Chapter 3: Chemical Residues in Rio Grande Silvery Minnows collected from the Middle Rio Grande, New Mexico, 2006-2008

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INTRODUCTION

The net accumulation of a chemical by an organism as a result of uptake from all environmental sources is termed bioaccumulation (USEPA 1995). Determining the extent of bioaccumulation in organisms is widely used as a method to monitor and assess pollutant distribution and bioavailability geographically and over time (Crawford and Luoma 1993; Schmitt et al. 1999). Phillips (1980), identified three benefits from using fish and other aquatic organisms in monitoring programs. First, concentrations of contaminants are often greater in tissue than in water and therefore, the probability of detecting trace amounts of contaminants in the environment is increased. Second, resident organisms provide a time-integrated assessment of a contaminant in question. Third, the direct bioavailability of contaminants that accumulate can be measured and temporally and spatially compared.

According to the critical body burden concept of McCarty (1986) and Escher and Hermens (2004), chemicals will have a specific effect (lethal or sublethal) upon fish once they have attained a specific internal tissue concentration. The internal concentration is independent of external influences and mode of exposure (i.e., in water or in the diet) and should occur at the same level regardless of the organism or species used. The objectives of this portion of the Rio Grande Silvery Minnow Health Study were to identify the body burden of chemicals in Rio Grande silvery minnow and review the extent of contamination, evaluate any possible health consequences associated with the concentrations, and provide baseline contaminant data. We analyzed Rio Grande silvery minnow carcasses (*Hybognathus amarus*; silvery minnows) for a variety of inorganic and organic chemicals, as well as lipid and moisture content. Organic chemicals analyzed included aliphatic hydrocarbons, polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs), dioxins and dibenzofurans, polybrominated diphenyl ethers (PBDEs), and a variety of organochlorine insecticides, including 1,1,1-trichloro-2,2-di(4-chlorophenyl)ethane, also known as DDT.

METHODS

Chemical Analyses, Quality Control and Methods of Interpretation of Contaminants in Fish

Lusk (2012) described the methods, dates, and locations of silvery minnow carcasses collected from six sites along the Rio Grande during 2006-2008. Study location and collection sites referred to in this report are described in Table 1 and depicted in Figure 1 and Figure 2. Silvery minnows that were dissected for fish health assessments (Davis and Lusk 2012) or pathogen assessments (Woodland 2012) were wrapped in aluminum foil or placed in polyethylene bags and frozen (4°Centigrade). Later, carcasses were removed from bags and foil and composited by site and date (up to 70 individual fish) in chemically clean jars and shipped frozen to the Service's contract laboratories for a variety of chemical analyses. These carcass composites (i.e., whole body minus the gonads or gills and some body fluids removed during their necropsy) were variously analyzed for 19 elements, methyl mercury, moisture and lipid contents, and 353 organic compounds including a variety of organochlorine (OC) pesticides, PCBs, PBDEs (flame retardants), dioxins, as well as aliphatic and aromatic hydrocarbons. A list of the chemicals and elements analyzed, abbreviations and names used, analytical methods used, and method detection limits for the chemical analyses, are provided in Table 2.

Briefly, silvery minnow carcass composites were homogenized and aliquot samples were digested and extracted for a variety of chemical analyses using gas chromatography, mass spectroscopy, atomic fluorescence or cold-vapor atomic absorption spectroscopy (Trace Element Research Laboratory 2010; Appendix 3A; and, TDI Brooks International, Inc., 2010; Appendix 3B). Percent lipid and percent moisture content were also determined. A single grab sample of water was also collected for analyses, and while the water and its results were not relevant to this study; however, the water sample provided for additional measures of laboratory quality control.

Quality-control measures included the analysis of method and procedural blanks (to measure contamination during sample preparation and analysis and to determine detection limits), duplicate samples (to measure precision), fortified (spiked) samples (to measure matrix interferences), and laboratory standards and certified reference material (to measure accuracy). Contaminants in silvery minnow carcasses are reported as micrograms per gram (µg/g; i.e., approximately parts per million) or micrograms per kilogram (µg/kg; i.e., approximately parts per billion) dry-weight and wet weight-weight concentrations; the latter was calculated from the moisture content of the sample.

Data Treatment and Statistics

Data were imported into spreadsheets (Microsoft Office Excel 2007, Microsoft Corp., Redmond, WA) or imported and analyzed using the statistical software Statistica (version 9.0, StatSoft, Tulsa, OK) or imported into a database using Microsoft Office Access 2007 (Microsoft Corp., Redmond, WA). Prior to final statistical analysis, the assumptions of normality and equal variance were formally tested by the Shapiro-Wilks test and Levene's test, respectively. Data that did not meet these assumptions were transformed to their natural logarithms and retested. If data were normally distributed comparisons were made using a one-way analysis of variance (ANOVA). Post-hoc comparisons between sites were conducted using a students' t-test (Bailey

1981) or Tukey's Honestly Significant Difference (HSD) test (for equal sample sizes) or Tukey-Spjotvoll/Stoline tests (for unequal sample sizes) (Statsoft Inc., 2010). If the natural log-transformation did not satisfy the assumptions, data were transformed to ranks and a Kruskal-Wallis and median test was applied to test the ranks (Statsoft Inc., 2010). Other parametric and nonparametric statistical tests were also employed and are so indicated in the text. For statistical purposes, as well as simplicity, when less than 25 percent of the analytical results were below the laboratory Method Detection Limit (MDL), the censored results were replaced with a value one-half the method detection limit prior to further statistical treatment. All censored data from comparison studies (e.g. Schmitt et al. 1999) were replaced with a value one-half the MDL (when reported) prior to comparison. Analytical results where chemicals were detected less frequently were only briefly summarized and results were provided in the appendices. Statistical significance probability (p) was set at $p < 0.05$ for all tests. Some of the fish tissue contaminant data were reported in either dry weight or wet weight concentrations and were so indicated. To convert dry weight concentrations into wet weight concentrations, the following equation was used:

Wet weight = (dry weight) * (1 - (percent moisture)/100)) Equation 1

Moisture content was determined by both inorganic and organic analytical laboratories. Values of moisture content conducted on same samples by the different laboratories were normalized and averaged. These average moisture contents were reported and evaluated. Only dry weight concentrations were reported by Mora (2001); therefore, a value of 72.1 percent moisture (average was derived using Schmitt et al. 1999) was used in Equation 1 to convert concentrations in lower Rio Grande fish into wet weight.

PCBs are a complex mixture of 209 isomers and congeners with 1 to 10 chlorines attached to the biphenyl structure in various arrangements with congeners identified by a suffix according to the nomenclature systems of Ballschmiter and Zell (1980) and Schultz and Malisch (1983). Aroclors are commercial PCB preparations that were produced up until 1977 by the Monsanto Chemical Company that contained various amounts of chlorine by weight. Total PCBs refers to the sum of the aroclors. Sum total PCBs refers to the summation of all PCB congeners and includes values where one-half the limit of detection (LOD) was substituted for PCB congener data below the LOD regardless of detection frequency. Certain PCB congeners are considered "dioxin-like" as they can have a fraction of the toxicity of dioxin (2,3,7,8-TCDD) and are often modified by their Toxic Equivalent Factors (TEFs; Van den Berg et al. 1998). TEFs from Van den Berg et al. (1998) for PCB congeners were used to calculate Toxicity Equivalents (TEQs) for fish. The TEQ scheme refers only to adverse effects (e.g., cancer) following interactions with the cellular Ah receptors (Wenning et al. 2011). For the purposes of the TEQ calculation, all values for dioxins, dibenzofurans, and PCB congeners below LOD were substituted with a zero.

Interpretation of Chemical Residues

Identification of contaminants of concern in whole body fish collected for this study was accomplished on a site or stream segment basis. Data from two sites each were combined into three groups (upper, middle, and lower) for further analysis at the segment scale. The evaluation methods included a comparison of the concentrations of chemicals in tissues collected from sites

or segments to various concentrations reported in the literature that affect fish (Jarvinen and Ankley 1999; USDI 1998; USACE 2010), to concentrations in fish collected from the Rio Grande or other watersheds (Roy et al. 1992; Simpson and Lusk 2000; Mora 2001; Abeyta and Lusk 2004; Schmitt et al. 2004; Buhl 2011), or to fish collected nationwide (Schmitt et al. 1999).

The significance of the concentrations of chemical contaminants in fish is not always clear, as elevated chemical concentrations are found in apparently healthy individuals. Identifying the presence of a chemical in silvery minnows was not considered sufficient information to conclude that the chemical would produce an adverse effect. The likelihood that a chemical substance in silvery minnows would produce an adverse effect is likely a function of the physical and chemical properties of the substance, the concentration of the chemical in the tissues, and the length of time the fish is exposed to the compound. Because environmental contaminants vary so widely in their potential to produce toxicity, comparison with an average literature-based contaminant-specific concentration was used to reach a determination regarding the potential for contaminant to produce adverse effects or identified the need for more research.

The Environmental Residue-Effects Database (USACE 2010 and references cited therein) were used for the majority tissue residue effects data comparisons. Emphasis was placed on contaminants that were known to pose serious health risks to fish or studies involving freshwater or marine fish in either carcasses or whole body fish in those studies cited in the USACE (2010) database. Those studies indicated concentrations above which a toxic effect due to a contaminant was likely to occur (i.e., adverse effects) including physiological, biochemical, growth, reproductive, and lethal effects. Those study also identify concentrations below which no adverse effects were likely to occur. For those cases where the concentrations in fish associated with adverse effects overlapped those concentrations where no adverse effects were identified, then comparisons were made to the average concentration associated with each type of effect (i.e., adverse or not). Where literature-based adverse effects thresholds were exceeded, then the corresponding adverse effect was further reviewed in relation to silvery minnow health.

Differentiating petrogenic from biogenic compounds in the interpretation of hydrocarbon residues in fish tissues is important. We compared the proportions of odd- and even-numbered long chain hydrocarbons to determine if sources were petrogenic (crude oil, diesel, light oil, etc.) as petrogenic compounds have approximately equal proportions of odd- and even-numbered long chain hydrocarbons (Hall and Coon 1988). We used ratios of pristane and phytane to n-heptadecane and n-octadecane, respectively, that when elevated indicate whether fish were recently or chronically exposed to petroleum compounds (Farrington et al. 1976; Anderson et al. 1978; Hall and Coon 1988).

RESULTS AND DISCUSSION

Quality Control of the Chemical Analyses

The U.S. Fish and Wildlife Service (USFWS) uses several contract laboratories to provide routine chemical analyses of chemical residues in biota and environmental samples (USFWS 2010a). The laboratories that conducted the chemical analyses of silvery minnow carcasses for this study were responsible for establishing the precision and accuracy of their analytical procedures. Quality assurance included cataloging silvery minnow sample collection, storage, and data transfer (Appendix 3C). Quality control procedures included analysis of procedural blanks, duplicate, and spiked samples as well as analysis of standard reference materials for inorganic chemicals (Appendices 3A and 3B). Data from such procedures were evaluated and documented by laboratory chemists and USFWS Analytical Control Facility (USFWS 2010a) and specific examples of quality control review are further described below.

Quality control described below was reviewed for the silvery minnow carcass samples only. Concentrations of mercury in the procedural blanks were slightly above the MDL in fish analyzed and reported by Trace Element Research Laboratory (2010; Appendix 3A). However, the concentrations of mercury detected in the procedural blanks (0.0028 µg/g dry weight) were within a factor of two of the MDL (0.002 µg/g dry weight) and were considered to be within the acceptable control limits. Concentrations of mercury in the procedural blanks were also 100 times less than the average concentration detected in silvery minnows and therefore were considered unlikely to affect interpretation. Concentrations in 10 of 41 duplicate element analyses of fish samples were below the MDL; thus no measure of the method precision was calculated. Quality control of analysis was maintained as the relative percent difference measured for the remaining 31 duplicate analyses averaged 6.1 percent with a range of 0 to 16.8 percent. Percent recovery of 41 spiked element analyses averaged 106 percent recovery with a range of 86 to 127 percent recovery of spiked amounts. Analysis of mercury in standard reference materials was within 87 to 107 percent of certified values, which indicated that mercury was accurately measured. Additional elements were analyzed in standard reference materials, and except for manganese (117 percent) and strontium (70.1 percent), the analysis of standard reference samples were within the acceptable range (80-110 percent) of certified values.

Concentrations of Aroclors (i.e., PCB-1242, PCB-1248, PCB-1254, PCB-1260) and PCB-total were detected in procedural blanks, but results were less than 10 times the MDL and were considered to be within the acceptable control limits (TDI–Brooks International, Inc. 2010; Appendix 3B). Concentrations in 91 of 224 duplicate analyses of fish samples were below the MDL and thus no measure of their method precision was calculated nor statistical analyses were conducted. The relative percent difference measured for the remaining 133 duplicate analyses averaged 6.3 percent with a range of 0 to 43.5 percent. Relatively high duplicate analysis results were found with biphenyl (43.5 percent) and heptachlor epoxide (29.1 percent); without these two duplicate analytes, the range of 131 duplicate analyses averaged 5.9 percent and ranged from 0 to 22.2 percent. Analysis of duplicate moisture content was 0 percent. Analysis of duplicate lipid content ranged from 0.7 to 12.6 percent and averaged 4.8 percent. All spike results for organic chemicals in silvery minnows were within normal limits (60 to 120 percent), except for

endosulfan II, n-heptacosane, and n-pentacosane, where the spike recoveries were low and therefore, the results of these three analytes may be biased low. As endosulfan II was not detected in greater than 75 percent of the samples analyzed, it was not further analyzed. Standard reference materials for organic chemicals were not analyzed as costs of analysis for over 350 organic chemicals in standard reference materials exceeded available funding.

Inorganic Contaminants, Methyl Mercury, and Lipid or Moisture Content

All analytical data on the concentrations of inorganic chemicals, methyl mercury or moisture content in silvery minnows are found in Appendix 3D and selectively summarized in Table 3. Nineteen inorganic contaminants were analyzed in carcass samples (Appendix 3D), however, molybdenum was below the MDL in all samples, and 4 elements (B, Be, Cd, and Cr) were detected in less than 75 percent of the samples analyzed. These 5 elements (B, Be, Cd, Cr, and Mo) were not compared statistically nor evaluated further. Concentrations of methyl mercury (MeHg) were also detected in all silvery minnows analyzed (Table 3). There were insufficient toxicity residue effects data available to assess the effects of iron, magnesium, manganese, and strontium in silvery minnows. The remaining inorganic contaminants in silvery minnows are discussed below. Average concentrations are provided below plus or minus one standard deviation (average±1SD).

A number of metals (Al, Cu, Fe, Mg, Mn, Ni, Pb, V, Zn) appeared to decrease in those fish collected on a downstream basis; however, this trend (or decrease) was not statistically significant (Table 3). Some of the differences in the metal contaminants in silvery minnows (Cu, Fe, Mg, Mn, Ni, Pb, V, and Zn) may partly be attributable to their ingestion of sediment (Buhl 2011). Silvery minnow feeding behavior from sediment substrates could increase their metal uptake by way of gill respiration or through dietary routes (Magaña 2009; Buhl 2011). Several of these elements were reported in underlying parent material (USGS 2008) and also generally decrease in a downstream direction. However, sediment samples reported by NMED (2009) collected near the same sites of fish collection did not show consistent or decreasing trends downstream for a number of elements (Cu, Fe, Mg, Mn, Ni, Pb, V, and Zn). Though three of these elements (As, Fe, and Mg) increased in water samples collected downstream (NMED 2009). Therefore, concentrations of these elements (Cu, Fe, Mg, Mn, Ni, Pb, V, Zn) in silvery minnows may reflect the general elemental composition of the geology around them, or reflect elevated sources, or reflect dietary exposures, which were not quantified by this study. Two elements (Cu and Ni) appeared to be elevated in silvery minnows collected near Site 2 (Table 3), though statistically insignificant in silvery minnows from other sites, this suggested there may be additional sources of copper and nickel near Site 2 as compared with other sites. Several heavy metals (Pb, Zn, Cu, Cr, As, Cd, and Ni) are common pollutants associated with urban stormwater (USEPA 1999), and Site 2 was below stormwater discharges from the Albuquerque urban area.

Aluminum

Aluminum concentrations in silvery minnow carcasses were elevated (82.3±75.0 µg/g wet weight), and although concentrations of aluminum generally decreased downstream, aluminum concentrations were not significantly different by site ($H=10.54$, $p=0.06$). Aluminum concentrations in silvery minnows (Table 4) were similar to aluminum concentrations in a

variety of fish collected from the Rio Grande by Roy et al. (1992; 45.3±60 µg/g wet weight), Mora (2001; 43.9±106.8 µg/g wet weight), or Schmitt et al. (2004; 16.1±11.6 µg/g wet weight), and to silvery minnows analyzed after 26 days accumulation in the Rio Grande (Buhl 2011; 15.1±7 µg/g wet weight). Aluminum concentrations in silvery minnows were similar to those in fish sampled from the San Juan River (Simpson and Lusk 1999; 152±193 µg/g wet weight).

Aluminum concentrations in whole fish associated with adverse effects (USACE 2010; average 32, range 8 to 77 µg/g wet weight) or with no effects (average 11, range 2 to 9 µg/g wet weight) ranged widely for freshwater fish (Cleveland et al. 1991; Handy 1993; Peterson et al. 1989). The majority of aluminum concentrations in silvery minnows were above the average concentration associated with adverse effects (32 µg/g wet weight). Interestingly, Poston (1991) showed no effects on growth, survival, or feeding in Atlantic salmon (*Salmo salar*) fed up to 200 µg/g wet weight dietary aluminum and salmon carcass concentrations reached 6 µg/g wet weight.

Sites that are acidic (i.e., with pH less than 7) and that have elevated aluminum in water can result in adverse effects to plants, invertebrates and fish (Sparling et al. 1997). Aluminum burdens in fish exposed to acidic conditions have been implicated in reduced growth and survival for both brook trout (Smith and Haines 1985) and smallmouth bass (Kane and Rabeni 1987). For example, adverse effects in Atlantic salmon occurred when aluminum concentrations of 20 µg/g wet weight were reached in fish when exposed to aluminum in water at a pH less than 5.0 (Peterson et al. 1989).

Aluminum is only sparingly soluble in water between pH 6 and pH 8 (ATSDR 2008). New Mexico Environment Department (NMED 2009) reported approximately 2 percent of aluminum was dissolved (0.1±0.2 mg/L) in the water column compared to the total concentration of aluminum (5.3±8.0 mg/L) in the Rio Grande. The minimum average pH in this study (Lusk 2012) was often slightly basic (pH>8) and few studies conducted have explored aluminum toxicity to fish in alkaline conditions. Considering that aluminum is often a gill toxicant and damage to the gill is greater in acidic water than in neutral or basic waters (Gensemer and Playle 1999), the toxicity residue relation for aluminum under acidic conditions in toxicity effect studies may not be representative of those under slightly basic conditions found in the Rio Grande.

It is also possible that a portion of the aluminum concentrations measured in silvery minnows may have come from ingested or sorbed sediment. Brumbaugh and Kane (1985) reported that whole-body aluminum concentrations in smallmouth bass (*Micropterus dolomieu*) were higher and more variable in fish analyzed with the gastrointestinal tract intact compared to those with the gut contents removed, presumably because of sediment in the gut.

Aluminum concentrations in silvery minnows as well as in other fish collected from the Rio Grande are at concentrations of concern and exceed adverse effect thresholds reported in literature-based studies associated with acidic conditions. Additional research should be conducted to determine if the elevated aluminum concentrations are associated with adverse effects in silvery minnows.

Arsenic

There was a significant difference in dry weight concentrations of (log transformed) arsenic (ANOVA, $F(5,27) = 4.60$, $p = 0.004$) in silvery minnows collected from among the six sites. Post hoc comparisons (using Tukey HSD test) indicated that average arsenic concentrations were highest in fish collected from Site 1 and Site 2 compared with those from Site 6 (Table 3). Arsenic concentrations in fish tissues often increased proportionately to increased dietary and water exposures and duration (Gilderhus 1966; Rankin and Dixon 1994; Buhl 2002). NMED (2009) reported arsenic dissolved in water samples collected near Site 1 (2.9±0.8 µg/L) and near Site 6 (4.4±1.3 µg/L). Between these two sites the average dissolved (log transformed) arsenic concentrations were statistically different ($t(25)=3.38$, $p=0.002$). Chapin and Dunbar (1994) reported that arsenic in the Rio Grande reflects the contributions of groundwater and the Jemez River, due to inputs of high-arsenic waters from the Jemez volcanic field. NMED (2009) reported arsenic concentrations in sediment samples collected near Site 1 (3.3±1.1 µg/g) or near Site 6 (4.0±2.2 µg/g), and while average arsenic concentrations generally increased downstream, they were not statistically different. This suggested that differential exposure to arsenic in water or sediment from these two sites might not reflect exposure and arsenic accumulation in silvery minnows collected from these two sites. Arsenic in silvery minnows may instead reflect dietary arsenic or other biological factors (Valette-Silver et al. 1999; Dutton and Fisher 2011).

