1.5 Viable	500 – 749	30%	150 – 225	40%	40%
1.5 – 2.0 Viable	750 – 999	25%	188 – 250	25%	50%
>2.0 Times Viable	> 1,000	25%	> 250	<10%	100%

¹ Lostine River contributes about 50% of production for Wallowa/Lostine Population - Viable level is 50% of TRT recommended minimum abundance threshold for Wallowa/Lostine population (1,000) after broodstock collection and fishery.

Table 5. Sliding scale management tool for Imnaha Chinook hatchery program utilized for managing disposition of Chinook salmon adults for broodstock and escapement to natural spawning areas (Table 1 excerpted from Imnaha spring Chinook HGMP May 2011).

Estimated natural run of ADULTS to river mouth as a proportion of minimum interior TRT minimum abundance threshold (MAT)	Number of ADULT natural fish to river mouth	Expected handle rate at weir of ADULT natural fish (50%)	Max % natural ADULTS for broodstock²	Number of ADULT natural fish retained for broodstock	Max proportion of ADULT hatchery fish released above weir	% Natural ADULTS in Broodstock³
< .05 of Critical ⁴	< 15 ⁴	< 8 ⁴	0	0	NA	NA
.05 - .5 of Critical	15 - 149	8 - 74	50%	04 - 37	NA	NA
.5 – Critical	150 -299	75 -149	40%	30 - 60	70%	20%
Critical - .5 of MAT	300 - 499	150 -249	40%	60 - 100	60%	25%
.5 Viable – MAT	500 - 999	250 - 499	30%	75 - 150	50%	30%
			40%¹			
Viable - 1.5 MAT	1000 - 1499	500 - 749	30%	150 - 225	40%	40%
			40%¹		30%	
1.5 - 2 x MAT	1500 - 1999	750 - 999	25%	188 - 250	25%	50%
> 2 x MAT	> 2000	> 1000	25%	> 250	<10%	100%

¹ Percentage highlighted will be implemented after three consecutive years (or 3 year mean) at viable or greater

² Maximum percent of wild adults handled at weir collected for Broodstock

³ Target percent natural adults in Broodstock

⁴ In the submitted HGMP, the less-than (<) symbols shown above are displayed in error as greater-than (>) symbol

Appendix A. Disinfections and Sanitation Guidelines for all LSRCP Hatcheries.

Specific Operational Recommendations

For background on the importance of these recommendations see page 1 of Appendix C (2013 AOP)

Applies to Who?	Prevention Control Measure or Sanitary Practice	Guideline Comment
All	Disinfect all gear/equipment prior to entering or leaving hatchery grounds	-As per attached iodophor protocol -Hatchery crew responsible for providing tub of 100 ppm iodophor
Hatchery Crew	Do not go from adult handling operations to juvenile operations activities unless all bib gear is thoroughly disinfected.	-As per attached iodophor protocol -it would be preferable to have bib gear designated for either adult or juvenile use.
Hatchery Crew	Pick mortality on a daily basis	-This is consistent with ODFW's statewide Fish Hatchery and Fish Health Management Policy.
All	Disinfect equipment when moving from raceway to raceway or tank to tank for <u>any</u> fish handling or pond cleaning activities	-As per attached iodophor protocol -Includes CWting, fin clipping and PIT tag operations. See footnote for marking.
All	Use footbaths upon entering or leaving the work area for a given program	-Use larger tub of disinfectant if involved in a spawning
All	Use a new disposable apron or disinfected personal rain gear while working with fish	
CTUIR Personnel operations at Lookingglass Hatchery	Disinfect all gear/equipment prior to entering or leaving hatchery grounds, Lookingglass Creek, or the intake building and when done with operations at intake	-CTUIR personnel responsible to maintain and use a tub of 100 ppm iodophor at intake building workstation
Hatchery Crew	Assure that individual raceway and tank mortality "picker equipment" is in place at each raceway and tank	-All use these for the specifically designated Raceway
Hatchery Crew	Sanitize each raceway prior to use for the next brood year. (see page 3 for recommendation)	-dry for a minimum of three days
Hatchery Crew	Keep footbaths located at strategic locations refreshed with disinfectant	-As per iodophor label, refreshed as needed
People at Spawnings	Disinfect the spawning table and spawning work area between stocks and at the end of the day	-As per attached iodophor protocol
Research, Hatchery Crew & Fish Health Personnel	Handle and necropsy dead fish only in designated areas	-Adult morts: use concrete pad outside spawn area or concrete pad in endemic building at LGH -store snouts only in adult mortality freezer -Juvenile morts: store in freezer in designated area for this purpose.
PIT taggers	-PIT tagging supervisors maintain and keep footbaths by each door of PIT tagging trailer for use during operations -Assure that PIT tagging needles are new or clean and sharp -Disinfect in 70% Isopropyl alcohol -No re-use of PIT tag needles until air dried	-if PIT tag needles are re-used disinfect as per isopropyl protocol attached
Lib Truck Operators	Assure proper disinfection of tank and equipment prior to collection or transfer	-As per attached disinfectant application Summary