Average arsenic concentrations in silvery minnow carcasses (0.9±0.1 µg/g wet weight, Table 4) were higher than average arsenic concentrations in a variety of fish collected from the Rio Grande by Roy et al. (1992; 0.2±0.5 µg/g wet weight), Abeyta and Lusk (2004; 0.2±0.2 µg/g wet weight), Schmitt et al. (2004; 0.2±0.1 µg/g wet weight), but similar to those in silvery minnows after 26 days accumulation (Buhl 2011; 0.6±0.2 µg/g wet weight). Average arsenic concentrations in silvery minnows were elevated compared to those in fish collected from the San Juan River (Simpson and Lusk 1999; 0.1±0.1 µg/g wet weight) and to those in fish collected nationwide (Schmitt et al. 1999; 0.2±0.3 µg/g wet weight). Depending on the form of arsenic in fish tissue, elevated arsenic in fish tissues from the Rio Grande may be appropriate for fish consumption advisories (Wilcox 1997). However, arsenic concentrations in silvery minnows were not considered high enough to represent a hazard to the fish itself, or piscivorous wildlife (Schmitt et al. 2004). For example, McGreachy and Dixon (1992) suggested a threshold for adverse effects of arsenic in fish occurred when concentrations exceeded 3 to 4 µg/g wet weight. Also, Gilderhus (1966) reported significant abnormal ovary and oocyte development occurred in adult fish at 1.7 µg/g wet weight, growth reduction occurred at 2.2 µg/g wet weight, and mortality occurred at 11.6 µg/g wet weight when compared with control fish. While elevated, arsenic in silvery minnows did not exceed levels of concern and likely reflected dietary sources.

Barium

There was a significant difference in the dry weight concentrations of barium (ANOVA, $F(5,27)=18.97$, $p=<0.05$) in silvery minnows collected from among the six sites. Post hoc comparisons (using Tukey HSD test) indicated that the average barium concentrations were highest at Site 1 and Site 2 and decreased significantly in fish collected downstream (Table 3). NMED (2009) reported barium concentrations in sediment samples collected near the upper sites (174±57 µg/g), near the middle sites (123±49 µg/g), or near the lower sites (170 ±49 µg/g); and

average concentrations were not statistically different. NMED (2009) reported barium concentrations in water samples collected near the upper sites (0.10±0.03 mg/L), near the middle sites (0.13±0.05 mg/L), or near the lower sites (0.18±0.14mg/L); and average concentrations were also not statistically different. Barium concentrations in silvery minnows did not reflect those in water or sediment, and also may reflect dietary concentrations. Barium is accumulated by several algal species, including diatoms and desmids (Havlik et al. 1980; Brook et al. 1988; Sedláček et al.1989). Diatoms are often part of the diet of larval and adult silvery minnows (Shirey 2004; Magaña 2009; USFWS 2010b).

Average barium concentrations in silvery minnows (15.5±4.8 µg/g wet weight,) appeared higher than average barium concentrations in a variety of fish collected from the Rio Grande by Roy et al. (1992; 2.9±2.5 µg/g wet weight,), Mora (2001; 3.5±5.4 µg/g wet weight) and Schmitt et al. (2004; 1.2±0.9 µg/g wet weight), but similar to those in silvery minnows after 26 days accumulation in the Rio Grande (8.7±3.6 µg/g wet weight) as reported by Buhl (2011). Barium concentrations in a variety of fish (other than silvery minnows) collected from the Rio Grande did not appear to differ from those in fish collected from the San Juan River (Simpson and Lusk 1999; 3.5±3.4 µg/g wet weight), from fish collected from the Colorado River (Hinck et al. 2006; 2.5±3.7 µg/g wet weight) or from fish collected nationwide (3.3±6.1 µg/g wet weight). Therefore, barium may also reflect a dietary exposure and accumulation in silvery minnows as compared with accumulation found in other fish collected from the Rio Grande and other watersheds. There were insufficient data available to assess the effects of elevated barium in silvery minnows. Additional research should be conducted to determine if elevated concentrations of barium in silvery minnows are associated with adverse effects.

Copper

Copper concentrations in whole fish associated with adverse effects (average 6.6, range 0.7 to 40 µg/g wet weight) or with no effects (average 5.1, range 0.02 to 16.3 µg/g wet weight) ranged widely and overlaps for freshwater fish (Handy 1992; Bonham et al. 1987; Jarvinen and Ankley 1999; Kamunde et al. 2005; Kolok et al. 2005). The majority of copper concentrations in silvery minnows (1.8±2.5 µg/g wet weight) were below the average concentration associated with adverse effects (6.6 µg/g wet weight). However, several high copper concentrations in silvery minnows collected from Site 1 and Site 2 (8.5-11.8 µg/g wet weight) were above this level of concern. Additional research should be conducted to determine if the elevated concentrations of copper are associated with adverse effects in silvery minnows at these sites.

Lead

Lead concentrations in whole fish associated with adverse effects (average 3.5, range 0.2 to 6.0 µg/g wet weight) or with no effects (average 1.3, range 0.2 to 6.0 µg/g wet weight) overlaps for freshwater fish (Hodson et al. 1978; Weber et al. 1991; Borgman et al. 1993). Lead concentrations in silvery minnows (0.1±0.1 µg/g wet weight) were below the average concentration associated with adverse effects (1.3 µg/g wet weight). However, silvery minnows collected in summer 2006 from Sites 1 through 4 had lead concentrations (>0.2 µg/g wet weight) greater than those found in trout associated with decreased iron concentrations in blood (Hodson et al. 1978). It was unclear why lead was elevated in silvery minnows during summer 2006, or

whether decreased iron may have occurred in silvery minnows that could reduce oxygenation and swimming performance (Hodson et al. 1978). Additional research could be conducted on the physiological effects of episodic or elevated lead concentrations in silvery minnows.

Mercury and Methyl Mercury

Methyl mercury averaged 88 (±10) percent of the total mercury in silvery minnows. While not statistically significant, mercury (Hg and MeHg) in silvery minnows (on a dry weight basis) appeared to increase downstream (Table 3). The highest concentrations of mercury (0.10 µg/g wet weight) were found in silvery minnows collected from Site 6. Wiener and Spry (1996) described concentrations between 0.01 and 0.1 µg/g wet weight as typical in fish collected from environments (without conditions such as low pH or warm temperature) that promote mercury accumulation. Beckvar et al. (2005) reviewed the available residue-effects literature and identified mercury concentrations less than 0.2 µg/g wet weight in whole body fish as protective. No silvery minnows exceeded this level of concern.

Nickel

Nickel concentrations in silvery minnows (range <0.1 to 3.6 µg/g wet weight) were highest in fish collected at Site 1 and Site 2 (Table 3). Few studies have been conducted evaluating the effects of nickel in whole fish (Jarvinen and Ankley 1999), but recently, Lapointe and Couture (2010) reported increased metabolic rates and enzyme activity in larval fathead minnow (*Pimephales promelas*) at whole body nickel concentrations greater than 2.6 µg/g wet weight. That is, elevated nickel exposure and accumulation increased energetic demand and metabolic costs in fathead minnows. While there may be differences between the species life histories, diet, contaminant exposure history, nickel concentrations measured in silvery minnows collected from Site 1 and Site 2 during summer 2006 suggest that they were at concentrations associated with increased metabolic rates in fathead minnows. Additional research should be conducted to determine the effects of elevated nickel concentrations in silvery minnows at upstream sites.

Selenium

While not statistically significant, selenium concentrations (on a dry weight basis) were elevated in silvery minnows collected near Site 5 (Table 3). Site 5 is below the confluence of the Rio Puerco, which drains a watershed that contains selenium-rich soils (Seiler et al. 1999). However, selenium concentrations in silvery minnows were less than the whole-body toxicity threshold of 4 µg/g dry weight advocated by Lemly (1996) and Hamilton (2002, 2003). Concentrations in silvery minnows were also lower than those linked with reduced growth (5.4 to 7.0 µg/g dry weight) in fathead minnows exposed to dietary selenium (Ogle and Knight 1989). Selenium concentrations in silvery minnows (0.5±0.1 µg/g wet weight) were similar to those in fish collected from the Rio Grande reported by Roy et al. (1992; 0.4±0.3 µg/g wet weight), Mora (2001; 0.3±0.2 µg/g wet weight), Abeyta and Lusk (2004; 0.8±0.7 µg/g wet weight), Schmitt et al. (2004; 0.7±0.4 µg/g wet weight), and in silvery minnows after 26 days accumulation (Buhl 2011; 0.8±0.2 µg/g wet weight), or in fish collected nationwide (0.5±0.5 µg/g wet weight).

Vanadium

Vanadium concentrations in silvery minnows ranged from 0.1 to 0.8 µg/g wet weight. No toxic threshold has been proposed for vanadium concentrations in fish, but Hilton and Bettger (1988) reported that a dietary toxic threshold to juvenile rainbow trout (*Oncorhynchus mykiss*). Reduced feeding and growth were seen in trout with carcass concentrations averaging 0.4 µg/g wet weight. At higher concentrations in trout (>5.3 µg/g wet weight), feed avoidance was increased, body weight was reduced, and mortalities were apparent (Hilton and Bettger 1988). Hilton and Bettger (1988) observed that the major physiological effect of vanadium in the trout was increased in vivo lipid oxidation. Silvery minnows from several upstream sites had vanadium concentrations above 0.4 µg/g wet weight when reduced feeding and growth were observed in trout. However, as the physiology of trout, a carnivore, and silvery minnow, an initial herbivore and later omnivore, may substantially differ, and there are few studies with which to compare, additional research should be conducted to determine whether growth or metabolic effects occur in silvery minnows associated with elevated vanadium residues.

Zinc

Average zinc concentrations in silvery minnows (38±6 µg/g wet weight) were similar to those in a variety of fish collected from the Rio Grande and reported by Roy et al. (1992; 33±16 µg/g wet weight), Schmitt et al. (2004; 39±24 µg/g wet weight), and in silvery minnows after 26 days accumulation (Buhl 2011; 50±18 µg/g wet weight), but higher than fish collected from the Rio Grande in Texas (Mora 2001; 13±13 µg/g wet weight). Zinc concentrations in a variety of other fish collected from the Rio Grande were similar to those in fish collected from the San Juan River (Simpson and Lusk 1999; 29±23 µg/g wet weight), in fish collected from the Colorado River (Hinck et al. 2006; 47±31 µg/g wet weight), and in fish collected nationwide (Schmitt et al. 1999; 39±24 µg/g wet weight).

Zinc concentrations in whole fish associated with adverse effects (average 58, range 5 to 220 µg/g wet weight) or with no effects (average 72, range 0.3 to 300 µg/g wet weight) ranged widely and overlapped in both marine and freshwater fish (Spehar 1976; Spehar et al. 1978; Farmer et al. 1979; Pierson 1981; Sherwood et al. 2000; Dube et al. 2005; USACE 2010). No zinc concentrations in silvery minnows were above the average concentration associated with adverse effect (58 µg/g wet weight). However, zinc concentrations in silvery minnows from all sites were higher than that (40 µg/g wet weight) associated with reduced growth in flagfish (*Jordanella floridae*) (Spehar 1976). Additional research should be conducted to determine if elevated concentrations of zinc are associated with reduced growth in silvery minnows.

Lipid and Moisture Content

Lipids are one of the main energy sources in fish. The stored energy available to a fish, as reflected in its body composition and energy reserves, may also influence its survival and reproduction (Breck 2008). Organic chemicals partition primarily into lipids within an organism and lipid content represents the capacity of an animal to accumulate those chemicals (Di Toro et al. 2000) as well as provides some resistance to chemical toxicity (Lassiter and Hallam 1990). However, when lipids are mobilized as an energy source, lipid-soluble chemicals tend to

partition from the lipid reserves into the bloodstream and increase the potential adverse effects on an organism (de Freitas and Norstrom 1974; Paterson et al. 2007). Lipid contents are often used to normalize organic pollutant concentrations, to reduce the variability within and between species and sexes, and normalization improves toxicity relationships for lipid soluble compounds when lipid and contaminant concentrations vary proportionally (Hebert and Keenleyside 1995).

Lipid content of silvery minnows ranged from 4.1 to 10.8 percent and averaged 6.9(±1.4) percent, but did not vary significantly by site or season or sample year (Table 5). However, lipid results may have been confounded by compositing fish of different size classes. Similar lipid content in fish collected from the Rio Grande was reported by Roy et al. (1992; 6.0±3.4 percent) and Schmitt et al. (2004; 4.5±2.9 percent) and in fish collected nationwide (Schmitt et al. 1999; 6.2±4.2 percent).

Moisture content in silvery minnows were not significantly different by site (Table 3). The average moisture content in silvery minnows in this study (71.9±1.9 percent) were similar to those in a variety of fish collected from the Rio Grande by Roy et al. (1992; 73.0±4.9 percent), Schmitt et al. (2004; 72.9±3.0 percent), and silvery minnows after 26 days accumulation (Buhl 2011; 73.0±1.7 percent), or in fish collected nationwide ((Schmitt et al. 1999; 72.1±4.5 percent).

A Pearson product-moment correlation coefficient was computed to assess the relationship between the moisture content and lipid content in silvery minnows. There was a significant negative correlation between the two variables ($r = -0.731$, $n = 27$, $p < 0.05$). This suggests that increases in moisture content are correlated with decreases in lipid content in silvery minnows and vice versa. Therefore, a relationship between lipid content and moisture content was modeled using linear regression analysis to predict lipid content:

Percent Lipid	= 78.6877 - (0.9945 * Percent Moisture)	Equation 2
Percent Moisture	= 45.2892 - (0.5366 * Percent Lipid)	Equation 3

If lipid content and moisture content are related as described by Equation 3, then a lipid content of 3 percent would translate to a moisture content of 78.8 percent; which would likely be the maximum moisture content of healthy silvery minnows. Mortalities occurred when lipid content was 3.2 percent or less in fish (Simpkins et al. 2003). Silvery minnows were not found to have lipid contents less than 3.2 percent.

Aliphatic and Aromatic Hydrocarbons

Analytical data for contaminant concentrations of organic chemicals and lipid content in silvery minnow carcasses are found in Appendix 3E and selectively summarized in Table 5. Three hundred and fifty-three organic contaminants were analyzed in carcass samples (Appendix 3E), however, 256 chemicals (73 percent) were detected in less than 75 percent of the samples analyzed, and therefore, those organic chemicals were not compared statistically nor evaluated further with the exception of n-undecane. However, all aliphatic hydrocarbons were used to help identify sources and interpret results. Several of the organic contaminants in silvery minnows are further discussed below.

Aliphatic Hydrocarbons

Aliphatic hydrocarbons (1.2±2.9 µg/g wet weight) were detected in all silvery minnows collected from five sites during January 2008 (Appendix 3E, Table 5). Differentiating petrogenic from biogenic compounds is important to demonstrate a pathway between the sources of hydrocarbons and their deposition in fish tissues. In general, the origin of organic matter can be obtained by analyzing the composition of aliphatic hydrocarbons, their peaks and ratios, or through the composition of the carbon preference index (Bray and Evans 1961). None of the hydrocarbons detected in silvery minnows had equal proportions of odd- and even-numbered aliphatic hydrocarbons indicative of petrogenic sources such as oil spills (Hall and Coon 1988). Also, all ratios of pristane-to-n-heptadecane or phytane-to-n-octadecane were low in silvery minnows, indicating no recent or chronic exposure to petroleum compounds in their diet (Anderson et al. 1978, Farrington et al. 1973, Hall and Coon 1988). Pristane-to-phytane ratios were also low (0.73±0.03), except at Site 1 (>1.1), which indicated that petrogenic aliphatic hydrocarbons were not those present in silvery minnows. Petrogenic hydrocarbons have much lower pristane-to-phytane ratios ranging from 0.14 to 0.36 (Roushdy et al. 2010). Pristane-to-phytane ratios (approximately 0.8) are indicative of carbonate-dominated habitats (ten Haven et al. 1987).

The presence of aliphatic hydrocarbons with a maximum of 20 to 35 carbons and with dominance of odd numbers of carbon atoms can be used to indicate the prevalence of higher land plant waxes though source identification is not always certain (Tornabene 1980**;** Cranwell 1984; Jeng and Huh 2008). By using the presence of higher chain hydrocarbons, we estimated that 34 and 39 percent of the aliphatic hydrocarbons in silvery minnows at Site 3 and Site 4 were from terrestrial plant sources. Up to 12 percent of the aliphatic hydrocarbons in silvery minnows at Site 1 were from terrestrial plant sources; whereas, silvery minnows from Site 2 and Site 6 contained less than 5 percent of aliphatic hydrocarbons of terrestrial plant origin.

The contribution of phytoplankton and diatoms in silvery minnows can be difficult to recognize due to the low abundance of aliphatic hydrocarbons in certain species of algae (Tornabene 1980). However, significant amounts of 15, 19, and particularly 17 carbon chain aliphatic hydrocarbons, has been interpreted as an indicator of hydrocarbons of algal origin (Cranwell 1984; Jeng and Huh 2008). Using these carbon chain aliphatics as indicators, silvery minnows collected in January 2008 at Site 2 and Site 6 contained approximately 92 and 91 percent of aliphatic hydrocarbons from algal origin; whereas silvery minnows at Site 1 contained 28 percent; and, silvery minnows at Site 3 and Site 4 contained approximately 60 percent of aliphatic hydrocarbons from algal origin. These hydrocarbon indicators suggested a wide variety of plant materials can be present in silvery minnows at different locations and during different seasons.

Silvery minnows in January 2008 collected from Sites 2 though 6 contained 5 percent (or less) of petrogenic aliphatic hydrocarbons (that is, they had few aliphatic chains with less than 15 carbons associated with oil, gasoline, or diesel fuels) (Neff 1979). However, silvery minnows collected at Site 1 contained an inordinate amount of n-undecane (18.5 µg/g wet weight) with over 58 percent of their hydrocarbon burden being indicative of petrogenic origin. This is the highest concentration of n-undecane ever reported in freshwater fish. N-undecane is indicative of diesel range organic compounds such as kerosene-based fuels, solvents, paints, and soot

(Technical Resources International, Inc. 2003), perhaps from idling traffic on the 550 Bridge above Site 1, or from other unknown sources nearby.

Aromatic Hydrocarbons

A variety of aromatic hydrocarbons were detected in silvery minnows (total PAHs were 66±11 µg/kg wet weight) from five sites collected in January 2008 (Table 5, Appendix 3E). PAHs are found everywhere in the environment and are formed by burning organic matter (Neff 1979; Eisler 1987). PAHs are commonly found in used motor oil, exhaust from internal combustion engines and emissions from fossil fuel powered energy plants, grilled foods, forest fires, and volcanoes, as well as most products made from coal and petroleum (National Research Council 1983; Eisler 1987). PAH concentrations in freshwater varies widely, depending upon such factors as proximity of the waterbody to the source, source type, and season. PAH concentrations in fish depend upon their proximity to the source of pollution, the physical and chemical properties of the PAH molecule, environmental variables, the PAH content of the diet, and biological factors (Moore and Ramamoorthy 1984; Eisler 1987; McElroy et al. 1989).

Several PAHs, especially those containing 4 to 6 aromatic rings in their structure, have been shown to be mutagenic, carcinogenic, and inducers of tumors in animals exposed to high doses of PAHs in the laboratory. Studies directly linking PAHs to these effects in wild fish are few in number (Albers 2003). From studies conducted in the Great Lakes basin, the International Joint Commission recommended that concentrations of the PAH benzo[a]pyrene not exceed 1.0 µg/g in fish. Average PAH concentrations in silvery minnows (1.5±2.6 µg/kg wet weight) were nearly 10 times lower than this threshold of concern. Concentrations of PAHs in silvery minnows tended to reflect the patterns of PAHs in sediment samples (NMED 2009) with higher concentrations detected at the middle sites near Albuquerque, New Mexico (Table 5).

Certain PAHs, particularly naphthalene and its alkylated derivatives, such as 1-methyl-naphthalene, 2-methylnaphthalene, 2,6-dimethylnaphthalene, C1-methylnaphthalene, C2-naphthalene, C3-methylnaphthalene, C4-methylnaphthalene, but also phenanthrene, fluor-anthene, and fluorene, were relatively elevated in silvery minnow carcasses compared to the other PAHs (Table 5, Appendix 3E). Many of these PAHs either components of weathered oil or were combustion-related hydrocarbons. For example, combustion related hydrocarbons such as phenanthrene and its derivatives averaged 2.7 percent, fluoranthene and its derivatives averaged 1.4 percent, chrysene and its derivatives averaged 0.7 percent, and pyrene and its derivatives average 0.6 percent of the total PAHs measured in silvery minnows. Naphthalenes and its derivatives were the highest percentage (average 6 percent) of the total PAHs measured in silvery minnows. Biphenyl occurs in weathered coal tar, crude oil, and natural gas and averaged 4 percent of the total PAHs measured in silvery minnows.

Naphthalene and its alkylated derivatives were consistently detected in silvery minnows (15±2 µg/kg wet weight) (Table 5). Naphthalenes are not generally reported in fish, but were detected at concentrations below 3 µg/kg wet weight in fish from the Pacific Coast (Easton et al. 2002). Naphthalene is a white solid with a strong smell and is also called mothballs. Naphthalene is a component of fossil fuels such as gasoline, diesel, oils and coal; it is also formed when natural products such as wood or tobacco are burned. Naphthalene has a short half life in water.

Naphthalene derivatives were detected at low concentrations in sediment samples collected by the NMED (2009) from Site 3 (5.6 µg/kg wet weight) and were less than naphthalene sediment toxicity thresholds (Buchman 2008) . Notably, the pattern of accumulation of naphthalene and its derivatives was similar in silvery minnows collected along the Rio Grande suggesting exposure to widespread atmospheric sources of PAHs (Table 5).

High concentrations of PAHs in fish have been associated with decreases in health measures, increased lesions, increased genetic alterations, and altered populations (Albers 2003). PAHs in fish tissues from uncontaminated areas were generally less than 10 µg/kg wet weight and from contaminated areas were generally greater than 100 µg/kg wet weight (Eisler 1987). PAHs in fish are usually low because fish rapidly metabolize PAHs or excrete them, though there is a metabolic cost associated with the increased enzyme activity (Albers 2003; Colavecchia et al. 2004). Therefore, silvery minnows were considered moderately contaminated with PAHs (average total PAHs 66±11 µg/kg wet weight). However, effects of PAHs in silvery minnows are difficult to generalize as the incidence of lesions, effects of any genetic alterations and metabolic pathways of PAHs in fish varies by species (Albers 2003). Additionally, few studies have been conducted on wild fish (Albers 2003). Additional research should be conducted to determine the growth effects or lesions associated with elevated PAHs in silvery minnows.

Organochlorine (OC) Pesticides, Dioxins, PCBs, and PBDEs

A variety of OC pesticides (total OCs 54±26 µg/kg wet weight) were detected in silvery minnows collected variously from all six sites during 2006 to 2008 (Table 5, Appendix 3E). The DDT metabolite, p,p'-DDE, accounted for the majority (43±14 percent) of the sum total of the OC pesticides detected in silvery minnows. Trans-nonachlor (11±4 percent), alpha chlordane (9±4 percent), and endosulfan sulfate (6±2 percent) were also detected at greater concentrations than the remainder of the OC pesticides (<1 to 5 percent) and are discussed below.

Total DDTs

DDT became widely used in the 1940s and was extremely effective controlling pest vectors until it was banned by the USEPA in January 1973 due to its long residual life in the environment and its accumulation in food chains (Beckvar and Lotufo 2011). The term total DDT refers to the sum of DDT and its metabolites (DDE, DDD, and DDMU), including the ortho para (o,p' or 2, 4) and para para (p,p' or 4, 4) isomers. Total DDT in silvery minnow carcasses was mostly DDE (83 percent), DDD (12 percent) and DDMU (3 percent). However, approximately 2 percent of the parent chemical (DDT) was still detected suggesting use of DDT in the environment and transport to the Rio Grande watershed has not ceased. Similar proportions of DDT (8 percent) and its metabolites DDE (80 percent) and DDD (13 percent) were reported in fathead minnow by Jarvinen et al. (1976).

Average concentrations of total DDT in silvery minnow carcasses (18±7 µg/kg wet weight) were similar to those found in fish sampled throughout the Rio Grande (Schmitt et al. 2004; 43±16 µg/kg wet weight), but generally less than those found in fish sampled nationwide (Schmitt et al. 1999; 240±800 µg/kg wet weight). No significant linear relationships was found between the concentrations of total DDT in silvery minnows and their lipid content. An ANOVA tests found

significant differences in wet weight concentrations of total DDT ($F(2,9)=14.2$, $p=0.002$) in silvery minnows collected from the middle sites (Site 3 and Site 4) than in those collected from the upper (Site 1 and Site 2) or lower sites (Site 5 and Site 6). As there are no known sources of total DDT near Site 3 and Site 4, it is possible that the rate of bioaccumulation or diet of silvery minnows is greater in this section of the river. Concentrations of total DDT in silvery minnows were below the lethal effects threshold of 600 µg/kg wet weight reported by Beckvar and Lofuto (2011). However, recent research have focused on endocrine disruption and other sublethal effects that have been observed at concentrations of total DDT similar to and up to two orders of magnitude lower than in lethal studies with older fish life stages (Beckvar and Lofuto 2011). Therefore, additional research could be conducted to determine if total DDT detected in silvery minnows are associated with any adverse sublethal effects.

Endosulfan

Designed in the 1950s, endosulfan is an organochlorine insecticide that can be used on a wide variety of vegetables, nuts, fruits, cotton, and ornamental plants. Endosulfan is a member of the class of the cyclodiene pesticides and consists of two isomers (endosulfan I and II), and a sulfur metabolite (Beckvar and Lotufo 2011). Endosulfan was recently banned in the United States due to continued risk to farm workers, wildlife, fish quality, and the environment, with a complete phase out of agricultural use of the chemical by 2016 (USEPA 2010).

Concentrations of endosulfan sulfate in silvery minnow carcasses (2.4±1.5 µg/kg wet weight) were higher than endosulfan concentrations in larger-bodied fish (< 0.2 µg/kg wet weight) collected from the Rio Grande by NMED (2009). No significant linear relationship was found between concentrations of endosulfan in silvery minnows and their lipid content. There was a significant positive relationship ($r = 0.668$, $n = 10$, $p = 0.017$) between concentrations of endosulfan sulfate in silvery minnows and their average weight in composite samples. However, average weights of the composited fish were not significantly different by site or by section of river, even though some of the larger fish were in collections of fish from Site 3 and Site 4. An ANOVA test found significant differences in carcass wet weight concentrations of endosulfan sulfate ($F(2,9)=13.6$, $p=0.001$) collected from the middle sites (Site 3 and Site 4) than in those collected from the upper or lower sites (Table 5). The NMED (2009) sampled endosulfan in sediments collected from the Rio Grande during 2006 to 2008, and three of the highest concentrations (0.05 µg/g wet weight) were found in the Rio Grande near Site 3. Reasons for elevated endosulfan sulfate in silvery minnows from the middle sites are unclear. There were elevated sediment concentrations of endosulfan sulfate near Site 3, but also bioaccumulation rates or diets in silvery minnows from the middle sites may differ in this river section.

Several fish species experienced mortality associated with whole body concentrations of endosulfan sulfate as low 31 µg/kg wet weight (Beckvar and Lotufo 2011). Morphological changes in the liver and enzyme activity reductions in brain, liver, and skeletal muscle were reported in fish at concentrations as low as 1 µg/kg wet weight, becoming more severe at concentrations greater than 5 µg/kg wet weight (European Food Safety Authority 2011). As concentrations of endosulfan and endosulfan sulfate in silvery minnows exceeded levels of concern, particularly in those collected from Site 3 and Site 4, additional research should be conducted to determine the effects of elevated concentrations in silvery minnows.

Trans-nonachlor

Trans-nonachlor is the common name for 1,2,3,4,5,6,7,8,8-nona-chloro-3a,4,7,7a-tetrahydro-4,7-methanoindan, and it is a member of the class of the cyclodiene pesticides (e.g., "technical chlordane") that were often used for management of subterranean termites (Isoptera). Chlordane mixtures (including alpha chlordane) were first produced in 1948, though commercial uses were later banned (1988) in the U.S.A. due to concerns about the risk of cancer (USEPA 2004).

Trans-nonachlor is the most bioaccumulative component of chlordane (USEPA 2004). Average concentrations of trans-nonachlor in silvery minnow carcasses (5±3 µg/kg wet weight) were similar to those found in fish sampled throughout the Rio Grande (Schmitt et al. 2004; 10±13 µg/kg wet weight), but generally less than those found in fish sampled nationwide (Schmitt et al. 1999; 39±102 µg/kg wet weight). No significant linear relationships was found between concentrations of trans-nonachlor in silvery minnows and their lipid content. An ANOVA test found significant differences in wet weight concentrations of trans-nonachlor ($F(2,9)=24.6$, $p<0.001$) in silvery minnows collected from the middle sites (Site 3 and Site 4) than in those collected from the upper sites (Table 5). Unless there are sources of trans-nonachlor (and many other organic chemicals) in the vicinity of Site 3 and Site 4, it is likely that bioaccumulation rate in fish at these sites was greater in this section of the river. Concentrations of trans-nonachlor in silvery minnows do not exceed levels of concern (Eisler 1990; Beckvar and Lotufo 2011).

PCBs

PCBs do not occur naturally in the environment. PCBs have been used as hydraulic lubricants, insulators, heat transfer fluids, dielectric fluid for transformers and capacitors, pesticide extenders, dust-reducing agents, flame retardants, sealants, and organic diluents (Hutzinger et al. 1974). PCBs accumulate from air, water, and in animals through the food web because they are highly lipid-soluble and persistent in the environment. PCBs were introduced in the U.S.A. in 1929 and certain uses were banned in 1979 (USEPA 1979). PCBs have been shown to adversely affect reproduction, thymic atrophy, enzyme induction, nervous systems dysfunction, behavioral abnormalities, liver injury, estrogenic activity, endocrine disruption, immune system suppression, deformities, hepatotoxicity, and tumor promotion in fish, wildlife, and experimental animals, and it is toxic to people (Eisler and Belisle 1996; Hoffman et al. 1996; Niimi 1996; ATSDR 2000; NMED 2009; Wenning et al. 2011).

Average total PCB concentrations in silvery minnows (15.3±10.1 µg/kg wet weight) were higher than average concentrations in channel catfish (*Ictalurus punctatus*) fillets (4.6±5.3 µg/kg wet weight), or in white bass (*Morone chrysops*) fillets from Cochiti Reservoir (Gonzales and Fresquez 2006), but less than average total PCB concentrations in whole body carp (90.7±13.0 µg/kg wet weight) or in white bass fillets (177 µg/kg wet weight) collected from the Rio Grande (NMED 2009, New Mexico Water Quality Control Commission 2010). Average concentrations of total PCBs in silvery minnows were in the low range of average total PCB concentrations reported in whole body cyprinids (range 18-320 µg/kg wet weight) collected across the U.S.A. (Stahl et al. 2009). Interestingly, the pattern and ratios of the different PCB congeners in silvery minnows were generally consistent between sites, but varied by sample year, suggesting PCB

congener patterns may vary by season, reflect atmospheric deposition patterns and sources, or reflect different rates of bioaccumulation, diet, or elimination processes in these fish.

Total PCB concentrations in whole fish associated with adverse effects (average 118, range 0.1 to 1502 µg/g wet weight) or with no effects (average 431, range 0.1 to 4668 µg/g wet weight) ranged widely and overlapped for marine and freshwater fish (USACE 2010; Wenning et al. 2011). No total PCB concentrations in silvery minnows were above the average concentration associated with adverse effect (118 µg/g wet weight). However, effect studies using total PCB concentrations ranged so widely as to make comparisons with concentrations in silvery minnows unsatisfactory. For example, toxicity studies for PCB-153 and PCB-180 with effects in common carp (*Cyprinus carpio*) were 0.04 µg/g wet weight and 0.06 µg/g wet weight, respectively, while the same two PCB congeners have higher effect concentrations for fathead minnow, ranging from 6,030,000 and 4,565,000 µg/g wet weight, respectively. Concentrations of PCB-153/163 and PCB-180/193 in silvery minnows were 1.4±0.9 µg/g wet weight and 1.3±0.9 µg/g wet weight, respectively, and therefore, exceed the effect studies involving carp but do not exceed the no effect studies involving fathead minnow.

During a review of the PCB toxicity thresholds for endangered fish, Meador (2000) derived a concentration associated with adverse effects at 2400 µg/kg dry weight on a per lipid basis. Average total PCB concentrations in silvery minnows (838±499 µg/kg dry weight on a per lipid basis) were below this threshold. Also, Olsson et al. (1999) reported skeletal deformities including craniofacial malformations associated with fish injected with 360 µg/kg PCB-190 wet weight on a per lipid basis. Silvery minnows had PCB-190 concentrations lower than 36 µg/kg wet weight on a per lipid basis. This suggests that PCB-induced deformity would be unlikely in the silvery minnows unless they were more sensitive than the zebrafish (*Danio rerio*) used by Olsson et al. (1999), therefore, PCB-90 is unlikely the cause of the deformed opercula commonly observed in silvery minnows (Davis and Lusk 2012). However, given the wide range of effect concentrations in fish, additional research could be conducted to determine if individual PCB congener concentrations in silvery minnows are associated with adverse sublethal effects.

Dioxins, Dibenzofurans, and TEQs

Certain PCB congeners are able to line up in a flat configuration and are considered "dioxin-like" as they may have a fraction of toxic equivalency (TEQ) to dioxin (i.e., 2,3,7,8-TCDD) (Wenning et al. 2011). Dioxins and dibenzofurans were not detected above the LOD in silvery minnows. Toxic equivalency factors (TEFs) from Van den Berg et al. (1998) were used for certain PCB congeners (PCB-105, PCB-118, PCB-156/157, and PCB-167) in silvery minnows to calculate TEQs. TEQs for silvery minnows ranged from 0.08 picograms TCDD per gram (pg /g) wet weight in silver minnows collected from Site 6 to 1.2 pg/g wet weight in fish collected from Site 3. TEQs in silvery minnows averaged 0.3±0.3 pg/g wet weight and averaged 4.3±4.1 pg/g wet weight on a per lipid basis. TEQs in silvery minnows were below those reported by Gonzales and Fresquez (2006) in fillets from fish collected from Abiquiu and Cochiti Reservoirs (range 3 to 300 pg/g wet weight), in fish collected from the Rio Grande (Schmitt et al. 2004; 1.8±1.2 pg/g), or in large bodied fish collected from the Rio Grande (NMED 2009; 2.1±0.9 pg/g wet weight). Steevens et al. (2005) proposed no effect and low effect tissue residue benchmarks from 57 to 700 pg /g wet weight on a lipid basis to protect early life stage fish. Concentrations in

silvery minnows would fall well below that range of potential effects in fish. Therefore, dioxin toxicity or its toxic equivalence using PCB congeners is not expected in silvery minnows at the tissue concentrations detected.

PBDEs

Total BDE in silvery minnows carcasses averaged 138±142 µg/kg wet weight and ranged from 11 µg/kg wet weight at Site 1 to 477 µg/kg wet weight at Site 4 (Table 5, Appendix 3E). On average, 76 percent of total BDE in silvery minnow carcasses was BDE-47, followed by BDE-100, BDE-154, BDE-49/71 and BDE-28, at 9, 3, 2.5 and 2 percent, respectively, similar to BDE congener patterns detected in other feral fish (Wenning et al. 2011). Patterns of the relative ratios of BDEs were similar in fish by site within a sample year than between sample years. For example, BDE-206 was not detected in silvery minnows collected in 2006, but was detected in silvery minnows collected during 2008 (Appendix 3E).

Little information is available, at present, on the toxicity of PBDEs in fish (Wenning et al. 2011). BDE-47 concentrations in whole fish associated with adverse effects (average 180, range 70 to 410 µg/g wet weight) or with no effects (average 140, range 3 to 410 µg/g wet weight) ranged widely and overlapped for freshwater fish (Chen et al. 2010, Chou et al. 2010, USACE 2010). BDE-47 in silvery minnow carcasses averaged 110±117 µg/kg wet weight and ranged from 6 µg/kg wet weight at Site 1 to 385 µg/kg wet weight at Site 4 (Appendix 3E, Table 5). None of the BDE-47 concentrations in silvery minnows were above the average concentration associated with adverse effect (180 µg/g wet weight). In a fathead minnow study, Muirhead et al. (2006) reported that egg laying in breeding pairs stopped after accumulation of BDE-47 in 10 days and cessation of spawning appeared to have occurred when peak body levels of 15 µg/g wet weight were reached in male fathead minnow. BDE-47 was not elevated above these thresholds of concern in silvery minnows. However, additional research on the effects of BDEs is warranted as there are few studies on the effects of PBDEs in fish. Marr et al. (2010) described research into the effects of BDE-47 on silvery minnows being conducted by the U.S. Geological Survey's Columbia Environmental Research Center.

CONCLUSIONS

Concentrations of inorganic contaminants in silvery minnows were similar to those in other fish species collected from the Rio Grande and were generally low, except for aluminum and zinc. Aluminum and zinc were moderately elevated in silvery minnows throughout the Middle Rio Grande, but were similar to concentrations in fish from other watersheds in New Mexico. Several elements, arsenic and barium, decreased in silvery minnows in a downstream manner, though concentrations of these elements in water or sediment did not also decrease downstream. Certain elements, including copper, lead, and nickel, were elevated in silvery minnows collected from the upper sites above levels of concern for these elements in laboratory studies involving sublethal effects. Research into the sublethal effects of aluminum, copper, lead, nickel, and zinc on silvery minnows is warranted.

Generally, organic contaminant concentrations in silvery minnows were low, except for PAHs. Certain PAHs, mostly naphthalenes, were moderately elevated in silvery minnows sampled throughout the Middle Rio Grande. One aliphatic hydrocarbon, n-undecane, common in fuels and diesel soot, was also elevated in silvery minnows collected at Site 1, the Rio Grande at Bernalillo, New Mexico. It was unclear why concentrations of several organochlorine pesticides, PCBs, and PBDEs, were elevated in silvery minnows collected from Site 3 and Site 4. Flame retardant chemicals, such as PBDEs used in furniture foam, were higher in silvery minnows than were total DDTs or total PCBs, but were below levels of concern compared with laboratory studies. Research into the sublethal effects of PAHs on silvery minnows is warranted.

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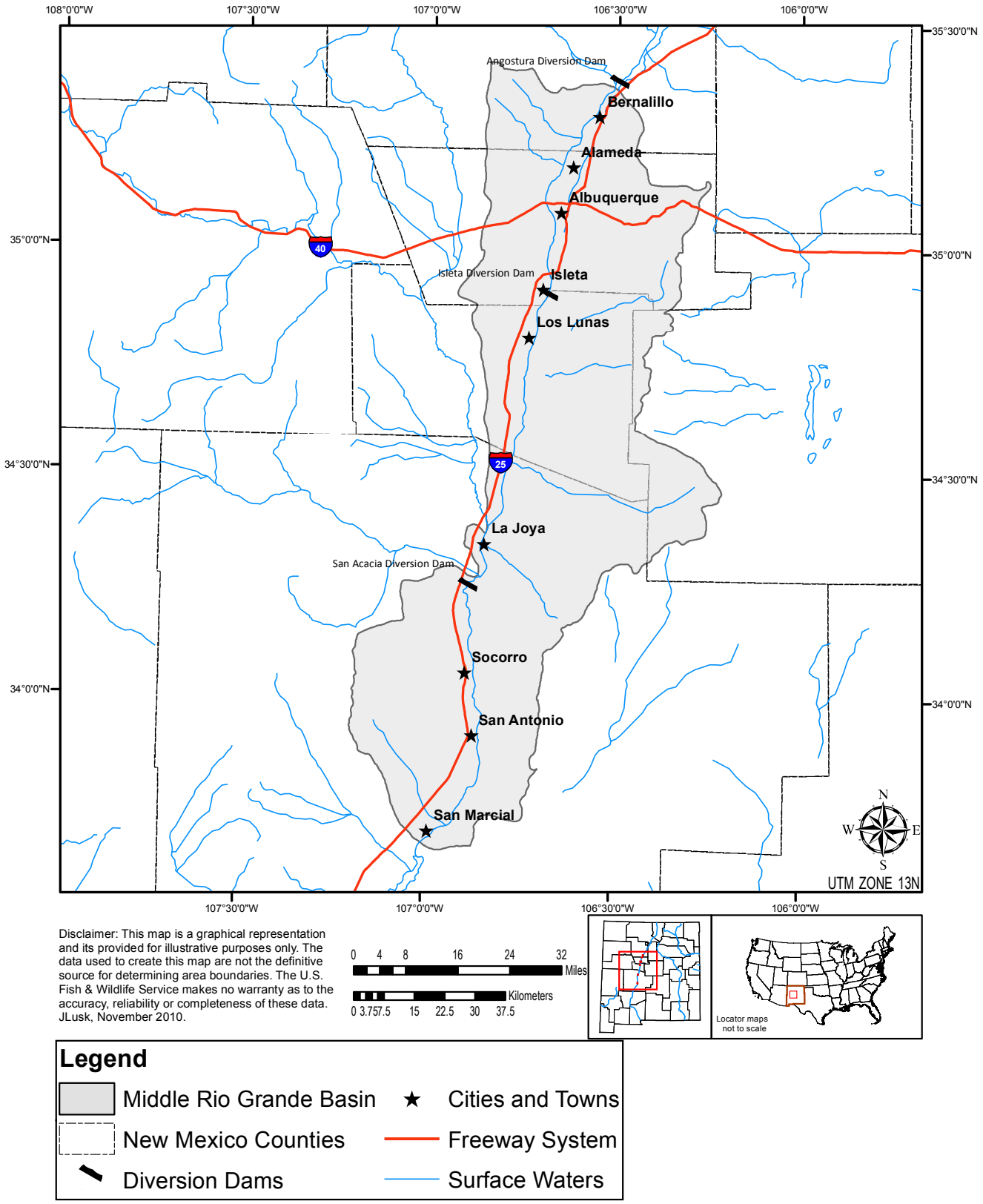

Figure 1. Location of the Middle Rio Grande Watershed and RGSM Health Study Area in New Mexico.