**Appendix A (page 2 of 2): Disinfection and Sanitation Guidelines for all LSRCP hatcheries
Summary of Recommended Disinfectants (Concentration and time) and for what Application**

Disinfectant*	Application	Concentration	Time	Comment
Iodophor	Nets, gear and equipment, clipping & tagging van, PIT tag stations, large tub disinfectant containers, spawning colanders and buckets, lib truck, footbaths, floors Note: For raceway sanitization** – thoroughly clean the unit to remove dirt, spray or brush on 75-100 ppm iodophor and let this remain for a minimum of 10 minutes. Leave it to dry for a minimum of 3 days. Allow iodophor to dry and break down with exposure to light. **If the above recommendation cannot be done then sanitize raceways by thoroughly cleaning them and leaving to dry for a minimum of 3 days.	100 ppm Note: to make 100 ppm solution mix 6.7 oz of jug strength iodophor to 5 gallons H ₂ O or 6.7 oz.=189ml	10 min.	-Equipment should be pre-rinsed to remove dirt, mucus or other organic material which reduces the efficacy of disinfection and sanitization -Rinse equipment to remove harmful residue if equipment is going into standing water containing fish or fish are being placed into the equipment (tank or bucket). Remember that iodine at 1:20,000 is harmful to fish. -Argentyne or other buffered iodophors such as Western Chemicals “PVP iodine” would be acceptable. Note: if DRAW 476 is used remember this product is 1.75% active iodine and unbuffered so should not be used for water- hardening eggs
	Water hardening eggs	• 100 ppm	Minimum 15 minutes	This is the statewide general practice
	Egg transfers - disinfection at receiving station	100 ppm	10 minutes	
Virkon Aquatic	Footbaths, nets, boots & gear			As per label
Chlorine or Aqueous solution as sodium hypochlorite (Household Bleach)	Lib truck tanks	10 ppm	10 min.	Organic matter binds and neutralizes
	Raceway disinfection	100 ppm		Left to dry and breakdown in sun. Need to assure that no bleach goes to effluent.

^aWithin a stock, operations will start with groups determined to be of lowest disease risk proceeding to raceways of higher disease risk. The latest fish health information should be used to determine the least risky raceway sequence.

^b All chemical use will be done in accordance with label use and reporting requirements. Disinfecting and disinfected water must be disposed of in an approved manner.

Appendix B. Juvenile Chinook fish health monitoring plan and disease treatments.