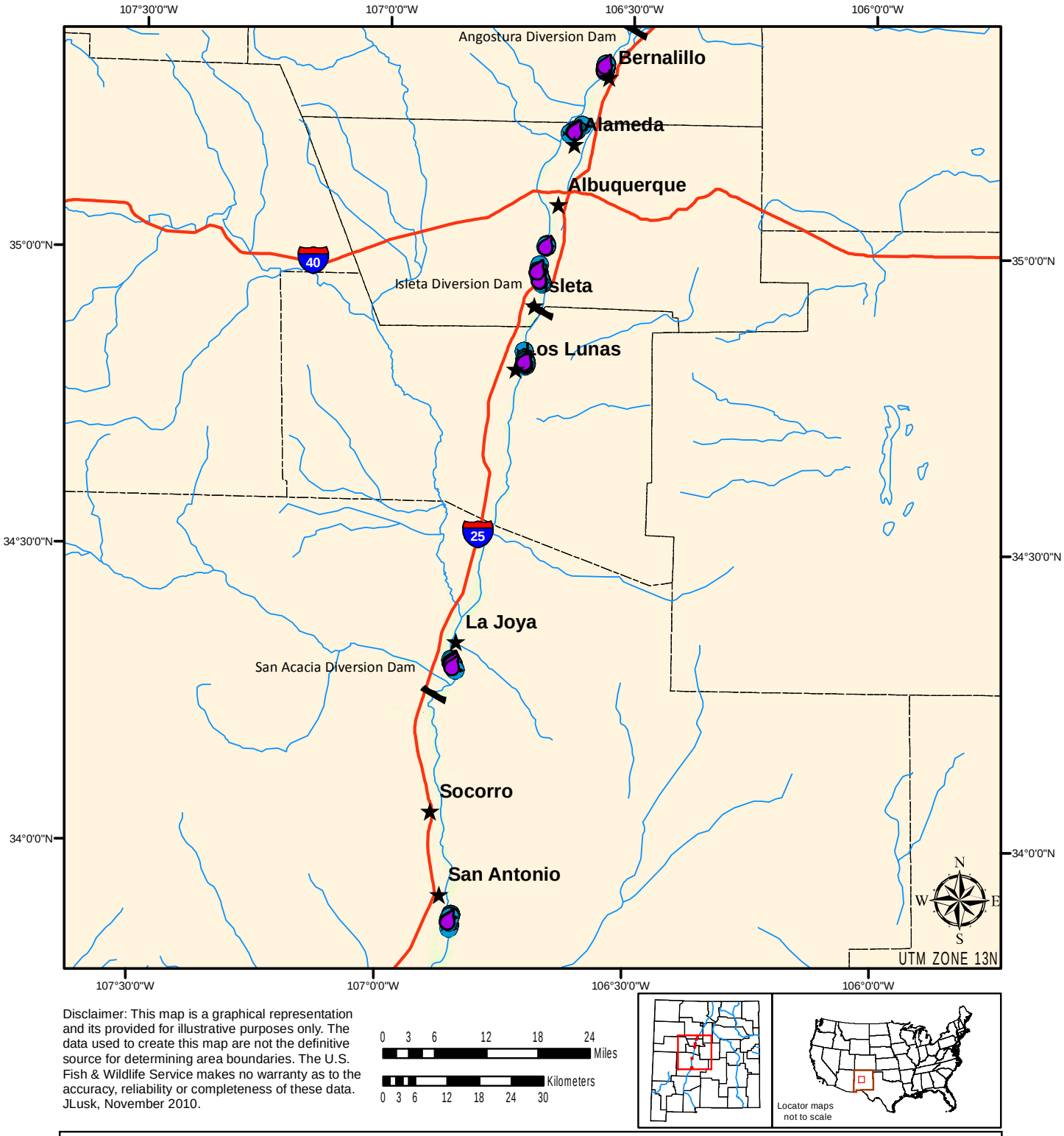

Figure 2. Location of the Six RGSM Health Study Sites along the Middle Rio Grande in New Mexico.

Table 1. Site number, site name name, latitude and longitude (in decimal degress) of a central point, and description of the RGSM Health Study sampling sites.

Site Number	Site Name	Latitude and Longitude (central point - decimal degrees)	Site Location Description
1	Rio Grande at Bernalillo, New Mexico	35.3181, -106.5589	approximately 0.1 mi upstream to 0.8 mi downstream of the Highway 550 Bridge crossing
2	Rio Grande at Alameda, New Mexico	35.2101, -106.6172	approximately 0.7 mi upstream and 1.1 mi downstream of AMAFCA North Diversion Channel mouth
3	Rio Grande at Los Padillas, New Mexico	34.9716, -106.6897	Just north of the Interstate 25 Bridge to approximately 5 mi upstream
4	Rio Grande at Los Lunas, New Mexico	34.8228, -106.7149	approximately 0.6 mi to 2.4 mi north of the Main Street (Highway 6) Bridge crossing
5	Rio Grande at La Joya, New Mexico	34.3073, -106.8493	approximately 2.4 mi to 3.8 mi north of the confluence with the Rio Salado
6	Rio Grande near San Antonio, New Mexico	33.8775, -106.8495	approximately 0.5 mi downstream and 1.2 mi upstream of Bosque del Apache National Wildlife Refuge (BdANWR) North Boundary

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Chemical Grouping	Analytical Method[1]	Limit of Detection[2]	Chemical Name or Abbreviation[3]	CAS[4] Number	Alternative Chemical Name
Aromatic hydrocarbon	GCMS	<0.45	1,6,7-trimethylnaphthalene	2131386	1,6,7-trimethylnaphthalene
Aromatic hydrocarbon	GCMS	<0.20	1-methylnaphthalene	90120	1-methylnaphthalene
Aromatic hydrocarbon	GCMS	<0.51	1-methylphenanthrene	832699	1-methylphenanthrene
Aromatic hydrocarbon	GCMS	<0.64	2,6-dimethylnaphthalene	28804888	2,6-dimethylnaphthalene
Aromatic hydrocarbon	GCMS	<0.20	2-methylnaphthalene	91576	2-methylnaphthalene
Aromatic hydrocarbon	GCMS	<0.24	acenaphthalene	208968	acenaphthalene
Aromatic hydrocarbon	GCMS	<0.41	acenaphthene	83329	acenaphthene
Aromatic hydrocarbon	GCMS	<0.20	anthracene	120127	anthracene
Aromatic hydrocarbon	GCMS	<0.37	benzo(a)anthracene	56553	benzo(a)anthracene
Aromatic hydrocarbon	GCMS	<0.45	benzo(a)pyrene	50328	benzo(a)pyrene
Aromatic hydrocarbon	GCMS	<0.32	benzo(b)fluoranthene	205992	benzo(b)fluoranthene
Aromatic hydrocarbon	GCMS	<0.46	benzo(e)pyrene	192972	benzo(e)pyrene
Aromatic hydrocarbon	GCMS	<0.53	benzo(g,h,i)perylene	191242	benzo(g,h,i)perylene
Aromatic hydrocarbon	GCMS	<0.41	benzo(k)fluoranthene	207089	benzo(k)fluoranthene
Aromatic hydrocarbon	GCMS	<0.31	biphenyl	92524	phenylbenzene
Aromatic hydrocarbon	GCMS	<0.94	C1-chrysenes	NA	Any or all methyl chrysene isomers
Aromatic hydrocarbon	GCMS	<0.68	C1-dibenzothiophenes	NA	Any or all methyl dibenzothiophene isomers
Aromatic hydrocarbon	GCMS	<1.26	C1-fluoranthenes & pyrenes	NA	Any or all methyl fluoranthene and methyl pyrene isomers
Aromatic hydrocarbon	GCMS	<0.81	C1-fluorenes	NA	Any or all methyl fluorene isomers
Aromatic hydrocarbon	GCMS	<2.26	C1-naphthalenes	NA	Any or all methyl naphthalene isomers
Aromatic hydrocarbon	GCMS	<0.60	C1-phenanthrenes & anthracenes	NA	Any or all methyl phenanthrene and methyl anthracenes isomer
Aromatic hydrocarbon	GCMS	<0.94	C2-chrysenes	NA	Any or all ethyl chrysene isomers
Aromatic hydrocarbon	GCMS	<0.66	C2-dibenzothiophenes	NA	Any or all ethyl dibenzothiophene isomers
Aromatic hydrocarbon	GCMS	<0.80	C2-fluorenes	NA	Any or all ethyl fluorenes isomers
Aromatic hydrocarbon	GCMS	<1.61	C2-naphthalenes	NA	Any or all ethyl naphthalenes isomers
Aromatic hydrocarbon	GCMS	<0.58	C2-phenanthrenes & anthracenes	NA	Any or all ethyl phenanthrenes and ethyl anthracenes isomers
Aromatic hydrocarbon	GCMS	<0.91	C3-chrysenes	NA	Any or all propyl chrysene isomers

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Aromatic hydrocarbon	GCMS	<0.66	C3-dibenzothiophenes	NA	Any or all propyl dibenzothiophene isomers
Aromatic hydrocarbon	GCMS	<0.81	C3-fluorenes	NA	Any or all propyl fluorene isomers
Aromatic hydrocarbon	GCMS	<1.94	C3-naphthalenes	NA	Any or all propyl naphthalene isomers
Aromatic hydrocarbon	GCMS	<0.58	C3-phenanthrenes & anthracenes	NA	Any or all propyl phenanthrene and propyl anthracene isomers
Aromatic hydrocarbon	GCMS	<0.91	C4-chrysenes	NA	Any or all butyl chrysene isomers
Aromatic hydrocarbon	GCMS	<1.94	C4-naphthalenes	NA	Any or all butyl naphthalene isomers
Aromatic hydrocarbon	GCMS	<0.58	C4-phenanthrenes & anthracenes	NA	Any or all butyl phenanthrene and butyl anthracene isomers
Aromatic hydrocarbon	GCMS	<0.46	chrysene	218019	chrysene
Aromatic hydrocarbon	GCMS	<0.41	dibenz(a,h)anthracene	53703	dibenz(a,h)anthracene
Aromatic hydrocarbon	GCMS	<0.33	dibenzothiophene	132650	dibenzothiophene
Aromatic hydrocarbon	GCMS	<0.28	fluoranthene	206440	fluoranthene
Aromatic hydrocarbon	GCMS	<0.24	fluorene	86737	fluorene
Aromatic hydrocarbon	GCMS	<0.67	indeno(1,2,3-cd)pyrene	193395	Indenopyrene
Aromatic hydrocarbon	GCMS	<0.52	naphthalene	91203	tar camphor
Aromatic hydrocarbon	GCMS	<0.63	perylene	198550	perylene
Aromatic hydrocarbon	GCMS	<0.20	phenanthrene	85018	Coal tar pitch volatiles
Aromatic hydrocarbon	GCMS	<0.40	pyrene	129000	Benzo[d,e,f]phenanthrene
Dioxin and Furans	GCMS	<0.009	1,2,3,4,6,7,8-HpCDD	35822469	1,2,3,4,6,7,8-heptachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.009	1,2,3,4,6,7,8-HpCDF	67562394	1,2,3,4,6,7,8-heptachlorodibenzofuran
Dioxin and Furans	GCMS	<0.009	1,2,3,4,7,8,9-HpCDF	55673897	1,2,3,4,7,8,9-heptachlorodibenzofuran
Dioxin and Furans	GCMS	<0.009	1,2,3,4,7,8-HxCDD	39227286	1,2,3,4,7,8-hexachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.009	1,2,3,4,7,8-HxCDF	70648269	1,2,3,4,7,8-hexachlorodibenzofuran
Dioxin and Furans	GCMS	<0.009	1,2,3,6,7,8-HxCDD	57653857	1,2,3,6,7,8-hexachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.009	1,2,3,6,7,8-HxCDF	57117449	1,2,3,6,7,8-hexachlorodibenzofuran
Dioxin and Furans	GCMS	<0.009	1,2,3,7,8,9-HxCDD	19408743	1,2,3,7,8,9-hexachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.009	1,2,3,7,8,9-HxCDF	72918219	1,2,3,7,8,9-hexachlorodibenzofuran
Dioxin and Furans	GCMS	<0.009	1,2,3,7,8-PeCDD	40321764	1,2,3,7,8-pentachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.009	1,2,3,7,8-PeCDF	57117416	1,2,3,7,8-pentachlorodibenzofuran
Dioxin and Furans	GCMS	<0.009	2,3,4,6,7,8-HxCDF	60851345	2,3,4,6,7,8-hexachlorodibenzofuran
Dioxin and Furans	GCMS	<0.009	2,3,4,7,8-PeCDF	57117314	2,3,4,7,8-pentachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.002	2,3,7,8-TCDD	1746016	2,3,7,8-tetrachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.002	2,3,7,8-TCDF	51207319	2,3,7,8-tetrachlorodibenzofuran

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Dioxin and Furans	GCMS	<0.002	Cl4-PCDD	NA	Any or all of the 22 possible tetrachlorinated dibenzo-p-dioxin isomers
Dioxin and Furans	GCMS	<0.002	Cl4-PCDF	NA	Any or all of the 38 possible tetrachlorinated dibenzofuran isomers.
Dioxin and Furans	GCMS	<0.009	Cl5-PCDD	NA	Any or all of the 14 possible pentachlorinated dibenzo-p-dioxin isomers.
Dioxin and Furans	GCMS	<0.009	Cl5-PCDF	NA	Any or all of the 28 possible pentaclorinated dibenzofuran isomers.
Dioxin and Furans	GCMS	<0.009	Cl6-PCDD	NA	Any or all of the 10 possible hexachlorinated dibenzo-p-dioxin isomers
Dioxin and Furans	GCMS	<0.009	Cl6-PCDF	NA	Any or all of the 16 possible hexachlorinated dibenzofuran isomers.
Dioxin and Furans	GCMS	<0.009	Cl7-PCDD	NA	Any or all of the 2 possible heptachlorinated dibenzo-p-dioxin isomers
Dioxin and Furans	GCMS	<0.009	Cl7-PCDF	NA	Any of all of the 4 possible heptachlorinated dibenzofuran isomers.
Dioxin and Furans	GCMS	<0.002	OCDD	3268879	octachlorodibenzo-p-dioxin
Dioxin and Furans	GCMS	<0.002	OCDF	39001020	octachlorodibenzofuran
Flame-resistant chemical	GCMS	<0.17	BDE-1	NA	4-monobromodiphenyl ether
Flame-resistant chemical	GCMS	<0.17	BDE-10	NA	2,6-dibromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.20	BDE-100	189084648	2,2’,4,4’,6-pentabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.17	BDE-11	6903635	3,3’-dibromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.18	BDE-116	NA	2,3,4,5,6-pentabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.18	BDE-118	NA	2,3',4,4',5-pentabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.18	BDE-119	189084660	2,3’,4,4’6-pentabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.18	BDE-12	NA	2,3,4,4',5,6'-hexabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.18	BDE-126	NA	3,3’,4,4’,5-pentabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.17	BDE-13	NA	3,4’-dibromodiphenyl ether
Flame-resistant chemical	GCMS	<0.14	BDE-138	182677301	2,2',3,4,4',5'-hexabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.18	BDE-15	2050477	2,2’,4,4’,5,5’ hexabromobiphenyl ether

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Flame-resistant chemical	GCMS	<0.20	BDE-153	68631492	2,2',4,4',5,5'-hexabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.20	BDE-154	207122154	2,2’,4,4’,5,6’-hexabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.14	BDE-155	35854945	2,2’,4,4’,6,6’-hexabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.14	BDE-166	189084580	2,3,4,4',5,6-hexabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.18	BDE-17	147217752	2,2’,4-tribromodiphenyl ether
Flame-resistant chemical	GCMS	<0.13	BDE-181	189084671	2,2’,3,4,4’,5,6-heptabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.13	BDE-183	207122165	2,2',3,4,4',5',6-heptabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.10	BDE-190	NA	2,3,3',4,4',5,6-heptabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.10	BDE-194	NA	2,2′,3,3′,4,4′,5,5′-Octabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-195	NA	2,2′,3,3′,4,4′,5,6-Octabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-196	NA	2,2’,3,3’,4,4’,5’,6-octabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.10	BDE-197	NA	2,2’,3,3’,4,4’,6,6’-Octabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-198/199/203/200	NA	BDE-198+BDE-199+BDE-203+BDE-200
Flame-resistant chemical	GCMS	<0.17	BDE-2	6876002	3-bromodipheyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-201	NA	2,2’,3,3’,4,5’,6,6’-Octabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-202	NA	2,2’,3,3’,5,5’,6,6’-Octabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-204	NA	2,2’,3,4,4’,5,6,6’-Octabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-205	NA	2,3,3’,4,4’,5,5',6-Octabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.10	BDE-206	63936561	2,2',3,3',4,4',5,5',6-nonabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.10	BDE-207	NA	2,2’,3,3’,4,4’,5,6,6’-nonabromodiphenyl ether

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Flame-resistant chemical	GCMS	<0.10	BDE-208	NA	2,2',3,3',4,5,5',6,6'-nonabromodiphenyl ether
Flame-resistant chemical	GCMS	<1.40	BDE-209	NA	2,2’,3,3’,4,4’,5,5’,6,6’-decabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.17	BDE-25	147217774	2,3’,4-tribromodiphenyl ether
Flame-resistant chemical	GCMS	<0.20	BDE-28	41318756	2,4,4’-tribromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-3	NA	2,2',3,4,4',5'-hexabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-30	NA	2,4,6-tribromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-32	NA	2,4’,6-tribromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-33	NA	2’,3,4-tribromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-35	NA	3,3’,4-tribromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-37	NA	3,4,4’-tribromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.20	BDE-47	54364312	2,2’,4,4’ tetrabromodiphenyl ether
Flame-resistant chemical	GCMS	<0.20	BDE-49/71	NA	BDE-49+BDE-71
Flame-resistant chemical	GCMS	<0.11	BDE-66	187084615	2,3',4,4'-tetrabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-7	NA	2,2',4,4',5,5'-hexabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.20	BDE-75	189084637	2,4-dibromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.20	BDE-77	NA	3,3’,4,4’-tetrabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.17	BDE-8	NA	2,2',4,4',5,6'-heptabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.36	BDE-85	182346210	2,2',3,4,4'-pentabromodiphenyl Ether
Flame-resistant chemical	GCMS	<0.18	BDE-99	603486092	2,2’,4,4’,5-pentabromodiphenyl ether
Flame-resistant chemical	ICP-AES	<1.4	BDE-Total	NA	Sum of the BDE congeners
Organic mercury	AFS	<0.002	Methyl mercury	22967926	Methyl mercury
Organo-halide pesticide	GCECD	<0.15	1,2,3,4-Tetrachlorobenzene	634662	1,2,3,4-benzene tetrachloride
Organo-halide pesticide	GCECD	<0.27	1,2,3,5-Tetrachlorobenzene	634902	1,2,3,5-benzene tetrachloride
Organo-halide pesticide	GCECD	<0.21	o,p'-DDD	72548	1,1-dichloro-2-(o-chlorophenyl)-2-(p-chlorophenyl)ethane
Organo-halide pesticide	GCECD	<0.45	o,p'-DDE	3424826	1,1-dichloro-2-(o-chlorophenyl)-2-(p-chlorophenyl)ethylene
Organo-halide pesticide	GCECD	<0.16	o,p'-DDT	789026	1,1,1-trichloro-2-(o-chlorophenyl)-2-(p-chlorophenyl)ethane
Organo-halide pesticide	GCECD	<0.45	p,p'-DDD	72548	1,1-dichloro-2,2-bis(p-chlorophenyl)ethane
Organo-halide pesticide	GCECD	<0.91	p,p'-DDE	72559	1,1-dichloro-2,2-bis(p-chlorophenyl)ethylene

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Organo-halide pesticide	GCECD	<0.91	p,p'-DDT	50293	1,1,1-trichloro-2,2-bis(p-chlorophenyl)ethane
Organo-halide pesticide	GCECD	<0.12	Aldrin	309002	1,4:5,8-dimethanonaphthalene
Organo-halide pesticide	GCECD	<0.21	alpha-BHC	319846	alpha HCH; alpha benzene hexachloride
Organo-halide pesticide	GCECD	<0.45	alpha-Chlordane	5103719	cis-Chlordane
Organo-halide pesticide	GCECD	<0.22	beta-BHC	319857	beta HCH; beta benzene hexachloride
Organo-halide pesticide	GCECD	<0.15	cis-Nonachlor	5103731	cis-Nonachlor
Organo-halide pesticide	GCECD	<0.13	44DDMU	1022226	4,4'-DDMU; metabolite of DDD
Organo-halide pesticide	GCECD	<0.91	DDT-total	NA	Sum of all DDT, DDE, DDD isomers
Organo-halide pesticide	GCECD	<0.11	delta-BHC	319868	delta HCH; delta benzene hexachloride
Organo-halide pesticide	GCECD	<0.19	dieldrin	60571	dieldrin
Organo-halide pesticide	GCECD	<0.50	endosulfan I	959988	endosulfan-alpha
Organo-halide pesticide	GCECD	<0.15	endosulfan II	33213659	endosulfan-beta
Organo-halide pesticide	GCECD	<0.45	endosulfan sulfate	1031078	endosulfan sulfate
Organo-halide pesticide	GCECD	<0.20	endrin	72208	endrin
Organo-halide pesticide	GCECD	<0.15	gamma-BHC	58899	lindane; gamma HCH
Organo-halide pesticide	GCECD	<0.15	gamma-Chlordane	5566347	gamma-Chlordane
Organo-halide pesticide	GCECD	<0.14	heptachlor	76448	heptachlor
Organo-halide pesticide	GCECD	<0.16	heptachlor epoxide	1024573	heptachlor Epoxide
Organo-halide pesticide	GCECD	<0.12	oxychlordane	27304138	Oxychlordane
Organo-halide pesticide	GCECD	<0.15	pentachloroanisole	1825214	pentachloroanisole
Organo-halide pesticide	GCECD	<0.36	BHC-total	NA	Sum of all BHC isomers
Organo-halide pesticide	GCECD	<10.5	toxaphene	8001352	toxaphene
Organo-halide pesticide	GCECD	<0.45	trans-Nonachlor	39765805	trans-Nonachlor
Organo-halide pesticide	GCMS	<0.23	HCB	118741	hexachlorobenzene
Organo-halide pesticide	GCECD	<0.67	pentachorobenzene	608935	pentachorobenzene
Organophosphate	GCMS	<0.15	chlorpyrifos	2921882	Chlorpyrifos
Paraffin hydrocarbon	GCMS	<20.0	n-decane	124185	n-decane
Paraffin hydrocarbon	GCMS	<20.0	n-docosane	629970	n-docosane
Paraffin hydrocarbon	GCMS	<20.0	n-dodecane	112403	n-dodecane
Paraffin hydrocarbon	GCMS	<14.5	n-dotriacontane	544854	n-dotriacontane
Paraffin hydrocarbon	GCMS	<16.2	n-eicosane	112958	n-eicosane
Paraffin hydrocarbon	GCMS	<17.1	n-heneicosane	629947	n-heneicosane
Paraffin hydrocarbon	GCMS	<14.8	n-hentriacontane	630046	n-hentriacontane
Paraffin hydrocarbon	GCMS	<20.0	n-heptacosane	593497	n-heptacosane
Paraffin hydrocarbon	GCMS	<20.0	n-heptadecane	629787	n-heptadecane