Location	Brood year	Stock	Examination Category	Protocol	Comment/Disease Treatment
Lookingglass Hatchery	2016	200 201 80 29 81	Monthly	-10 mort/moribund per stock, kidney smears on TYE-S agar, gill culture smears if suspect gill disease, R. salmoninarum (BKD) if suspect disease or starting monthly sampling by October 2015, Gill & skin wet mounts from a combination of moribund and healthy fish. -tissues (gill/ kidney/spleen) will be assayed for culturable viruses from a sub-sample of fish -5 grab-sampled fish every other month and any moribund fish for EIBS (blood smears and hematocrits).	A 21-day Aquamycin 2.25% feed will be implemented for all raceways for each stock starting in July 2017. Disease outbreaks - treated on a case-by-case basis. Therapies and remedial measures are based on conventional and available treatments, new information, and innovation. Warm water temperature therapy would be used if EIBS became a problem on a priority basis determined by co-managers. Formalin treatments may be implemented for Ichthyobodo (cosita) infestations. Fungus - Formalin flushes (1 hour) are prescribed after fin clipping, PIT tagging, VIE tagging and coded wire tagging for two consecutive days. Formalin is used under a veterinarian prescription. Treat CWD with Florfenicol using a Veterinary Feed Directive (VFD) INAD.
Lookingglass Hatchery	2015	200 201 80 29 81	Monthly Pre-transfer & Annual Myxobolus cerebralis testing	Monthly: As above Pre-transfer: 60 grab-sampled smolts per stock -R. salmoninarum by ELISA -tissues (gill/kidney/spleen) from 3 fish pools for culturable viruses -wet mounts of skin & gill tissue from a minimum of 5 fish -sub-sample for EIBS -one stock on water supply for 6 months (60 fish) for <i>Myxobolus cerebralis</i>	Pre-transfer grab-sample numbers may vary depending on disease history and number of fish for a given brood year. For future years a small group rainbow/steelhead should be reared at Lookingglass Hatchery for annual <i>M. cerebralis</i> testing 181 days after ponding.
Chinook acclimation IM, LR, CC & UGR	2015		Pre-liberation	-Smolt groups held at acclimation sites longer than 3 weeks will be evaluated with a lesser number of “grab-sampled” fish as in pre-transfer protocol above. -Mortalities will be examined (as in monthly)	Pre-liberation grab-sample numbers at acclimation sites may vary depending on disease history and number of fish for a given brood year.

Appendix C. Adult Chinook fish health monitoring plan and disease treatments at Lookingglass Hatchery.

Stock	Examination Category	Protocol	Comment
200 (LR) 201 (CC) 80 (GR) 29 (IM) 81 (LGC)	Adult Spawners (Broodstock)	-A minimum of 60 females will be sampled for virus from each stock (or all females if <60). Samples will be individual ovarian fluid and caeca/kidney/spleen sample pools not to exceed 5 fish. - All females for BKD by ELISA	ELISA results will be used to implement BKD prevention control through culling of eggs known to be of higher risk.
200 201 80 29 81	Prespawning Mortality	- Up to 5/stock (before Sep 1 st): -Kidney sampled for BKD by ELISA -systemic bacteria by culture	<ul style="list-style-type: none"> • Note: additional mortality may be sampled • Lookingglass Creek mortalities will be worked up with CTUIR staff to assure data collection covers all the needed information
81 or 201 LG-CK	Spawning Ground Survey	-Collect a sub-sample of 30 kidney samples from adult Chinook above the weir (hatchery intake)	<ul style="list-style-type: none"> • Fish Health Request

Appendix D. Disease treatments and other drugs for adult Chinook Broodstock.

Location	Brood Year	Stock	Treatment for	Chemical/Drug	Protocol	Comment
Lookingglass	2017	200 201 80 29 81	Fungus Control	Formalin Hydrogen Peroxide	Formalin administered a minimum of 3 days per week at 167 ppm for 1 hr. (Veterinary prescription) Hydrogen peroxide 3 days per week at 100 ppm	If formalin cannot be used then use hydrogen peroxide (second choice) Continue treatments throughout the entire spawning season.
Lookingglass, Catherine Creek, Upper Grande Ronde and weirs	2017	200 201 80 29 81	BKD Furunculosis- Enteric Redmouth & other gram negative bacterial infections	Tulathromycin DRAXXIN-25 Oxytetracycline	Injection 5 mg/Kg(Veterinary Prescription) Injection 10 mg/kg (Veterinary Prescription) Inject fish kept for broodstock and reinjection only if deemed necessary Note: For the Lostine River stock only 50% of the broodstock will be injected with both antibiotics to evaluate the efficacy of injections.	Erythromycin will be available via INAD for the 2017 season if desired. DRAXXIN (tulathromycin) should be used via prescription for continuity of the study. If DRAXXIN is used, injected fish will not be used for human consumption, released where legal harvest is possible or used for nutrient enhancement.