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Paraffin hydrocarbon	GCMS	<20.0	n-hexacosane	630013	n-hexacosane
Paraffin hydrocarbon	GCMS	<20.0	n-hexadecane	544763	n-hexadecane
Paraffin hydrocarbon	GCMS	<13.0	n-nonacosane	630035	n-nonacosane
Paraffin hydrocarbon	GCMS	<20.0	n-nonadecane	629925	n-nonadecane
Paraffin hydrocarbon	GCMS	<14.0	n-octacosane	630024	n-octacosane
Paraffin hydrocarbon	GCMS	<20.0	n-octadecane	593453	n-octadecane
Paraffin hydrocarbon	GCMS	<20.0	n-pentacosane	629992	n-pentacosane
Paraffin hydrocarbon	GCMS	<20.0	n-pentadecane	629629	n-pentadecane
Paraffin hydrocarbon	GCMS	<20.0	n-tetracosane	646311	n-tetracosane
Paraffin hydrocarbon	GCMS	<20.0	n-tetradecane	629594	n-tetradecane
Paraffin hydrocarbon	GCMS	<14.2	n-tetratriacontane	14167590	n-tetratriacontane
Paraffin hydrocarbon	GCMS	<13.8	n-triacontane	638686	n-triacontane
Paraffin hydrocarbon	GCMS	<0.02	n-tricosane	638675	n-tricosane
Paraffin hydrocarbon	GCMS	<20.0	n-tridecane	629505	n-tridecane
Paraffin hydrocarbon	GCMS	<12.3	n-tritriacontane	630057	n-tritriacontane
Paraffin hydrocarbon	GCMS	<20.0	n-undecane	1120214	hendecane
Paraffin hydrocarbon	GCMS	<20.0	phytane	638368	2,6,10,14-tetramethylhexadecane
Paraffin hydrocarbon	GCMS	<20.0	pristane	1921706	2,6,10,14-tetramethylpentadecane
Polychlorinated biphenyl	GCMS	<0.03	PCB-1	2051607	2-monochlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.008	PCB-10	33146451	2,6-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-103	60145213	2,2',4,5,6'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-104	56558168	2,2',4,6,6'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-105	32598144	2,3,3'4,4'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-106	70424690	2,3,3',4,5-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-107	70424689	2,3,3',4',5-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-108/124	NA	PCB-108+PCB-124
Polychlorinated biphenyl	GCMS	<0.15	PCB-11	2050671	3,3'-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-110/115	NA	PCB-110+PCB-115
Polychlorinated biphenyl	GCMS	<0.15	PCB-111	39635320	2,3,3',5,5'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-112	74472369	2,3,3',5,6-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-114	74472370	2,3,4,4',5-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-117	68194116	2,3,4',5,6-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.02	PCB-118	31508006	2,3',4,4',5-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.02	PCB-12/13	NA	PCB-12+PCB-13
Polychlorinated biphenyl	GCMS	<0.08	PCB-120	68194127	2,3',4,5,5'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-121	56558180	2,3',4,5,6-pentachlorobiphenyl

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Polychlorinated biphenyl	GCMS	<0.08	PCB-122	76842074	2',3,3',4,5-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-123	65510443	2',3,4,4',5-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-1242	53469219	Aroclor 1242 PCB mixture
Polychlorinated biphenyl	GCMS	<0.15	PCB-1248	12672296	Aroclor 1248 PCB mixture
Polychlorinated biphenyl	GCMS	<0.15	PCB-1254	11097691	Aroclor 1254 PCB mixture
Polychlorinated biphenyl	GCMS	<0.08	PCB-126	57465288	3,3',4,4',5-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-1260	11096825	Aroclor 1260 PCB mixture
Polychlorinated biphenyl	GCMS	<0.15	PCB-127	39635331	3,3',4,5,5'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-128/166	NA	PCB-128+PCB-166
Polychlorinated biphenyl	GCMS	<0.08	PCB-129/138/163	NA	PCB-129+PCB-138+PCB-163
Polychlorinated biphenyl	GCMS	<0.08	PCB-130	52663668	2,2',3,3',4,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-131	61798707	2,2',3,3',4,6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-132	38380051	2,2',3,3',4,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-133	35694043	2,2',3,3',5,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-134	NA	2,2',3,3',5,6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.10	PCB-135/151	NA	PCB-135+PCB-151
Polychlorinated biphenyl	GCMS	<0.03	PCB-136	38411222	2,2',3,3',6,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-137	35694065	2,2',3,4,4',5-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-139/140	NA	PCB-139+PCB-140
Polychlorinated biphenyl	GCMS	<0.15	PCB-14	34883415	3,5-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-141	52712046	2,2',3,4,5,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-142	41411614	2,2',3,4,5,6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-143	68194150	2,2',3,4,5,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-144	68194149	2,2',3,4,5',6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-145	74472405	2,2',3,4,6,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-146	51908168	2,2',3,4',5,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-147/149	NA	PCB-147+PCB-149
Polychlorinated biphenyl	GCMS	<0.15	PCB-148	74472416	2,2',3,4',5,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-15	2050682	4,4'-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-150	68194081	2,2',3,4',6,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-152	68194092	2,2',3,5,6,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-153/168	NA	PCB-153+PCB-168
Polychlorinated biphenyl	GCMS	<0.08	PCB-154	60145224	2,2',4,4',5',6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-155	33979032	2,2',4,4',6,6'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-156/157	NA	PCB-156+PCB-157
Polychlorinated biphenyl	GCMS	<0.03	PCB-158	74472427	2,3,3',4,4',6-hexachlorobiphenyl

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Polychlorinated biphenyl	GCMS	<0.15	PCB-159	39635353	2,3,3',4,5,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.02	PCB-16	38444789	2,2',3-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-160	41411625	2,3,3',4,5,6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-161	74472438	2,3,3',4,5',6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-162	39635342	2,3,3',4',5,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-164	74472450	2,3,3',4',5',6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-165	74472461	2,3,3',5,5',6-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-167	52663726	2,3,4,4',5,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-169	32774166	3,3',4,4',5,5'-hexachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-17	37680663	2,2',4-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-170	35065306	2,2',3,3',4,4',5-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-171/173	NA	PCB-171+PCB-173
Polychlorinated biphenyl	GCMS	<0.15	PCB-172	52663748	2,2',3,3',4,5,5'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-174	38411255	2,2',3,3',4,5,6'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-175	40186707	2,2',3,3',4,5',6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-176	52663657	2,2',3,3',4,6,6'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-177	52663704	2,2',3,3',4',5,6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-178	52663679	2,2',3,3',5,5',6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-179	52663646	2,2',3,3',5,6,6'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-18/30	NA	PCB-18+PCB-30
Polychlorinated biphenyl	GCMS	<0.15	PCB-180/193	NA	PCB-180+PCB-193
Polychlorinated biphenyl	GCMS	<0.15	PCB-181	74472472	2,2',3,4,4',5,6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-182	60145235	2,2',3,4,4',5,6'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.16	PCB-183	52663691	2,2',3,4,4',5',6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-184	74472483	2,2',3,4,4',6,6'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-185	52712483	2,2',3,4,5,5',6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-186	74472494	2,2',3,4,5,6,6'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-187	52663680	2,2',3,4,5,5',6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-188	74487857	2,2',3,4',5,6,6'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-189	39635319	2,3,3',4,4',5,5'-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.02	PCB-19	38444734	2,2',6-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-190	41411647	2,3,3',4,4',5,6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-191	74472507	2,3,3',4,4',5',6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-192	74472518	2,3,3',4,5,5',6-heptachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-194	35694087	2,2',3,3',4,4',5,5'-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-195	52663782	2,2',3,3',4,4',5,6-octachlorobiphenyl

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Polychlorinated biphenyl	GCMS	<0.15	PCB-196	42740501	2,2',3,3',4,4',5,6'-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-197	33091177	2,2',3,3',4,4',6,6'-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-198/199	NA	PCB-198+PCB-199
Polychlorinated biphenyl	GCMS	<0.002	PCB-2	2051618	3-monochlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-20/28	NA	PCB-20+PCB-28
Polychlorinated biphenyl	GCMS	<0.15	PCB-200	52663737	2,2',3,3',4,5,6,6'-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-201	40186718	2,2',3,3',4,5',6,6'-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-202	2136994	2,2',3,3',5,5',6,6'-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-203	52663760	2,2',3,4,4',5,5',6-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-204	74472529	2,2',3,4,4',5,6,6'-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-205	74472530	2,3,3',4,4',5,5',6-octachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-206	40186729	2,2',3,3',4,4',5,5',6-nonachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-207	52663793	2,2',3,3',4,4',5,6,6'-nonachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.15	PCB-208	52663771	2,2',3,3',4,5,5',6,6'-nonachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-209	2051243	decachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-21/33	NA	PCB-21+PCB-33
Polychlorinated biphenyl	GCMS	<0.03	PCB-22	38444858	2,3,4'-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-23	55720440	2,3,5-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-24	55702459	2,3,6-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-25	55712373	2,3',4-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-26/29	NA	PCB-26+PCB-29
Polychlorinated biphenyl	GCMS	<0.03	PCB-27	38444767	2,3',6-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-3	2051629	4-monochlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-31	16606023	2,4',5-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-32	38444778	2,4',6-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-34	37680685	2',3,5-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-35	37680696	3,3',4-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-36	38444870	3,3',5-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-37	38444905	3,4,4'-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-38	53555661	3,4,5-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-39	38444881	3,4',5-trichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-4	13029088	2,2'-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-40/41/71	NA	PCB-40+PCB-41+PCB-71
Polychlorinated biphenyl	GCMS	<0.03	PCB-42	36559225	2,2',3,4'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-43/73	NA	PCB-43+PCB-73
Polychlorinated biphenyl	GCMS	<0.03	PCB-44/47/65	NA	PCB-44+PCB-47+PCB-65

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Polychlorinated biphenyl	GCMS	<0.03	PCB-45/51	NA	PCB-45+PCB-51
Polychlorinated biphenyl	GCMS	<0.03	PCB-46	41464475	2,2',3,6'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-48	70362479	2,2',4,5-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-49/69	NA	PCB-49+PCB-69
Polychlorinated biphenyl	GCMS	<0.08	PCB-5	16605917	2,3-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-50/53	NA	PCB-50+PCB-53
Polychlorinated biphenyl	GCMS	<0.08	PCB-52	35693993	2,2',5,5'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-54	15968055	2,2',6,6'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-55	74338242	2,3,3',4'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-56	41464431	2,3,3',4'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-57	70424678	2,3,3',5-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-58	41464497	2,3,3',5'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-59/62/75	NA	PCB-59+PCB-62+PCB-75
Polychlorinated biphenyl	GCMS	<0.008	PCB-6	25569806	2,3'-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-60	33025411	2,3,4,4'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-63	74472347	2,3,4',5-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-64	52663588	2,3,4',6-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-66	32598100	2,3',4,4'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-67	73575538	2,3',4,5-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-68	73575527	2,3',4,5'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-7	33284503	2,4-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-70/61/74/76	NA	PCB-70+PCB-61+PCB-74+PCB-76
Polychlorinated biphenyl	GCMS	<0.08	PCB-72	41464420	2,3',5,5'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-73	74338231	2,3',5',6-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-77	32598133	3,3',4,4'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-78	70362491	3,3',4,5-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-79	41464486	3,3',4,5'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-8	34883437	2,4'-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-80	33284525	3,3',5,5'-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-81	70362504	3,4,4',5-tetrachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-82	52663624	2,2',3,3',4-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-83/99	NA	PCB-83+PCB-99
Polychlorinated biphenyl	GCMS	<0.08	PCB-84	52663602	2,2',3,3',6-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.03	PCB-85/116	NA	PCB-85+PCB-116
Polychlorinated biphenyl	GCMS	<0.08	PCB-86/87/97/109/119/125	NA	PCB-86+PCB-87+PCB-97+PCB-109+PCB-119+PCB-125

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

Polychlorinated biphenyl	GCMS	<0.08	PCB-88/91	NA	PCB-88+PCB-91
Polychlorinated biphenyl	GCMS	<0.08	PCB-89	73575572	2,2',3,4,6'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-9	34883391	2,5-dichlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.20	PCB-90/101/113	NA	PCB-90+PCB-101+PCB-113
Polychlorinated biphenyl	GCMS	<0.08	PCB-92	52663613	2,2',3,5,5'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-94	73575550	2,2',3,5,6'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-95	38379996	2,2',3,5',6-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-96	73575549	2,2',3,6,6'-pentachlorobiphenyl
Polychlorinated biphenyl	GCMS	<0.08	PCB-98/100	NA	PCB-98+PCB-100
Polychlorinated biphenyl	GCMS	<0.08	PCB-98/102	NA	PCB-98+PCB-102
Polychlorinated biphenyl	GCMS	<1.51	PCB-total	1336363	Sum of all PCB congeners
Trace element	ICP-AES	<1.11	Al	7429905	Aluminum
Trace element	ICP-MS	<0.51	As	7784421	Arsenic
Trace element	ICP-AES	<0.14	B	7440428	Boron
Trace element	ICP-AES	<0.02	Ba	7440393	Barium
Trace element	ICP-AES	<0.01	Be	7440417	Beryllium
Trace element	ICP-MS	<0.01	Cd	7440439	Cadmium
Trace element	ICP-AES	<0.11	Cr	7440473	Chromium
Trace element	ICP-AES	<0.11	Cu	7440508	Copper
Trace element	ICP-AES	<0.23	Fe	7439896	Iron
Trace element	CVAAS	<0.002	Hg	7439976	Mercury
Trace element	ICP-AES	<0.23	Mg	7439954	Magnesium
Trace element	ICP-AES	<0.05	Mn	7439965	Manganese
Trace element	ICP-AES	<0.25	Mo	7439987	Molybdenum
Trace element	ICP-AES	<0.11	Ni	7440020	Nickel
Trace element	ICP-MS	<0.01	Pb	7439921	Lead
Trace element	ICP-MS	<0.02	Se	7782492	Selenium
Trace element	ICP-AES	<0.01	Sr	7440246	Strontium
Trace element	ICP-AES	<0.25	V	7440622	Vanadium
Trace element	ICP-AES	<0.13	Zn	7440666	Zinc

Table 2. Chemical names, abbreviations, analytical methods, limit of detection. ["<", less than; "NA", not available]

1-Tissue processing and analytical methods are fully described in Appendices 3A and 3B. Analytical method identifiers: GCMS, gas chromatography, mass spectroscopy; ICP-AES, inductively coupled plasma mass spectroscopy; AFS, atomic fluorescence; GC-ECP, gas chromatography, electron capture detection; ICP-MS, inductively coupled plasma mass spectroscopy; CVAAS, cold-vapor atomic absorption spectroscopy.

2-Liimit of detection reported is the lowest method detection limit for the sample batch.

3-Chemical name used in this report. PCB and BDE congeners are designated using the Ballschmiter-Zell numbering system. Certain PCB or BDE congeners will co-elute during analysis.

4-Chemical Abstract Service (CAS) number. "NA" indicates that no CAS number is available at the time of this report.

Table 3. Moisture content, methyl mercury and element concentrations in Rio Grande silvery minnow carcasses collected from six sites (Table 1; Figure 2) along the Middle Rio Grande, New Mexico, during 2006-2008, and all site data combined.

[Moisture content, methyl mercury and element concentration data are the mean±1 standard deviation with the number of fish composite samples and range of concentrations in parentheses. Means within a row that have no upper case letters are not significantly different. Means within a row with upper case letters and sharing the same upper case letter are not significantly ($P<0.05$) different. "±", plus or minus; "MRG", Middle Rio Grande; "NM", New Mexico; Site 1, MRG at Bernalillo, NM; Site 2, MRG at Alameda, NM; Site 3, MRG at Los Padillas, NM; Site 4, MRG at Los Lunas, NM; Site 5, MRG at La Joya, NM; Site 6, MRG near San Antonio, NM; "µg/g", microgram per gram; dry, dry weight basis; wet, wet weight basis; "<", less than; "%", percent]

Contaminant (Unit Basis)	1. Bernalillo	2. Alameda	3. Los Padillas	4. Los Lunas	5. La Joya	6. San Antonio	All site data combined
Aluminum (µg/g dry)	519 ± 380.7 (5; 140-1150)	406 ± 302.2 (6; 141-970)	234 ± 268.9 (6; 81-767)	315 ± 212.5 (6; 61-626)	176 ± 110.2 (5; 36.4-328)	123 ± 99.1 (5; 30.8-289)	298 ± 266.9 (33; 30.8-1150)
Aluminum (µg/g wet)	140 ± 103.2 (5; 34.7-308)	109 ± 84.6 (6; 33.6-267)	64.6 ± 77.5 (6; 22-219)	90.6 ± 69.6 (6; 17-210)	50.8 ± 34.6 (5; 9.9-102)	35.5 ± 30.4 (5; 8.8-87.3)	82.3 ± 75.0 (33; 8.8-308)
Arsenic (µg/g dry)	3.7^{A} ± 0.6 (5; 3.0-4.5)	3.5^{A} ± 0.3 (6; 3.1-3.9)	3.0^{AB} ± 0.5 (6; 2.6-3.8)	3.0^{AB} ± 0.3 (6; 2.7-3.4)	3.0^{AB} ± 0.2 (5; 2.8-3.2)	2.8^{AB} ± 0.3 (5; 2.6-3.3)	3.2^{BC} ± 0.5 (33; 2.6-4.5)
Arsenic (µg/g wet)	1.0 ± 0.2 (5; 0.8-1.2)	0.9 ± 0.1 (6; 0.8-1.1)	0.8 ± 0.1 (6; 0.7-1.0)	0.8 ± 0.1 (6; 0.7-1.0)	0.8 ± 0.1 (5; 0.8-1.0)	0.8 ± 0.1 (5; 0.7-0.9)	0.9 ± 0.1 (33; 0.7-1.2)
Barium (µg/g dry)	79.3^{A} ± 8.9 (5; 69.5-88.4)	71.4^{AB} ± 4.6 (6; 67.6-79.3)	58.7^{BC} ± 8.3 (6; 47.5-68.0)	58.0^{BC} ± 6.5 (6; 47.8-65.3)	36.6^{C} ± 12.3 (5; 22.1-54.4)	34.8^{C} ± 13.9 (5; 22.1-57.6)	57.0 ± 18.2 (33; 22.1-88.4)
Barium (µg/g wet)	21.0 ± 1.8 (5; 18.8-23.6)	18.7 ± 1.2 (6; 17.3-20.4)	15.6 ± 1.7 (6; 12.9-17.0)	16.3 ± 3.0 (6; 13.3-21.9)	10.5 ± 4.2 (5; 6.1-17.0)	9.9 ± 4.4 (5; 6.5-17.4)	15.5 ± 4.8 (33; 6.1-23.6)
Copper (µg/g dry)	9.2 ± 12.6 (5; 3.3-31.7)	13.0 ± 17.6 (6; 3.2-46.7)	5.9 ± 6.9 (6; 2.6-20.0)	5.1 ± 4.8 (6; 2.7-14.9)	2.8 ± 0.3 (5; 2.3-3.2)	3.5 ± 1.9 (5; 2.3-6.9)	6.7 ± 9.6 (33; 2.3-46.7)
Copper (µg/g wet)	2.5 ± 3.4 (5; 0.9-8.5)	3.4 ± 4.5 (6; 0.8-11.8)	1.6 ± 2.0 (6; 0.7-5.8)	1.4 ± 1.3 (6; 0.7-4.0)	0.8 ± 0.8 (5; 0.7-0.9)	1.0 ± 0.6 (5; 0.7-2.0)	1.8 ± 2.5 (33; 0.7-11.8)
Iron (µg/g dry)	589 ± 393.3 (5; 149-1180)	476 ± 325.6 (6; 181-1060)	279 ± 280.7 (6; 125-838)	375 ± 229.4 (6; 106-634)	236 ± 117.7 (5; 66.1-344)	164 ± 92.6 (5; 68.7-309)	355 ± 282.1 (33; 66.1-1180)
Iron (µg/g wet)	159 ± 107.9 (5; 37.0-316)	128 ± 92.0 (6; 45.8-292)	76.6 ± 81.1 (6; 33.5-239)	107 ± 71.2 (6; 29.5-212)	67.5 ± 35.7 (5; 18.0-107)	46.9 ± 28.3 (5; 19.7-93.3)	98.0 ± 79.4 (33; 18.0-316)
Lead (µg/g dry)	0.6 ± 0.4 (5; 0.2-1.1)	0.6 ± 0.4 (6; 0.3-1.5)	0.4 ± 0.2 (6; 0.2-0.8)	0.5 ± 0.3 (6; 0.2-0.9)	0.4 ± 0.1 (5; 0.2-0.6)	0.2 ± 0.03 (5; 0.03-0.1)	0.4 ± 0.3 (33; 0.2-1.5)
Lead (µg/g wet)	0.2 ± 0.1 (5; 0.04-0.3)	0.2 ± 0.1 (6; 0.07-0.4)	0.1 ± 0.1 (6; 0.04-0.2)	0.1 ± 0.1 (6; 0.05-0.2)	0.1 ± 0.04 (5; 0.06-0.1)	0.1 ± 0.03 (5; 0.03-0.1)	0.1 ± 0.1 (33; 0.03-0.4)

Table 3. Moisture content, methyl mercury and element concentrations in Rio Grande silvery minnow carcasses collected from six sites (Table 1; Figure 2) along the Middle Rio Grande, New Mexico, during 2006-2008, and all site data combined.