Appendix E. Standard Operating Procedures for Imnaha River Weir

Standard Operating Procedures (SOP) for processing fish at the Imnaha River Weir

Operations Year 2017

Purpose: The Imnaha satellite facility is an adult collection and juvenile acclimation and release facility for the Imnaha River Spring/Summer Chinook Salmon Program. The adult weir and trap collects returning Chinook salmon for program broodstock and allows for selective management of natural spawners upstream of the facility. During annual operations, non-target ESA-listed species are also encountered at the weir site. Bull trout are collected and marked for ongoing population monitoring, and post-spawn steelhead kelts are migrating downstream past the weir. This document is intended to provide guidance to operational staff for handling and processing ESA-listed species at the Imnaha facility, with the goal of minimizing stress and risk of latent injury to each individual.

Annual Coordination: Imnaha River weir operations are coordinated with funding agencies and co-managers during the Lower Snake River Compensation Plan (LSRCP) annual operations plan (AOP) process. Co-managers agree that the annual planning process will be consistent with this SOP, and meet the intent of this guidance. Significant deviations in annual operations may require broader coordination and an update of the SOP. Prior to each trapping season, staff involved in operating the weir facility should review this document, the AOP, ESA-permits, and any associated Section 7 consultation documents, and understand any constraints regarding changes and/or modifications to procedures pursuant to these other documents.

Equipment List by Species

<u>Chinook Salmon</u>	<u>Bull Trout</u>	<u>Steelhead</u>
<ul style="list-style-type: none">• Large dip net(s)• Electronarcosis unit• Measuring board• PIT tag reader• Coded wire tag (CWT) reader• Extra batteries (AA)• Operculum punches• Genetic samples• Syringes• Needles• Antibiotics• Sharps container• Electronic thermometer• Datasheets	<ul style="list-style-type: none">• Dip net(s)• Electronarcosis unit• Measuring board• PIT tag reader• Electronic thermometer• PIT tag injectors• PIT tag needles and tag (single use)	<ul style="list-style-type: none">• Long-handled dip nets• Anesthesia vessel• MS-222• Scale Cards• Genetic samples• Measuring board• PIT tag reader• PIT tagging equipment

Trapping operations

1. The Imnaha River weir is scheduled to be installed by June 5th and operated until the second Friday in September, or until the last scheduled spawning ground survey for Chinook salmon. The exact installation date will depend on environmental conditions.

2. The adult weir and trap will be staffed 24 hours per day, 7 days per week, while operational, beginning with the capture of the first Chinook or bull trout.
3. From initial operations through the last week of July, fish will be removed and processed daily.
4. After the last week of July and through the end of operations, fish will be removed and processed daily from Monday-Friday. During this latter period, if 10 or more natural Chinook salmon, or 30 or more fish total, are estimated in the trap on Friday and/or Saturday fish will be removed and processed during the weekend. If a bull trout is observed in the trap on Friday and/or Saturday, all fish will be removed and processed during the weekend.
5. Trap modifications
 - a. A finger weir in the bottom of the ladder with a mesh cover will be installed. This should address potential issues and concern with fish movement in and out of the ladder.
 - b. A set of grader bars may be installed over the entrance to the tower. Bars will be spaced (2.5 inches) to exclude Chinook salmon, but allow bull trout to find refuge in the tower.
 - c. Modifications to the trap can occur in-season if impacts to fish health/survival are being addressed.

Morning trap and weir check – Each morning, on-site staff at the Imnaha satellite facility will check trap conditions, recover weir and/or trap mortalities, and communicate staffing needs to Lookingglass Hatchery.