[Moisture content, methyl mercury and element concentration data are the mean±1 standard deviation with the number of fish composite samples and range of concentrations in parentheses. Means within a row that have no upper case letters are not significantly different. Means within a row with upper case letters and sharing the same upper case letter are not significantly (P<0.05) different. "±", plus or minus; "MRG", Middle Rio Grande; "NM", New Mexico; Site 1, MRG at Bernalillo, NM; Site 2, MRG at Alameda, NM; Site 3, MRG at Los Padillas, NM; Site 4, MRG at Los Lunas, NM; Site 5, MRG at La Joya, NM; Site 6, MRG near San Antonio, NM; "µg/g", microgram per gram; dry, dry weight basis; wet, wet weight basis; "<", less than; "%", percent]

Contaminant (Unit Basis)	1. Bernalillo	2. Alameda	3. Los Padillas	4. Los Lunas	5. La Joya	6. San Antonio	All site data combined
Manganese (µg/g dry)	38.4 ± 18.1 (5; 17.8-58.2)	35.5 ± 18.3 (6; 17.0-62.8)	19.9 ± 10.9 (6; 10.7-39.9)	23.9 ± 11.1 (6; 12.5-36.8)	30.6 ± 15.6 (5; 13.1-49.9)	17.8 ± 7.9 (5; 11.8-31.7)	27.6 ± 15.1 (33; 10.7-62.8)
Manganese (µg/g wet)	10.4 ± 5.2 (5; 4.4-16.1)	9.5 ± 5.4 (6; 4.3-12.9)	5.4 ± 3.2 (6; 2.9-11.4)	6.8 ± 3.6 (6; 3.4-12.3)	8.8 ± 4.9 (5; 3.6-15.6)	5.1 ± 2.5 (5; 3.5-9.6)	7.6 ± 4.4 (33; 2.9-17.9)
Magnesium (µg/g dry)	1654 ± 224.1 (5; 1410-2000)	1528 ± 100.7 (6; 1440-1700)	1462 ± 187.6 (6; 1300-1780)	1517 ± 155.9 (6; 1330-1710)	1380 ± 186.4 (5; 1150-1640)	1354 ± 128.6 (5; 1240-1530)	1484 ± 181.9 (33; 1150-2000)
Magnesium (µg/g wet)	440 ± 62.5 (5; 381-536)	401 ± 38.1 (6; 364-468)	392 ± 57.4 (6; 354-507)	426 ± 72.8 (6; 370-566)	392 ± 76.1 (5; 320-512)	385 ± 45.2 (5; 350-462)	406 ± 58.5 (33; 320-566)
Mercury (µg/g dry)	0.22 ± 0.07 (5; 0.13-0.28)	0.21 ± 0.08 (6; 0.12-0.32)	0.18 ± 0.07 (6; 0.11-0.30)	0.15 ± 0.05 (6; 0.09-0.22)	0.18 ± 0.05 (5; 0.10-0.22)	0.21 ± 0.11 (5; 0.10-0.36)	0.19 ± 0.07 (33; 0.09-0.36)
Mercury (µg/g wet)	0.06 ± 0.02 (5; 0.04-0.07)	0.06 ± 0.02 (6; 0.03-0.08)	0.05 ± 0.02 (6; 0.03-0.07)	0.04 ± 0.02 (6; 0.02-0.06)	0.05 ± 0.01 (5; 0.03-0.06)	0.06 ± 0.03 (5; 0.03-0.10)	0.05 ± 0.02 (33; 0.02-0.10)
Methyl mercury (µg/g dry)	0.18 (1)	0.21 ± 0.02 (2; 0.20-0.22)	0.20 ± 0.07 (2; 0.14-0.24)	0.17 ± 0.04 (2; 0.14-0.20)	0.16 ± 0.06 (3; 0.10-0.22)	0.31 ± 0.08 (2; 0.26-0.36)	0.20 ± 0.07 (12; 0.10-0.36)
Methyl mercury (µg/g wet)	0.05 (1)	0.05 ± 0.003 (2; 0.05-0.06)	0.05 ± 0.01 (2; 0.04-0.06)	0.05 ± 0.01 (2; 0.04-0.05)	0.05 ± 0.02 (3; 0.03-0.06)	0.09 ± 0.02 (2; 0.07-0.10)	0.06 ± 0.02 (12; 0.03-0.10)
Nickel (µg/g dry)	3.5 ± 5.5 (5; 0.7-13.3)	4.0 ± 5.1 (6; 0.5-12.9)	1.6 ± 1.6 (6; 0.2-4.6)	1.4 ± 1.1 (6; 0.2-3.2)	0.5 ± 0.4 (5; 0.2-1.1)	1.1 ± 1.3 (5; 0.2-3.5)	2.0 ± 3.2 (33; 0.2-13.3)
Nickel (µg/g wet)	0.9 ± 1.5 (5; 0.2-3.6)	1.1 ± 1.4 (6; 0.1-3.6)	0.6 ± 0.5 (6; 0.1-0.8)	0.4 ± 0.3 (6; 0.1-0.9)	0.2 ± 0.1 (5; 0.1-0.3)	0.3 ± 0.4 (5; 0.1-1.0)	0.6 ± 0.9 (33; 0.1-3.6)
Selenium (µg/g dry)	1.9 ± 0.3 (5; 1.4-2.1)	2.0 ± 0.3 (6; 1.7-2.4)	1.8 ± 0.4 (6; 1.3-2.5)	1.8 ± 0.2 (6; 1.5-2.1)	2.2 ± 0.7 (5; 1.4-3.2)	1.9 ± 0.6 (5; 1.4-2.8)	1.9 ± 0.4 (33; 1.3-3.2)
Selenium (µg/g wet)	0.5 ± 0.1 (5; 0.4-0.6)	0.5 ± 0.1 (6; 0.4-0.7)	0.5 ± 0.1 (6; 0.4-0.6)	0.5 ± 0.1 (6; 0.4-0.6)	0.6 ± 0.2 (5; 0.4-0.9)	0.6 ± 0.2 (5; 0.4-0.8)	0.5 ± 0.1 (33; 0.3-0.9)

Table 3. Moisture content, methyl mercury and element concentrations in Rio Grande silvery minnow carcasses collected from six sites (Table 1; Figure 2) along the Middle Rio Grande, New Mexico, during 2006-2008, and all site data combined.

[Moisture content, methyl mercury and element concentration data are the mean±1 standard deviation with the number of fish composite samples and range of concentrations in parentheses. Means within a row that have no upper case letters are not significantly different. Means within a row with upper case letters and sharing the same upper case letter are not significantly ($P<0.05$) different. "±", plus or minus; "MRG", Middle Rio Grande; "NM", New Mexico; Site 1, MRG at Bernalillo, NM; Site 2, MRG at Alameda, NM; Site 3, MRG at Los Padillas, NM; Site 4, MRG at Los Lunas, NM; Site 5, MRG at La Joya, NM; Site 6, MRG near San Antonio, NM; "µg/g", microgram per gram; dry, dry weight basis; wet, wet weight basis; "<", less than; "%", percent]

Contaminant (Unit Basis)	**1. Bernalillo**	**2. Alameda**	**3. Los Padillas**	**4. Los Lunas**	**5. La Joya**	**6. San Antonio**	**All site data combined**
Strontium (µg/g dry)	136 ± 22.9 (5; 107-162)	137 ± 24.0 (6; 105-176)	137 ± 24.4 (6; 104-164)	126 ± 15.3 (5; 108-154)	149 ± 23.4 (5; 128-187)	153 ± 35.9 (5; 98.8-186)	139 ± 24.4 (33; 98.8-187)
Strontium (µg/g wet)	36.1 ± 5.3 (5; 28.9-43.4)	35.7 ± 3.9 (5; 29.9-41.9)	36.6 ± 6.5 (5; 28.3-46.7)	36.1 ± 5.3 (5; 28.9-43.4)	42.5 ± 9.1 (5; 36.5-58.3)	43.4 ± 10.5 (5; 28.4-51.3)	38.1 ± 7.6 (33; 28.3-58.3)
Vanadium (µg/g dry)	1.6 ± 1.0 (5; 0.6-3.2)	1.5 ± 0.6 (6; 0.9-2.5)	1.1 ± 0.9 (6; 0.2-0.9)	1.1 ± 0.6 (6; 0.2-1.7)	0.8 ± 0.4 (5; 0.2-1.2)	0.7 ± 0.3 (5; 0.2-1.1)	1.1 ± 0.7 (33; 0.2-3.2)
Vanadium (µg/g wet)	0.4 ± 0.3 (5; 0.1-0.8)	0.4 ± 0.2 (6; 0.2-0.7)	0.3 ± 0.3 (6; 0.1-0.8)	0.3 ± 0.2 (6; 0.1-0.6)	0.2 ± 0.1 (5; 0.1-0.4)	0.2 ± 0.1 (5; 0.1-0.4)	0.3 ± 0.2 (33; 0.1-0.8)
Zinc (µg/g dry)	149 ± 11.8 (5; 136-165)	143 ± 19.2 (6; 122-179)	139 ± 13.3 (6; 119-159)	136 ± 19.6 (6; 114-170)	130 ± 18.6 (5; 112-153)	134 ± 19.7 (5; 110-165)	139 ± 17.1 (33; 110-179)
Zinc (µg/g wet)	39.6 ± 2.9 (5; 36.3-42.3)	37.4 ± 3.8 (6; 33.3-42.6)	37.2 ± 2.5 (6; 34.5-41.9)	38.3 ± 9.5 (6; 31.7-57.0)	36.9 ± 7.1 (5; 30.6-45.6)	38.3 ± 7.3 (5; 29.8-49.8)	37.9 ± 5.7 (33; 29.8-57.0)
Moisture Content (%)	72.7 ± 1.6 (7; 69.9-75.0)	72.8 ± 1.7 (8; 69.8-74.6)	71.7 ± 1.5 (8; 70.0-74.9)	71.8 ± 2.7 (8; 66.8-76.4)	70.9 ± 1.5 (7; 69.5-73.7)	71.7 ± 1.8 (6; 69.5-74.5)	71.9 ± 1.9 (44; 66.8-76.4)

Table 4. Concentrations of elements and moisture content in whole fish or carcasses collected from the Rio Grande and collected nationwide.

[Moisture content and element data are mean ± 1 standard deviation with the number of fish composite samples and range of concentrations in parenthesis. "NCBP", National Contaminant Biomonitoring Program; "µg/g wet", micrograms per gram on a wet weight basis; "%", percent]

Element	This study	Abeyta and Lusk 2004	Roy et al. 1992	Schmitt et al. 2004	Mora 2001 (est. wet weight using 72.1% moisture)	Schmitt et al. 1999 (NCBP)
Aluminum (µg/g wet)	82.3 ± 75.0 (33; 8.8-308)		45.3 ± 60.15 (109; 0.4-364.3)	16.1 ± 11.6 (47; 6.4-61.8)	43.9 ± 106.8 (23; 2.1-477)	
Arsenic (µg/g wet)	0.9 ± 0.1 (33; 0.7-1.2)	0.3 ± 0.2 (27; 0.1-0.7)	0.1 ± 0.2 (94; 0.01-0.8)	0.2 ± 0.1 (47; 0.03-0.6)	0.2 ± 0.4 (55; <0.01-2.6)	0.2 ± 0.2 (2873; 0.01-3.4)
Barium (µg/g wet)	15.5 ± 4.8 (33; 6.1-23.6)		2.9 ± 2.5 (93; 0.15-17.3)	1.2 ± 0.9 (47; 0.2-3.7)	1.0 ± 1.5 (23; 0.1-7.1)	
Copper (µg/g wet)	1.8 ± 2.5 (33; 0.7-11.8)	1.5 ± 1.8 (27; 0.7-8.9)	1.63 ± 1.0 (112; 0.32-6.14)	0.9 ± 0.4 (47; 0.3-1.8)	0.5 ± 0.5 (51; 0.1-2.7)	1.0 ± 1.6 (1255; 0.1-38.8)
Iron (µg/g wet)	98.0 ± 79.4 (33; 18.0-316)		82.5 ± 93.5 (112; 10.7-552)	107 ± 110 (47; 11.2-524)	43.6 ± 83.1 (22; 3.5-356)	
Lead (µg/g wet)	0.1 ± 0.1 (33; 0.03-0.4)	0.1 ± 0.1 (27; 0.03-0.4)	0.5 ± 0.4 (88; 0.1-1.7)	0.1 ± 0.1 (47; 0.01-0.8)	0.2 ± 0.1 (42; 0.01-0.6)	0.2 ± 0.4 (2873; <0.01-6.7)
Manganese (µg/g wet)	7.6 ± 4.4 (33; 2.9-17.9)		(8.5 ± 7.4) (112; 0.8-51.3)	(4.4 ± 2.5) (47; 1.0-9.9)	(4.5 ± 4.1) (17; 1.0-12.5)	
Magnesium (µg/g wet)	406 ± 58.5 (33; 320-566)		(344 ± 77.5) (112; 221-645)	(321 ± 62.8) (47; 217-480)	(280 ± 117) (22; 189-634)	
Mercury (µg/g wet)	0.05 ± 0.02 (33; 0.02-0.10)	(0.04 ± 0.03) (0.03; 0.01-0.16)	(0.07 ± 0.05) (112; 0.01-0.24)	(0.13 ± 0.09) (47; 0.01-0.46)	(0.07 ± 0.05) (75; 0.01-0.22)	(0.15 ± 0.18) (3410; <0.01-4.5)
Nickel (µg/g wet)	0.6 ± 0.9 (33; 0.1-3.6)	(0.3 ± 0.3) (15; 0.03-1.1)	(0.4 ± 0.3) (91; 0.02-2.23)	(0.7 ± 0.8) (47; 0.2-4.2)	(0.3 ± 0.9) (20; 0.01-3.9)	
Selenium (µg/g wet)	0.5 ± 0.1 (33; 0.3-0.9)	(0.8 ± 0.7) (27; 0.2-3.3)	(0.4 ± 0.3) (112; 0.07-1.3)	(0.7 ± 0.4) (47; 0.2-1.9)	(0.3 ± 0.2) (75; <0.01-0.9)	(0.5 ± 0.5) (2205; 0.03-6.6)
Strontium (µg/g wet)	38.1 ± 7.6 (33; 28.3-58.3)		(26.0 ± 15.2) (100; 5.3-68.4)	(33.7 ± 17.3) (47; 5.9-78.4)	(41.5 ± 22.7) (23; 13.5-108)	
Vanadium (µg/g wet)	0.3 ± 0.2 (33; 0.1-0.8)		(0.4 ± 0.3) (63; 0.03-1.7)	(0.1 ± 0.1) (47; 0.05-0.4)	(0.2 ± 0.2) (20; 0.03-0.8)	
Zinc (µg/g wet)	37.9 ± 5.7 (33; 29.8-57.0)	(59.2 ± 57.7) (27; 21.3-230)	(32.5 ± 16.3) (112; 9.8-83.2)	(39.3 ± 23.9) (47; 11.1-83.6)	(12.9 ± 13.2) (53; 0.9-76.5)	(29.8 ± 22.8) (1255; 7.7-168)
Moisture (percent)	71.9 ± 1.9 (33; 66.5-76.6)	(77.8 ± 2.7) (7; 75-83)	(73.0 ± 4.9) (112; 56.5-82.6)	(72.9 ± 3.0) (47; 65.3-79.8)		(72.1 ± 4.5) (1421; 43.5-86.2)

Table 5. Carcass concentrations of organic chemicals and lipid content in Rio Grande silvery minnow collected from 6 sites along the Middle Rio Grande, New Mexico, during 2006-2008, (Table 1, Figure 1) and grouped into river sections including the Upper Sites, Middle Sites, and Lower Sites, as well as all data combined.

[Organic chemical concentration data are mean ± 1 standard deviation with the number of fish composite samples and range of concentrations in parenthesis. Means within a row that have no upper case letters are not significantly different. Means within a row with upper case letters and sharing the same upper case letter are not significantly (P<0.05) different. "±", plus or minus; "MRG", Middle Rio Grande; "NM", New Mexico; Site 1. MRG at Bernalillo, NM; Site 2. MRG at Alameda, NM; Site 3. MRG at Los Padillas, NM; Site 4. MRG at Los Lunas, NM; Site 5. MRG at La Joya, NM; Site 6. MRG near San Antonio, NM; µg/kg, microgram per kilogram; wet weight; <, less than]

Organic Chemical (µg/kg wet weight)	Upper Sites (Site 1 & Site 2 combined)	Middle Sites (Site 3 & Site 4 combined)	Lower Sites (Site 5 & Site 6 combined)	All Sites Combined
Lipid (percent)	5.9 ± 1.3 (10; 4.1-7.7)	7.0 ± 1.7 (10; 5.0-10.8)	6.9 ± 1.1 (7; 5.6-8.0)	6.9 ± 1.4 (27; 4.1-10.8)
alpha chlordane	1.5 ± 0.9^{A} (4; 0.3-2.5)	7.1 ± 1.6^{B} (4; 5.7-9.4)	2.3 ± 1.3^{AB} (4; 0.4-3.2)	3.6 ± 2.9 (12; 0.3-9.4)
cis-nonachlor	0.7 ± 0.4^{A} (4; 0.2-1.2)	3.0 ± 0.6^{B} (4; 2.2-3.4)	1.1 ± 0.7^{AB} (4; 0.3-1.7)	1.6 ± 1.1 (12; 0.2-3.4)
dieldrin	0.5 ± 0.3^{A} (4; 0.1-0.9)	3.3 ± 0.8^{B} (4; 2.5-4.2)	1.5 ± 1.0^{AB} (4; 0.4-2.8)	1. 8 ± 1.4 (12; 0.1-4.2)
o,p'-DDE	0.3 ± 0.2 (4; 0.1-0.5)	0.5 ± 0.2 (4; 0.4-0.7)	0.4 ± 0.3 (4; 0.2-0.9)	0.4 ± 0.2 (12; 0.1-0.9)
p,p'-DDD	1.2 ± 0.5^{A} (4; 0.7-1.8)	2.4 ± 0.6^{A} (4; 1.8-3.0)	1.2 ± 0.4^{A} (4; 0.8-1.6)	1.6 ± 0.7 (12; 0.7-3.0)
p,p'-DDE	10.2 ± 2.9^{A} (4; 7.8-13.9)	20.8 ± 3.4^{B} (4; 17.4-24.6)	11.6 ± 2.4^{A} (4; 8.7-13.7)	14.2 ± 6.0 (12; 7.8-24.6)
p,p'-DDT	0.2 ± 0.06^{A} (4; 0.2-0.3)	0.7 ± 0.1^{B} (4; 0.7-0.8)	0.2 ± 0.1^{A} (4; 0.1-0.4)	0.4 ± 0.3 (12; 0.1-0.8)
DDT-total	12.5 ± 4.1^{A} (4; 8.8-17.1)	25.8 ± 4.5^{B} (4; 21.2-30.7)	14.4 ± 2.6^{A} (4; 11.5-17.4)	17.5 ± 7.0 (12; 8.8-30.7)
endosulfan I	1.1 ± 0.7^{A} (4; 0.4-1.7)	3.5 ± 0.8^{B} (4; 2.5-4.2)	1.6 ± 0.8^{AB} (4; 0.6-2.4)	2.1 ± 1.3 (12; 0.3-4.2)
endosulfan sulfate	1.3 ± 0.8^{A} (4; 0.4-2.4)	4.1 ± 0.6^{B} (4; 3.2-4.6)	1.7 ± 1.0^{AB} (4; 0.5-2.8)	2.4 ± 1.5 (12; 0.4-4.6)
gamma chlordane	0.8 ± 0.5^{A} (4; 0.1-1.2)	3.8 ± 1.1^{B} (4; 3.2-5.5)	1.1 ± 0.7^{AB} (4; 0.2-1.6)	1. 9 ± 1.6 (12; 0.1-5.5)
hexachlorobenzene	0.3 ± 0.2 (4; 0.1-0.6)	0.6 ± 0.2 (4; 0.4-0.8)	0.3 ± 0.1 (4; 0.1-0.4)	0.4 ± 0.2 (12; 0.1-0.8)
pentachlorobenezene	2.5 ± 0.2 (4; 2.2-2.6)	3.0 ± 0.2 (4; 2.7-3.2)	3.1 ± 0.5 (4; 2.5-3.8)	2.8 ± 0.4 (12; 2.2-3.8)
pentachloroanisole	0.2 ± 0.2 (4; 0.1-0.5)	0.8 ± 0.3 (4; 0.4-1.1)	0. 6 ± 0.5 (4; 0.3-1.4)	0.5 ± 0.4 (12; 0.1-1.4)
trans-nonachlor	2.1 ± 1.2^{A} (4; 0.4-3.4)	8.7 ± 1.5^{B} (4; 7.21-10.7)	2.8 ± 1.6^{AB} (4; 0.6-4.1)	4.5 ± 3.4 (12; 0.4-10.7)
BDE-100	4.3 ± 2.7^{A} (4; 1.0-6.8)	24.2 ± 15.6^{B} (4; 8.9-44.7)	9.4 ± 6.0^{AB} (3; 3.5-15.5)	12.9 ± 12.9 (11; 1-44.7)
BDE-154	1.2 ± 0.7^{A} (4; 0.4-1.9)	6.9 ± 4.3^{B} (4; 2.7-12.8)	2.7 ± 1.5^{AB} (3; 1.2-4.2)	3.7 ± 3.6 (11; 0.4-12.8)
BDE-206	0.5 ± 0.3 (4; 0.4-0.7)	0.3 ± 0.3 (4; 0.1-0.6)	0.2 ± 0.3 (3; 0.1-0.5)	0.5 ± 0.1 (11; 0.4-0.7)
BDE-28	1.5 ± 1.1 (4; 0.3-2.7)	4.5 ± 3.1 (4; 1.7-8.5)	2.1 ± 1.5 (3; 0.6-3.6)	2.8 ± 2.4 (11; 0.3-8.5)
BDE-47	31.8 ± 21.6^{A} (4; 6.0-50.6)	214 ± 140^{B} (4; 75.1-385)	74.7 ± 50.3^{AB} (3; 26.6-127)	110 ± 117 (11; 6.0-385)

Table 5. Carcass concentrations of organic chemicals and lipid content in Rio Grande silvery minnow collected from 6 sites along the Middle Rio Grande, New Mexico, during 2006-2008, (Table 1, Figure 1) and grouped into river sections including the Upper Sites, Middle Sites, and Lower Sites, as well as all data combined.