1. Measure facility temperatures in the raceway using an electronic thermometer. If water temperatures are expected to exceed 16⁰C (61⁰F) by 10:00 AM, notify staff of expedited bull trout procedures (see below).
 - a. If temperatures in excess of 16⁰C (61⁰F) are anticipated during subsequent processing days, notify Lookingglass Hatchery manager. Processing times may be adjusted to start earlier to avoid higher temperatures for all species.
2. Estimate the numbers of Chinook (by-origin) and bull trout in the trap.
 - a. If more than 30 Chinook salmon are estimated in the trap, assume that at least 10 are natural-origin.
 - b. Bull trout can be differentiated from Chinook salmon by the presence of a white leading edge on their fins.
 - c. On Fridays and Saturdays, if trap abundance thresholds are met or a bull trout is encountered notify Lookingglass Hatchery manager to ensure staff is scheduled for weekend processing (*see above*; Trapping Operations, item 4).
3. Recover trap and weir mortalities.
 - a. Each afternoon/evening, on-site staff will check for Chinook salmon, steelhead, and bull trout mortalities.
 - b. Inspect the intake and inspect the trap and surrounding walkways (for jump-outs). Remove any mortalities from the trap.
 - c. Inspect the upstream face of the weir for mortalities. Recover if flows allow.
 - d. Mortality Recovery Procedures
 1. Bull Trout: Scan for existing PIT tag, measure fork and total length, and inspect for hooking wounds. Record location of recovery.
 - a. Retain all recovered mortalities in the freezer at the Imnaha satellite facility for further investigation. Immediately notify and provide recovery information to Jeff Yanke, Wallowa District Fisheries Biologist, Mark Robertson LSRCP, and Rick Wilkison IPC. Care must be taken in handling sick or injured fish to ensure effective treatment and care, and in handling dead specimens to preserve biological material in the best possible state. In

conjunction with the care of sick or injured fish, or the preservation of biological materials from a dead animal, LSRCP has the responsibility to ensure that information relative to the date, time, and location of the fish when found, and possible cause of injury or death be recorded and provided to the Service.

2. Steelhead: NPT will provide a steelhead carcass tote to hold carcasses until NPT staff can examine the fish.
 - a. NPT will process the carcasses similar to methods used for live upstream steelhead.
 - b. After examination carcasses will be passed downstream.
3. Chinook salmon: All weir mortalities recovered before the first regularly scheduled spawning ground survey will be processed by on-site staff.
 - a. After the first annual spawning ground survey, which typically occurs the last week in August or the first week in September, weir carcasses should be processed from Friday to Tuesday.
 - b. For each carcass, weir staff will document the fork length (mm), sex (M, F), fin mark (e.g., AD = adipose clip; none = Unclipped), type of opercle mark present on the carcass (e.g., 1LOP, 1ROP), % spawn for female carcasses (0% = full of eggs, 100% = completely spawned), and presence/absence of a CWT. If there is a CWT, staff will collect a snout and each snout will receive a unique snout identification number (SNID). Weir staff does not need to collect scales or kidney samples. After the carcass is processed, the tail will be removed and the carcass tossed back into the river below the weir.

Processing shed setup – Prior to staff arrival for daily processing, on-site staff at the weir will setup the processing shed and test equipment.

1. Test the tower door and braille to ensure operation before fish are crowded.
2. Inspect hoses, valves, and fittings used to supply water to the processing shed and liberation trucks.
3. Inspect and test oxygen supplies in the holding tank.
4. Setup processing tables and tanks. Place rubber mats around fish processing stations.
5. Ensure that all items listed above (*see*: equipment list by species) are present and operational.
 - a. Test PIT tag reader with a dummy tag.
6. Fill liberation trucks upon arrival. Install electronic thermometers and oxygen sensors, and monitor throughout fish processing.

Crowding / Sorting

1. Begin filling the holding tank. Turn on the oxygen supply and monitor levels.
2. Herd fish in the trap towards the tower door using long handled dip nets.
3. Lower the mechanical crowder on the west end of the trap.
4. Slowly move the mechanical crowder toward the tower door, monitoring for fish that may get impinged on the trap walls or floor.
5. Depending on trap densities, leave enough space for fish to move around freely in-between loading groups of fish into the tower.
6. While loading fish in the tower, move the mechanical crowder as far as possible towards the east end of the trap. Use a long-handed dip net to herd fish along the trap walls towards the tower door.
 - a. When loading a group into the tower, attempt to target no more than 20 Chinook salmon.
7. Once the trap is full and processing equipment is ready, raise the braille slowly to move fish into the holding tank.
 - a. Attempt to transfer no more than 20 Chinook salmon into the holding tank at one time.
8. The holding tank will have a divider, with a cover, that allows separation between Chinook salmon and bull trout and prevents fish from jumping out of the holding tank.
9. To the best degree possible, bull trout should be isolated from Chinook salmon as soon as possible to minimize interactions. Staff can opportunistically net bull trout as they spill from the tower into the holding tank, or net bull trout directly from the larger portion of the holding tank into the portion reserved for bull trout.
 - a. It is not necessary to separate *every* bull trout before processing begins. Additional stress will be caused by attempting to net fish, so staff should isolate what is immediately possible and separate the rest as Chinook salmon are processed.

Bull Trout Processing

1. General guidelines

- a. Bull trout will normally be processed before Chinook salmon. On high volume days, both species can be processed concurrently if staff resources allow.
- b. Bull trout will be processed in a manner that minimizes time out of water and risk of dropping (i.e., use nets at all time).
- c. Bull trout are thermally-intolerant; therefore, staff will note water temperatures prior to processing trap catch each day. Expedited processing procedures will be followed when water temperatures are expected to exceed 16°C (61°F) before processing is completed.

Normal procedures ($\leq 61^{\circ}\text{F}$)	Expedited procedures ($> 61^{\circ}\text{F}$)
<ol style="list-style-type: none"> 1. Anesthetize 2. Scan for existing PIT tags 3. Measure for fork and total length (mm) 4. Inspect for previous injuries, including hooking scars 5. Mark new captures with PIT tag 6. Transfer to liberation truck for release 	<ol style="list-style-type: none"> 1. Anesthetize 2. Scan for existing PIT tags 3. Estimate fork length (within 2-inch size class) 4. Inspect for previous injuries, including hooking scars 5. Transfer to liberation truck for release

2. Anesthetization

- a. Tricaine methanesulfonate (MS-222) will be used as a primary anesthesia; however, electronarcosis (EN) will be the secondary method for anesthetizing bull trout including use of the EN gloves experimentally in 2016. When EN is used, it will be noted on data forms or for individual fish.
- b. Bull trout will be processed in a separate MS-222/EN tank.
 - i. For MS-222 as an Anesthetic
 1. Bull trout will be placed in separate MS-222 tank for anesthesia. The tank will be monitored for temperature to ensure dosage guidelines are met and/or changed with cool water regularly.
 - a. Dosage = 50-60 mg/L at 13-16C (moderate dosage)
 - b. Duration = lack of equilibrium before 5 minutes, up to 5 minutes in solution.
 - c. Recovery of anesthetized bull trout will take place in the transport truck tank or tank placed on the ground near the fish processing area. Recovery timing will range from a minimum of 10 minutes and a maximum of 120 minutes until release during which time fish will have supplemental oxygen (see Section 5 - Release).
 - d. Fish will be observed in the liberation tank after at least 10 minutes after processing to ensure recovery.
 - ii. For EN use as an anesthetic

1. The tank will be equipped with a cradle and adjustable electrodes to account for variable fish sizes. If EN gloves are used, the bull trout will be held in a freshwater tank during measurements/tagging.
2. Each fish will be oriented with the head facing the anode and tail towards the cathode, and the electrodes near each end of the fish. Orientation is more important than distance to the electrodes.
3. The EN unit will be turned on at the lowest settings and gradually increased until the fish enters a state of narcosis, as described by immobilization, relaxation of muscles, and a lack of upright orientation. Voltage treatments will depend on fish size:
 - a. 600-800mm will be exposed to voltages between 20-25 volts
 - b. <600mm will be exposed to voltages between 35-45 volts.