[Organic chemical concentration data are mean ± 1 standard deviation with the number of fish composite samples and range of concentrations in parenthesis. Means within a row that have no upper case letters are not significantly different. Means within a row with upper case letters and sharing the same upper case letter are not significantly (P<0.05) different. "±", plus or minus; "MRG", Middle Rio Grande; "NM", New Mexico; Site 1. MRG at Bernalillo, NM; Site 2. MRG at Alameda, NM; Site 3. MRG at Los Padillas, NM; Site 4. MRG at Los Lunas, NM; Site 5. MRG at La Joya, NM; Site 6. MRG near San Antonio, NM; µg/kg, microgram per kilogram; wet weight; <, less than]

Organic Chemical (µg/kg wet weight)	Upper Sites (Site 1 & Site 2 combined)	Middle Sites (Site 3 & Site 4 combined)	Lower Sites (Site 5 & Site 6 combined)	All Sites Combined
BDE-49/71	1.3 ± 1.0 (4;0.1-2.1)	5.9 ± 4.7 (4; 2.0-12.5)	3.8 ± 2.8 (3; 1.0-6.5)	3.6 ± 3.6 (11; 0.1-12.5)
BDE-total	43.4 ± 26.9^{A} (4; 11.0-66.7)	265 ± 171^{B} (4; 94.6-477)	96.7 ± 63.3^{AB} (3; 35.7-162)	138 ± 142 (11; 11.0-477)
PCB-total	8.9 ± 2.4^{A} (4; 5.7-11.1)	26.9 ± 9.0^{B} (4; 16.2-38.2)	10.1 ± 4.3^{A} (4; 4.8-15.0)	15.3 ± 10.1 (12; 4.8-38.2)
PCB-1254	3.8 ± 1.1^{A} (4; 2.6-5.1)	13.8 ± 9.7^{B} (4; 4.9-26.7)	4.2 ± 1.2^{A} (4; 3.2-5.8)	7.3 ± 7.0 (12; 2.6-26.7)
PCB-1260	4.1 ± 3.5 (4; 0.8-7.8)	11.2 ± 6.4 (4; 3.8-19.3)	5.6 ± 4.3 (4; 0.1-10.5)	6.9 ± 5.4 (12; 0.1-19.3)
PCB-105	0.09 ± 0.03^{A} (4; 0.06-0.12)	0.32 ± 0.23^{A} (4; 0.11-0.63)	0.10 ± 0.04^{A} (4; 0.05-0.13)	0.17 ± 0.17 (12; 0.05-0.63)
PCB-129/138/163	0.67 ± 0.29^{A} (4; 0.26-0.89)	2.40 ± 0.90^{B} (4; 1.3-3.5)	0.88 ± 0.43^{A} (4; 0.35-1.36)	1.32 ± 0.97 (12; 0.27-3.5)
PCB-132	0.14 ± 0.07^{A} (4; 0.05-0.22)	0.57 ± 0.24^{B} (4; 0.31-0.88)	0.20 ± 0.09^{A} (4; 0.09-0.29)	0.31 ± 0.24 (12; 0.05-0.88)
PCB-141	0.12 ± 0.07^{A} (4; 0.05-0.22)	0.48 ± 0.14^{B} (4; 0.31-0.65)	0.17 ± 0.08^{A} (4; 0.06-0.25)	0.25 ± 0.19 (12; 0.05-0.65)
PCB-153/168	0.7 ± 0.4^{A} (4; 0.3-1.0)	2.3 ± 0.7^{B} (4; 1.4-3.1)	0.9 ± 0.5^{AB} (4; 0.4-1.4)	1.3 ± 0.9 (12; 0.3-3.1)
PCB-180/193	0.49 ± 0.34^{A} (4; 0.222-0.981)	2.11 ± 0.55^{B} (4; 1.4-2.66)	0.75 ± 0.47^{AB} (4; 0.18-1.32)	1.12 ± 0.85 (12; 0.18-2.66)
PCB-187	0.28 ± 0.17^{A} (4; 0.11-0.47)	0.94 ± 0.25^{B} (4; 0.66-1.24)	0.38 ± 0.23^{AB} (4; 0.11-0.67)	0.53 ± 0.37 (12; 0.11-1.24)
PCB-2	0.003 ± 0.0008^{ABC} (4; 0.003-0.005)	0.004 ± 0.0002^{AB} (4; 0.004-0.004)	0.003 ± 0.0003^{AC} (4; 0.002-0.003)	0.003 ± 0.001 (12; 0.002-0.005)
PCB-52	0.16 ± 0.03^{A} (4; 0.12-0.18)	0.38 ± 0.21^{B} (4; 0.23-0.69)	0.13 ± 0.03^{A} (4; 0.09-0.16)	0.22 ± 0.16 (12; 0.10-0.69)
PCB-83/99	0.14 ± 0.11 (4; 0.05-0.28)	0.43 ± 0.29 (4; 0.17-0.83)	0.14 ± 0.04 (4; 0.09-0.18)	0.24 ± 0.22 (12; 0.05-0.83)
PCB-190	<0.10 ± 0.0 (4; <0.10-<0.10)	0.17 ± 0.04 (4; 0.12-0.21)	0.06 ± 0.04 (4; <0.10-0.11)	0.09 ± 0.07 (12; <0.10-0.21)
n-decane	124 ± 107 (2; 47.9-199)	68 ± 2.5 (2; 66.0-69.5)	90 (1)	95 ± 60.3 (5; 47.9-199)
n-hexacosane	153 ± 89 (2; 90.7-216)	429 ± 165 (2; 312-545)	567 (1)	346 ± 207 (5; 90.7-567)
n-hexadecane	304 ± 46.7 (2; 271-337)	815 ± 48.8 (2; 780-849)	517 (1)	551 ± 258 (5; 271-849)
n-nonadecane	319 ± 141 (2; 219-419)	2747 ± 3639 (2; 173-5320)	825 (1)	1391 ± 2211 (5; 173-5320)
n-octadecane	3055 ± 431 (2; 2750-3360)	5855 ± 1506 (2; 4790-6920)	4660 (1)	4496 ± 1607 (5; 2750-6920)

Table 5. Carcass concentrations of organic chemicals and lipid content in Rio Grande silvery minnow collected from 6 sites along the Middle Rio Grande, New Mexico, during 2006-2008, (Table 1, Figure 1) and grouped into river sections including the Upper Sites, Middle Sites, and Lower Sites, as well as all data combined.

[Organic chemical concentration data are mean ± 1 standard deviation with the number of fish composite samples and range of concentrations in parenthesis. Means within a row that have no upper case letters are not significantly different. Means within a row with upper case letters and sharing the same upper case letter are not significantly (P<0.05) different. "±", plus or minus; "MRG", Middle Rio Grande; "NM", New Mexico; Site 1. MRG at Bernalillo, NM; Site 2. MRG at Alameda, NM; Site 3. MRG at Los Padillas, NM; Site 4. MRG at Los Lunas, NM; Site 5. MRG at La Joya, NM; Site 6. MRG near San Antonio, NM; µg/kg, microgram per kilogram; wet weight; <, less than]

Organic Chemical (µg/kg wet weight)	Upper Sites (Site 1 & Site 2 combined)	Middle Sites (Site 3 & Site 4 combined)	Lower Sites (Site 5 & Site 6 combined)	All Sites Combined
n-pentacosane	779 ± 639 (2; 327-1230)	640 ± 291 (2; 434-846)	353 (1)	638 ± 392 (5; 327-1230)
n-pentadecane	4380 ± 1739 (2; 3150-5610)	8945 ± 3189 (2; 6690-11200)	2870 (1)	5904 ± 3374 (5; 2870-11200)
n-tetracosane	387 ± 248 (2; 211-562)	79 ± 16 (2; 67.1-90.2)	134 (1)	213 ± 203 (5; 67.1-562)
n-tridecane	251 ± 117 (2; 168-333)	745 ± 237 (2; 577-912)	1090 (1)	616 ± 385 (5; 168-1090)
n-undecane	9271 ± 13052 (2; 41-18500)	83 ± 15 (2; 73-93)	91 (1)	3760 ± 8240 (5; 41-18500)
phytane	190 ± 21 (2; 175-204)	395 ± 47 (2; 362-428)	278 (1)	289 ± 106 (5; 175-428)
pristane	184 ± 73 (2; 132-235)	280 ± 40 (2; 251-308)	210 (1)	227 ± 64 (5; 132-308)
acenaphthalene	0. 5 ± 0.1 (2; 0.4-0.5)	1.2 ± 0.2 (2; 1-1.3)	0.7 (1)	0.8 ± 0.4 (5; 0.4-1.3)
acenaphthene	0.2 ± 0 (2; 0.2-0.2)	0.7 ± 0 (2; 0.7-0.7)	0.5 (1)	0.5 ± 0.3 (5; 0.2-0.7)
anthracene	0.3 ± 0 (2; 0.3-0.3)	0.5 ± 0.1 (2; 0.4-0.5)	0.3 (1)	0.4 ± 0.1 (5; 0.3-0.5)
chrysene	0.5 ± 0 (2; 0.5-0.5)	0.7 ± 0.3 (2; 0.5-0.9)	0.2 (1)	0.5 ± 0.2 (5; 0.2-0.9)
fluoranthene	1.4 ± 0.6 (2; 1.0-1.8)	2 ± 1.1 (2; 1.2-2.8)	0.5 (1)	1.5 ± 0.9 (5; 0.5-2.8)
fluorene	1.4 ± 0.2 (2; 1.2-1.5)	2.1 ± 0.2 (2; 1.9-2.2)	1.4 (1)	1.6 ± 0.4 (5; 1.2-2.2)
naphthalene	13.5 ± 0.6 (2; 13.0-13.9)	15.1 ± 0.42 (2; 14.8-15.4)	17.3 (1)	14.9 ± 1.6 (5; 13.0-17.3)
C1-naphthalenes	5.9 ± 1.3 (2; 5.0-6.8)	6.0 ± 0 (2; 6.0-6.0)	3.8 (1)	5.5 ± 1.2 (5; 3.8-6.8)
C2-naphthalenes	3.0 ± 1.0 (2; 2.3-3.7)	3.7 ± 0.14 (2; 3.6-3.8)	2.7 (1)	3.2 ± 0.7 (5; 2.3-3.8)
C3-naphthalenes	1.0 ± 0.01 (2; 1.0-1.0)	2.8 ± 0.21 (2; 2.6-2.9)	1.0 (1)	1.7 ± 1.0 (5; 1.0-2.9)
phenanthrene	3.8 ± 1.3 (2; 2.9-4.7)	4.7 ± 0.4 (2; 4.4-5)	1.9 (1)	3.8 ± 1.3 (5; 1.9-5)
pyrene	0.7 ± 0.1 (2; 0.6-0.8)	1.3 ± 0.8 (2; 0.7-1.8)	0.2 (1)	0.8 ± 0.6 (5; 0.2-1.8)

Appendix 3A. Inorganic Analytical Results for Rio Grande Silvery Minnow Health Study Catalog 2020128 by Trace Element Research Laboratory, College Station, Texas.

Analytical Results Report TOC

1. ECDMS Analytical Results Report 5/27/2010

Catalog Number	Purchase Order Number	Lab ID	Catalog Submitter	ECDMS User ID
2020128	94420-09-Y004	TERL	Lusk, Joel - Albuquerque, NM	r2alfo

Catalog Title	Rio Grande Silvery Minnow Health Study
Lab Name:	Trace Element Research Laboratory

Notes, Symbols and Abbreviations Used
Based on the report options selected the report should be **printed in landscape mode**
Notes, Symbols and Abbreviations Used The following may appear before a reported result (e.g. < 1234). < - Less than symbol indicates that the actual result is less than the reported detection limit. > - Greater than symbol indicates that the actual result is greater than the reported result.
All results are reported as 3 significant digits.
All results are reported as parts per million (ppm), or percent, unless otherwise noted.

1. Integrity Report

Lab Receipt Date	10/01/2009	Lab Approval Date	10/01/2009

Catalog Problems
No problems reported
Problem Resolution

2. Bulk Data

Sample Number	Sample Matrix	Sample Weight (grams)	Percent Moisture
1-07-2	Partial Carcass	34.582	76.2
1-07-3	Partial Carcass	87.213	76.6
1-07-5	Partial Carcass	33.67	72.2
1-07-6	Partial Carcass	102.092	72.9
1-08-1	Partial Carcass	4.752	75.2
1-08-2	Partial Carcass	4.685	74.7
1-08-3	Partial Carcass	4.725	73.2
1-08-4	Partial Carcass	4.509	72.6
1-08-5	Partial Carcass	65.723	72.7
1-08-6	Partial Carcass	4.595	70.7
2-07-4	Partial Carcass	103.398	72.2
4-07-1	Partial Carcass	56.026	74.1
4-07-2	Partial Carcass	37.471	75.3
4-07-3	Partial Carcass	30.856	74.0
4-07-4	Partial Carcass	60.238	74.3
4-07-5	Partial Carcass	27.374	73.5
4-08-1	Partial Carcass	4.255	73.0
4-08-2	Partial Carcass	4.405	72.0
4-08-3	Partial Carcass	4.302	71.0
4-08-4	Partial Carcass	4.676	73.0
4-08-6	Partial Carcass	4.405	73.3
7-06-1	Partial Carcass	3.869	73.2
7-06-2	Partial Carcass	4.069	72.5
7-06-3	Partial Carcass	3.891	71.5
7-06-4	Partial Carcass	4.417	66.5
7-06-5	Partial Carcass	4.199	68.8
7-06-6	Partial Carcass	4.354	69.8
7-08-1	Partial Carcass	62.399	71.6
7-08-2	Partial Carcass	53.332	71.5
7-08-3	Partial Carcass	23.286	72.8
7-08-4	Partial Carcass	35.884	73.5

Sample Number	Sample Matrix	Sample Weight (grams)	Percent Moisture
7-08-5	Partial Carcass	18.078	71.5
7-08-6	Partial Carcass	23.443	71.3
06385810	Water	125	

4. Contaminant Concentrations

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
06385810						
	Aluminum	Water			0.905	0.00500
	Arsenic	Water			0.00440	0.0000500
	Boron	Water			0.116	0.00500
	Barium	Water			0.106	0.00100
	Beryllium	Water			< 0.000500	0.000500
	Cadmium	Water			0.000260	0.0000100
	Chromium	Water			< 0.00500	0.00500
	Copper	Water			0.0130	0.00500
	Iron	Water			1.04	0.0100
	Mercury	Water			0.00000300	0.00000100
	Magnesium	Water			4.48	0.0200
	Manganese	Water			0.0750	0.00200
	Molybdenum	Water			< 0.0100	0.0100
	Nickel	Water			0.00800	0.00500
	Lead	Water			0.0120	0.0000500
	Selenium	Water			0.000200	0.000100
	Strontium	Water			0.264	0.000500
	Vanadium	Water			0.00700	0.00500
	Zinc	Water			0.0700	0.00200
1-07-2						
	Aluminum	Partial Carcass	141	0.462	33.6	0.110
	Arsenic	Partial Carcass	3.50	0.0462	0.833	0.0110
	Boron	Partial Carcass	< 0.462	0.462	< 0.110	0.110
	Barium	Partial Carcass	79.3	0.0920	18.9	0.0219
	Beryllium	Partial Carcass	< 0.0462	0.0462	< 0.0110	0.0110
	Cadmium	Partial Carcass	0.0695	0.00924	0.0165	0.00220

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Chromium	Partial Carcass	< 0.462	0.462	< 0.110	0.110
	Copper	Partial Carcass	3.30	0.462	0.785	0.110
	Iron	Partial Carcass	203	0.920	48.3	0.219
	Mercury	Partial Carcass	0.315	0.00510	0.0750	0.00121
	Magnesium	Partial Carcass	1550	1.85	369	0.440
	Manganese	Partial Carcass	24.0	0.185	5.71	0.0440
	Molybdenum	Partial Carcass	< 0.920	0.920	< 0.219	0.219
	Nickel	Partial Carcass	0.472	0.462	0.112	0.110
	Lead	Partial Carcass	0.416	0.0462	0.0990	0.0110
	Selenium	Partial Carcass	2.16	0.0462	0.514	0.0110
	Strontium	Partial Carcass	176	0.0462	41.9	0.0110
	Vanadium	Partial Carcass	1.26	0.462	0.300	0.110
	Zinc	Partial Carcass	179	0.185	42.6	0.0440
	Methyl Mercury	Partial Carcass	0.221	0.00845	0.0526	0.00201
1-07-3						
	Aluminum	Partial Carcass	95.6	0.431	22.4	0.101
	Arsenic	Partial Carcass	3.43	0.0431	0.803	0.0101
	Boron	Partial Carcass	< 0.431	0.431	< 0.101	0.101
	Barium	Partial Carcass	68.0	0.0860	15.9	0.0201
	Beryllium	Partial Carcass	< 0.0431	0.0431	< 0.0101	0.0101
	Cadmium	Partial Carcass	0.0371	0.00862	0.00868	0.00202
	Chromium	Partial Carcass	< 0.431	0.431	< 0.101	0.101
	Copper	Partial Carcass	3.20	0.431	0.749	0.101
	Iron	Partial Carcass	151	0.860	35.3	0.201
	Mercury	Partial Carcass	0.296	0.00480	0.0693	0.00112
	Magnesium	Partial Carcass	1590	1.72	372	0.402
	Manganese	Partial Carcass	16.4	0.172	3.84	0.0402
	Molybdenum	Partial Carcass	< 0.860	0.860	< 0.201	0.201
	Nickel	Partial Carcass	0.537	0.431	0.126	0.101

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Lead	Partial Carcass	0.395	0.0431	0.0924	0.0101
	Selenium	Partial Carcass	2.52	0.0431	0.590	0.0101
	Strontium	Partial Carcass	147	0.0431	34.4	0.0101
	Vanadium	Partial Carcass	0.852	0.431	0.199	0.101
	Zinc	Partial Carcass	159	0.172	37.2	0.0402
	Methyl Mercury	Partial Carcass	0.241	0.0101	0.0564	0.00236
1-07-5						
	Aluminum	Partial Carcass	109	0.425	30.3	0.118
	Arsenic	Partial Carcass	2.96	0.0425	0.823	0.0118
	Boron	Partial Carcass	0.477	0.425	0.133	0.118
	Barium	Partial Carcass	22.1	0.0850	6.14	0.0236
	Beryllium	Partial Carcass	< 0.0425	0.0425	< 0.0118	0.0118
	Cadmium	Partial Carcass	< 0.00851	0.00851	< 0.00237	0.00237
	Chromium	Partial Carcass	< 0.425	0.425	< 0.118	0.118
	Copper	Partial Carcass	2.69	0.425	0.748	0.118
	Iron	Partial Carcass	162	0.850	45.0	0.236
	Mercury	Partial Carcass	0.198	0.00490	0.0550	0.00136
	Magnesium	Partial Carcass	1150	1.70	320.	0.473
	Manganese	Partial Carcass	16.3	0.170	4.53	0.0473
	Molybdenum	Partial Carcass	< 0.850	0.850	< 0.236	0.236
	Nickel	Partial Carcass	0.470	0.425	0.131	0.118
	Lead	Partial Carcass	0.272	0.0425	0.0756	0.0118
	Selenium	Partial Carcass	3.19	0.0425	0.887	0.0118
	Strontium	Partial Carcass	143	0.0425	39.8	0.0118
	Vanadium	Partial Carcass	0.669	0.425	0.186	0.118
	Zinc	Partial Carcass	114	0.170	31.7	0.0473
	Methyl Mercury	Partial Carcass	0.164	0.00837	0.0456	0.00233
1-07-6						
	Aluminum	Partial Carcass	127	0.439	34.4	0.119