3. Marking / PIT-tagging

- a. PIT tagging equipment will be supplied by Idaho Power (IPC) and procedure/guidance provided by IPC staff will be followed for the PIT tagging process.
- b. All bull trout will be scanned for an existing PIT tag using a handheld PIT tag reader. While scanning, emphasize the area of the fishes' body around the dorsal fin (see P4 instructions below).
- c. Record existing PIT tag codes as 'recaptures'.
- d. Measure
- e. Record both fork and total length (mm) for new and recaptured bull trout.
- f. Inspect for injuries or Record external appearance and identify wounds
 - i. Wound location: Mouth/maxillary, head, body
 - ii. Size: 1) = less than 1 inches; 2) = 1 to 3 inches; 3) = over 3 inches
 - iii. Record whether wound is healed or unhealed?
- g. For previously unmarked (new) fish, apply a 12mm PIT tag to the dorsal sinus using a preloaded syringe.
- h. Properly dispose used PIT tag needles in a designated sharps container.

4. Release

- a. A separate, aerated tank will be reserved for bull trout recovery. Holding in the tank truck (as an option for a separate tank) will be at a minimum of 10 minutes and a maximum of 120 minutes until release. After processing, bull trout will be placed into the release channel via the release tube.
- b. The recovery tank will be monitored for oxygen saturation and water temperature with an in-tank probe. Bull trout will be released immediately should oxygen levels fall below specified minimum levels (80% saturation with 100% goal), or water temperatures rise above 61°F (16°C).
- c. Release Protocol
 - i. Bull trout will be released into the release channel and allowed to voluntarily leave the facility. The exact time of release into the recovery channel will be noted. Fish can be hand netted from the liberation truck or recovery tank and released directly into the recovery channel as well.

1. If this release protocol progresses without bull trout impacts/concerns for the first weeks of 2017 operations, handled bull trout can eventually be released via the release tube and allowed recovery in the release channel.
 - a. Undesirable impacts/concerns would be observation of fish against the weir or any weir mortalities following releases in the channel.
 - ii. Should undesirable impacts/concerns be observed during initial 2017 operations, handled bull trout will be placed into a recovery tank on a truck and then driven up to intake for release (by hand/net) identical to 2016 operations.
- d. Fish will be observed in the recovery tank for at least 10 minutes after processing to ensure recovery.

Chinook salmon processing

1. General guidelines

- a. Natural-origin fish will be differentiated from hatchery-origin fish by the presence of an adipose fin.
- b. ODFW staff will be responsible for determining fish disposition on-site. Weekly and/or daily guidance will be provided by the Lookingglass Hatchery Manager and/or Wallowa District staff.
- c. Co-managers have agreed to the following dispositions, in order of priority:

Natural-origin

1. Broodstock
2. Pass above weir for natural spawning

Hatchery-origin

1. Broodstock
2. Pass above weir for natural spawning
3. Recycle downstream for fishery contribution
4. Tribal distribution and/or foodbank
5. Outplant

2. Anesthetization

- a. Electronarcosis (EN) will be the primary method for anesthetizing Chinook salmon; however, tricaine methanesulfonate (MS-222) can be used as a backup. There will be 2 EN units available, so MS-222 would be a 3rd choice.
- b. MS-222 *cannot* be used for fish dispositioned for recycling and/or tribal distribution/foodbank. If the EN unit is inoperable, these fish will have to be processed without anesthesia.
- c. Each fish will be oriented with the head facing the anode and the tail toward the cathode. Orientation is more important than distance to the electrodes, which can be variable.
- d. The EN unit will be turned on at the lowest settings and gradually increased until the fish enters a state of narcosis, as described by immobilization, relaxation of muscles, and a lack of upright orientation.
- e. Voltage treatments should range between 20 and 45 volts Chinook salmon.
 - i. 600-800mm will be exposed to voltages between 20-25 volts
 - ii. <600mm will be exposed to voltages between 35-45 volts.

3. Processing / Marking

- a. After each fish succumbs to the EN, identify the origin of fish (natural or hatchery), estimate sex (male or female), and record the fork length (mm).
 - i. Sex identification will be based on phenotypic traits (snout shape/size and anal vent characteristics).
- b. Scan for a coded wire tag (CWT) near the snout. Note presence or absence of a CWT.
- c. Scan for a PIT tag near belly and sides of the fish. If present, record the PIT tag code with other biological information.

- d. Designated staff will select a disposition for each fish based on broodstock and natural spawning objectives for that week.
 - e. Salmon selected to spawn upstream will be released in the existing upstream release tube. All other dispositions will be transferred to the respective liberation truck for transport.
 - i. Fish designated for tribal distribution / food bank, or outplanting, may be held on-site until transportation is available. These fish will be held in a net pen in the rearing pond.
4. Electronic PIT tag data collection (P4)
- a. This section to be developed if provided with necessary equipment.
5. Release
- a. After processing, Chinook salmon will be transferred via hand net to a liberation truck.
 - b. The tank will be monitored for oxygen saturation and water temperature while in transfer.

Steelhead Processing – During the early portion of the trapping season, upstream-migrating steelhead may be encountered in the Imnaha weir trap. The Nez Perce Tribe will provide staff, on request, to assist with processing steelhead. Adult steelhead observed in the Imnaha trap box they will be handled, processed and passed upstream. NPT staff will be at the Imnaha weir to assist on a daily basis until June 20 or until it can be determined that adult steelhead will no longer be present at the weir site. In an emergency situation, NPT can also be reached at the College Creek facility via telephone (541-577-3254).

1. Procedures

- a. Steelhead removed from the trap will be placed in an anesthetic vessel provided by NPT containing a buffered solution of tricaine methanesulfonate (MS-222) at a concentration of 80 mg/L.
- b. After anesthetization, each fish will be examined for fin-clips, marks and/or tags, measured for fork length (cm), categorized by gender using secondary sexual characteristics, and origin determined (hatchery or natural).
- c. Scales will be collected from the preferred area of the fish and a 1ROP opercle punch given to each steelhead. The resulting tissue will be retained for future genetic analysis.
- d. Steelhead without a preexisting PIT tag will receive a tag for future detections on passover PIT tag arrays and downstream dams (kelts). These fish will be tagged in the cartilage of the pelvic girdle to facilitate tag retention during spawning.
- e. Steelhead will be recovered in fresh water. After recovery, steelhead will be released in the upstream release tube.

Downstream migrants – Chinook salmon, bull trout, and steelhead (kelts) may be observed on the upstream side of the weir. On-site staff will make efforts (within reason) to safely pass steelhead kelts and bull trout downstream, while restricting the downstream movement of Chinook salmon. NPT staff will be at the Imnaha weir to assist on a daily basis until June 20 or until it can be determined that adult steelhead will no longer be present at the weir site. In an emergency situation, NPT can also be reached at the College Creek facility via telephone (541-577-3254).

1. If bull trout and/or steelhead are observed on the upstream side of the weir, on-site staff (with assistance from NPT) will assess feasibility of passing fish downstream.
 - a. Criteria should depend on flows and presence of Chinook salmon. Chinook salmon that are passed above the weir are uniquely marked and abundance is managed. Therefore, all steps should be taken to avoid allowing Chinook salmon downstream.
 - b. On-site staff will monitor the relative presence of fish above the weir, and select the best time of day to pass fish without allowing salmon downstream.
2. Staff will use long-handled nets to herd steelhead and bull trout toward a selected weir panel.
3. The selected weir panel will be temporarily raised to allow bull trout and steelhead downstream.