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Arsenic	Partial Carcass	3.26	0.0439	0.883	0.0119
	Boron	Partial Carcass	< 0.439	0.439	< 0.119	0.119
	Barium	Partial Carcass	37.3	0.0880	10.1	0.0238
	Beryllium	Partial Carcass	< 0.0439	0.0439	< 0.0119	0.0119
	Cadmium	Partial Carcass	< 0.00878	0.00878	< 0.00238	0.00238
	Chromium	Partial Carcass	< 0.439	0.439	< 0.119	0.119
	Copper	Partial Carcass	2.58	0.439	0.699	0.119
	Iron	Partial Carcass	178	0.880	48.2	0.238
	Mercury	Partial Carcass	0.258	0.00460	0.0699	0.00125
	Magnesium	Partial Carcass	1290	1.76	350.	0.477
	Manganese	Partial Carcass	15.6	0.176	4.23	0.0477
	Molybdenum	Partial Carcass	< 0.880	0.880	< 0.238	0.238
	Nickel	Partial Carcass	< 0.439	0.439	< 0.119	0.119
	Lead	Partial Carcass	0.290	0.0439	0.0786	0.0119
	Selenium	Partial Carcass	1.85	0.0439	0.501	0.0119
	Strontium	Partial Carcass	134	0.0439	36.3	0.0119
	Vanadium	Partial Carcass	0.656	0.439	0.178	0.119
	Zinc	Partial Carcass	110.	0.176	29.8	0.0477
	Methyl Mercury	Partial Carcass	0.256	0.00652	0.0694	0.00177
1-08-1						
	Aluminum	Partial Carcass	140.	0.519	34.7	0.129
	Arsenic	Partial Carcass	4.49	0.0490	1.11	0.0122
	Boron	Partial Carcass	< 0.519	0.519	< 0.129	0.129
	Barium	Partial Carcass	88.4	0.104	21.9	0.0258
	Beryllium	Partial Carcass	< 0.0519	0.0519	< 0.0129	0.0129
	Cadmium	Partial Carcass	0.0627	0.0196	0.0155	0.00486
	Chromium	Partial Carcass	0.753	0.519	0.187	0.129
	Copper	Partial Carcass	3.78	0.519	0.937	0.129
	Iron	Partial Carcass	149	1.04	37.0	0.258

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Mercury	Partial Carcass	0.150	0.00610	0.0372	0.00151
	Magnesium	Partial Carcass	1560	2.07	387	0.513
	Manganese	Partial Carcass	17.8	0.207	4.41	0.0513
	Molybdenum	Partial Carcass	< 1.04	1.04	< 0.258	0.258
	Nickel	Partial Carcass	0.741	0.519	0.184	0.129
	Lead	Partial Carcass	0.152	0.0490	0.0377	0.0122
	Selenium	Partial Carcass	2.06	0.0490	0.511	0.0122
	Strontium	Partial Carcass	154	0.0519	38.2	0.0129
	Vanadium	Partial Carcass	0.550	0.519	0.136	0.129
	Zinc	Partial Carcass	165	0.207	40.9	0.0513
1-08-2						
	Aluminum	Partial Carcass	162	0.508	41.0	0.129
	Arsenic	Partial Carcass	3.92	0.0480	0.992	0.0121
	Boron	Partial Carcass	< 0.508	0.508	< 0.129	0.129
	Barium	Partial Carcass	69.3	0.102	17.5	0.0258
	Beryllium	Partial Carcass	< 0.0508	0.0508	< 0.0129	0.0129
	Cadmium	Partial Carcass	0.0359	0.0192	0.00908	0.00486
	Chromium	Partial Carcass	1.23	0.508	0.311	0.129
	Copper	Partial Carcass	46.7	0.508	11.8	0.129
	Iron	Partial Carcass	181	1.02	45.8	0.258
	Mercury	Partial Carcass	0.146	0.00570	0.0369	0.00144
	Magnesium	Partial Carcass	1440	2.03	364	0.514
	Manganese	Partial Carcass	17.0	0.203	4.30	0.0514
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.258	0.258
	Nickel	Partial Carcass	7.50	0.508	1.90	0.129
	Lead	Partial Carcass	0.291	0.0480	0.0736	0.0121
	Selenium	Partial Carcass	2.08	0.0480	0.526	0.0121
	Strontium	Partial Carcass	144	0.0508	36.4	0.0129
	Vanadium	Partial Carcass	0.908	0.508	0.230	0.129

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Zinc	Partial Carcass	138	0.203	34.9	0.0514
1-08-3						
	Aluminum	Partial Carcass	95.0	0.520	25.5	0.139
	Arsenic	Partial Carcass	3.77	0.0490	1.01	0.0131
	Boron	Partial Carcass	< 0.520	0.520	< 0.139	0.139
	Barium	Partial Carcass	62.1	0.104	16.6	0.0279
	Beryllium	Partial Carcass	< 0.0520	0.0520	< 0.0139	0.0139
	Cadmium	Partial Carcass	< 0.0196	0.0196	< 0.00525	0.00525
	Chromium	Partial Carcass	< 0.520	0.520	< 0.139	0.139
	Copper	Partial Carcass	3.47	0.520	0.930	0.139
	Iron	Partial Carcass	125	1.04	33.5	0.279
	Mercury	Partial Carcass	0.125	0.00520	0.0335	0.00139
	Magnesium	Partial Carcass	1400	2.08	375	0.557
	Manganese	Partial Carcass	10.7	0.208	2.87	0.0557
	Molybdenum	Partial Carcass	< 1.04	1.04	< 0.279	0.279
	Nickel	Partial Carcass	0.856	0.520	0.229	0.139
	Lead	Partial Carcass	0.166	0.0490	0.0445	0.0131
	Selenium	Partial Carcass	1.70	0.0490	0.456	0.0131
	Strontium	Partial Carcass	147	0.0520	39.4	0.0139
	Vanadium	Partial Carcass	0.706	0.520	0.189	0.139
	Zinc	Partial Carcass	136	0.208	36.4	0.0557
1-08-4						
	Aluminum	Partial Carcass	142	0.508	38.9	0.139
	Arsenic	Partial Carcass	2.70	0.0480	0.740	0.0132
	Boron	Partial Carcass	< 0.508	0.508	< 0.139	0.139
	Barium	Partial Carcass	56.1	0.102	15.4	0.0279
	Beryllium	Partial Carcass	< 0.0508	0.0508	< 0.0139	0.0139
	Cadmium	Partial Carcass	< 0.0192	0.0192	< 0.00526	0.00526
	Chromium	Partial Carcass	< 0.508	0.508	< 0.139	0.139

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Copper	Partial Carcass	3.76	0.508	1.03	0.139
	Iron	Partial Carcass	178	1.02	48.8	0.279
	Mercury	Partial Carcass	0.0861	0.00870	0.0236	0.00238
	Magnesium	Partial Carcass	1390	2.03	381	0.556
	Manganese	Partial Carcass	12.9	0.203	3.53	0.0556
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.279	0.279
	Nickel	Partial Carcass	0.692	0.508	0.190	0.139
	Lead	Partial Carcass	0.191	0.0480	0.0523	0.0132
	Selenium	Partial Carcass	1.89	0.0480	0.518	0.0132
	Strontium	Partial Carcass	121	0.0508	33.2	0.0139
	Vanadium	Partial Carcass	0.788	0.508	0.216	0.139
	Zinc	Partial Carcass	132	0.203	36.2	0.0556
1-08-5						
	Aluminum	Partial Carcass	36.4	0.459	9.94	0.125
	Arsenic	Partial Carcass	3.10	0.0459	0.846	0.0125
	Boron	Partial Carcass	< 0.459	0.459	< 0.125	0.125
	Barium	Partial Carcass	34.7	0.0920	9.47	0.0251
	Beryllium	Partial Carcass	< 0.0459	0.0459	< 0.0125	0.0125
	Cadmium	Partial Carcass	< 0.00918	0.00918	< 0.00251	0.00251
	Chromium	Partial Carcass	< 0.459	0.459	< 0.125	0.125
	Copper	Partial Carcass	2.89	0.459	0.789	0.125
	Iron	Partial Carcass	66.1	0.920	18.0	0.251
	Mercury	Partial Carcass	0.102	0.00510	0.0278	0.00139
	Magnesium	Partial Carcass	1280	1.84	349	0.502
	Manganese	Partial Carcass	13.1	0.184	3.58	0.0502
	Molybdenum	Partial Carcass	< 0.920	0.920	< 0.251	0.251
	Nickel	Partial Carcass	< 0.459	0.459	< 0.125	0.125
	Lead	Partial Carcass	0.214	0.0459	0.0584	0.0125
	Selenium	Partial Carcass	2.03	0.0459	0.554	0.0125

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Strontium	Partial Carcass	134	0.0459	36.6	0.0125
	Vanadium	Partial Carcass	< 0.459	0.459	< 0.125	0.125
	Zinc	Partial Carcass	112	0.184	30.6	0.0502
	Methyl Mercury	Partial Carcass	0.0995	0.00755	0.0272	0.00206
1-08-6						
	Aluminum	Partial Carcass	73.3	0.502	21.5	0.147
	Arsenic	Partial Carcass	2.73	0.0474	0.800	0.0139
	Boron	Partial Carcass	< 0.502	0.502	< 0.147	0.147
	Barium	Partial Carcass	22.1	0.100	6.48	0.0293
	Beryllium	Partial Carcass	< 0.0502	0.0502	< 0.0147	0.0147
	Cadmium	Partial Carcass	< 0.0190	0.0190	< 0.00557	0.00557
	Chromium	Partial Carcass	< 0.502	0.502	< 0.147	0.147
	Copper	Partial Carcass	6.91	0.502	2.02	0.147
	Iron	Partial Carcass	101	1.00	29.6	0.293
	Mercury	Partial Carcass	0.102	0.00780	0.0299	0.00229
	Magnesium	Partial Carcass	1240	2.01	363	0.589
	Manganese	Partial Carcass	11.8	0.201	3.46	0.0589
	Molybdenum	Partial Carcass	< 1.00	1.00	< 0.293	0.293
	Nickel	Partial Carcass	3.45	0.502	1.01	0.147
	Lead	Partial Carcass	0.115	0.0474	0.0337	0.0139
	Selenium	Partial Carcass	2.84	0.0474	0.832	0.0139
	Strontium	Partial Carcass	175	0.0502	51.3	0.0147
	Vanadium	Partial Carcass	0.608	0.502	0.178	0.147
	Zinc	Partial Carcass	131	0.201	38.4	0.0589
2-07-4						
	Aluminum	Partial Carcass	61.3	0.434	17.0	0.121
	Arsenic	Partial Carcass	3.13	0.0434	0.870	0.0121
	Boron	Partial Carcass	< 0.434	0.434	< 0.121	0.121
	Barium	Partial Carcass	47.8	0.0870	13.3	0.0242

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Beryllium	Partial Carcass	< 0.0434	0.0434	< 0.0121	0.0121
	Cadmium	Partial Carcass	< 0.00867	0.00867	< 0.00241	0.00241
	Chromium	Partial Carcass	< 0.434	0.434	< 0.121	0.121
	Copper	Partial Carcass	2.67	0.434	0.742	0.121
	Iron	Partial Carcass	106	0.870	29.5	0.242
	Mercury	Partial Carcass	0.141	0.00450	0.0392	0.00125
	Magnesium	Partial Carcass	1330	1.73	370.	0.481
	Manganese	Partial Carcass	12.5	0.173	3.48	0.0481
	Molybdenum	Partial Carcass	< 0.870	0.870	< 0.242	0.242
	Nickel	Partial Carcass	< 0.434	0.434	< 0.121	0.121
	Lead	Partial Carcass	0.326	0.0434	0.0906	0.0121
	Selenium	Partial Carcass	1.81	0.0434	0.503	0.0121
	Strontium	Partial Carcass	108	0.0434	30.0	0.0121
	Vanadium	Partial Carcass	< 0.434	0.434	< 0.121	0.121
	Zinc	Partial Carcass	114	0.173	31.7	0.0481
	Methyl Mercury	Partial Carcass	0.141	0.00762	0.0392	0.00212
4-07-1						
	Aluminum	Partial Carcass	382	0.511	98.9	0.132
	Arsenic	Partial Carcass	3.02	0.0482	0.782	0.0125
	Boron	Partial Carcass	< 0.511	0.511	< 0.132	0.132
	Barium	Partial Carcass	79.0	0.102	20.5	0.0264
	Beryllium	Partial Carcass	< 0.0511	0.0511	< 0.0132	0.0132
	Cadmium	Partial Carcass	< 0.0193	0.0193	< 0.00500	0.00500
	Chromium	Partial Carcass	0.618	0.511	0.160	0.132
	Copper	Partial Carcass	3.49	0.511	0.904	0.132
	Iron	Partial Carcass	457	1.02	118	0.264
	Mercury	Partial Carcass	0.282	0.00460	0.0730	0.00119
	Magnesium	Partial Carcass	1730	2.04	448	0.528
	Manganese	Partial Carcass	32.4	0.204	8.39	0.0528

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.264	0.264
	Nickel	Partial Carcass	1.24	0.511	0.321	0.132
	Lead	Partial Carcass	0.376	0.0482	0.0974	0.0125
	Selenium	Partial Carcass	2.00	0.0482	0.518	0.0125
	Strontium	Partial Carcass	138	0.0511	35.7	0.0132
	Vanadium	Partial Carcass	1.32	0.511	0.342	0.132
	Zinc	Partial Carcass	140.	0.204	36.3	0.0528
4-07-2						
	Aluminum	Partial Carcass	429	0.513	106	0.127
	Arsenic	Partial Carcass	3.29	0.0484	0.813	0.0120
	Boron	Partial Carcass	< 0.513	0.513	< 0.127	0.127
	Barium	Partial Carcass	70.0	0.103	17.3	0.0254
	Beryllium	Partial Carcass	< 0.0513	0.0513	< 0.0127	0.0127
	Cadmium	Partial Carcass	< 0.0194	0.0194	< 0.00479	0.00479
	Chromium	Partial Carcass	0.783	0.513	0.193	0.127
	Copper	Partial Carcass	3.25	0.513	0.803	0.127
	Iron	Partial Carcass	496	1.03	123	0.254
	Mercury	Partial Carcass	0.251	0.00510	0.0620	0.00126
	Magnesium	Partial Carcass	1570	2.05	388	0.506
	Manganese	Partial Carcass	34.1	0.205	8.42	0.0506
	Molybdenum	Partial Carcass	< 1.03	1.03	< 0.254	0.254
	Nickel	Partial Carcass	1.15	0.513	0.284	0.127
	Lead	Partial Carcass	0.522	0.0484	0.129	0.0120
	Selenium	Partial Carcass	1.66	0.0484	0.410	0.0120
	Strontium	Partial Carcass	145	0.0513	35.8	0.0127
	Vanadium	Partial Carcass	1.84	0.513	0.454	0.127
	Zinc	Partial Carcass	135	0.205	33.3	0.0506
4-07-3						
	Aluminum	Partial Carcass	256	0.512	66.6	0.133

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Arsenic	Partial Carcass	2.74	0.0484	0.712	0.0126
	Boron	Partial Carcass	< 0.512	0.512	< 0.133	0.133
	Barium	Partial Carcass	65.2	0.102	17.0	0.0265
	Beryllium	Partial Carcass	< 0.0512	0.0512	< 0.0133	0.0133
	Cadmium	Partial Carcass	< 0.0193	0.0193	< 0.00502	0.00502
	Chromium	Partial Carcass	0.513	0.512	0.133	0.133
	Copper	Partial Carcass	2.79	0.512	0.725	0.133
	Iron	Partial Carcass	290.	1.02	75.4	0.265
	Mercury	Partial Carcass	0.202	0.00470	0.0525	0.00122
	Magnesium	Partial Carcass	1390	2.05	361	0.533
	Manganese	Partial Carcass	24.3	0.205	6.32	0.0533
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.265	0.265
	Nickel	Partial Carcass	0.917	0.512	0.238	0.133
	Lead	Partial Carcass	0.290	0.0484	0.0754	0.0126
	Selenium	Partial Carcass	1.31	0.0484	0.341	0.0126
	Strontium	Partial Carcass	150.	0.0512	39.0	0.0133
	Vanadium	Partial Carcass	1.49	0.512	0.387	0.133
	Zinc	Partial Carcass	140.	0.205	36.4	0.0533
4-07-4						
	Aluminum	Partial Carcass	450.	0.507	116	0.130
	Arsenic	Partial Carcass	3.09	0.0478	0.794	0.0123
	Boron	Partial Carcass	< 0.507	0.507	< 0.130	0.130
	Barium	Partial Carcass	59.0	0.101	15.2	0.0260
	Beryllium	Partial Carcass	< 0.0507	0.0507	< 0.0130	0.0130
	Cadmium	Partial Carcass	< 0.0191	0.0191	< 0.00491	0.00491
	Chromium	Partial Carcass	0.752	0.507	0.193	0.130
	Copper	Partial Carcass	3.57	0.507	0.917	0.130
	Iron	Partial Carcass	511	1.01	131	0.260
	Mercury	Partial Carcass	0.161	0.00490	0.0414	0.00126

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Magnesium	Partial Carcass	1710	2.03	439	0.522
	Manganese	Partial Carcass	28.4	0.203	7.30	0.0522
	Molybdenum	Partial Carcass	< 1.01	1.01	< 0.260	0.260
	Nickel	Partial Carcass	1.26	0.507	0.324	0.130
	Lead	Partial Carcass	0.557	0.0478	0.143	0.0123
	Selenium	Partial Carcass	2.07	0.0478	0.532	0.0123
	Strontium	Partial Carcass	129	0.0507	33.2	0.0130
	Vanadium	Partial Carcass	1.56	0.507	0.401	0.130
	Zinc	Partial Carcass	126	0.203	32.4	0.0522
4-07-5						
	Aluminum	Partial Carcass	201	0.493	53.3	0.131
	Arsenic	Partial Carcass	2.79	0.0493	0.739	0.0131
	Boron	Partial Carcass	0.758	0.493	0.201	0.131
	Barium	Partial Carcass	29.7	0.0990	7.87	0.0262
	Beryllium	Partial Carcass	< 0.0493	0.0493	< 0.0131	0.0131
	Cadmium	Partial Carcass	< 0.00987	0.00987	< 0.00262	0.00262
	Chromium	Partial Carcass	< 0.493	0.493	< 0.131	0.131
	Copper	Partial Carcass	3.00	0.493	0.795	0.131
	Iron	Partial Carcass	294	0.990	77.9	0.262
	Mercury	Partial Carcass	0.216	0.00490	0.0572	0.00130
	Magnesium	Partial Carcass	1360	1.97	360.	0.522
	Manganese	Partial Carcass	35.1	0.197	9.30	0.0522
	Molybdenum	Partial Carcass	< 0.990	0.990	< 0.262	0.262
	Nickel	Partial Carcass	< 0.493	0.493	< 0.131	0.131
	Lead	Partial Carcass	0.562	0.0493	0.149	0.0131
	Selenium	Partial Carcass	2.33	0.0493	0.617	0.0131
	Strontium	Partial Carcass	155	0.0493	41.1	0.0131
	Vanadium	Partial Carcass	0.977	0.493	0.259	0.131
	Zinc	Partial Carcass	125	0.197	33.1	0.0522

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
4-08-1						
	Aluminum	Partial Carcass	383	0.509	103	0.137
	Arsenic	Partial Carcass	3.41	0.0480	0.921	0.0130
	Boron	Partial Carcass	< 0.509	0.509	< 0.137	0.137
	Barium	Partial Carcass	69.5	0.102	18.8	0.0275
	Beryllium	Partial Carcass	< 0.0509	0.0509	< 0.0137	0.0137
	Cadmium	Partial Carcass	0.0436	0.0192	0.0118	0.00518
	Chromium	Partial Carcass	0.925	0.509	0.250	0.137
	Copper	Partial Carcass	3.34	0.509	0.902	0.137
	Iron	Partial Carcass	408	1.02	110.	0.275
	Mercury	Partial Carcass	0.129	0.00570	0.0348	0.00154
	Magnesium	Partial Carcass	1410	2.03	381	0.548
	Manganese	Partial Carcass	27.1	0.203	7.32	0.0548
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.275	0.275
	Nickel	Partial Carcass	1.25	0.509	0.338	0.137
	Lead	Partial Carcass	0.298	0.0480	0.0805	0.0130
	Selenium	Partial Carcass	1.36	0.0480	0.367	0.0130
	Strontium	Partial Carcass	107	0.0509	28.9	0.0137
	Vanadium	Partial Carcass	1.30	0.509	0.351	0.137
	Zinc	Partial Carcass	136	0.203	36.7	0.0548
4-08-2						
	Aluminum	Partial Carcass	317	0.511	88.8	0.143
	Arsenic	Partial Carcass	3.70	0.0482	1.04	0.0135
	Boron	Partial Carcass	< 0.511	0.511	< 0.143	0.143
	Barium	Partial Carcass	67.7	0.102	19.0	0.0286
	Beryllium	Partial Carcass	< 0.0511	0.0511	< 0.0143	0.0143
	Cadmium	Partial Carcass	< 0.0193	0.0193	< 0.00540	0.00540
	Chromium	Partial Carcass	0.662	0.511	0.185	0.143
	Copper	Partial Carcass	3.17	0.511	0.888	0.143

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Iron	Partial Carcass	344	1.02	96.3	0.286
	Mercury	Partial Carcass	0.123	0.00570	0.0344	0.00160
	Magnesium	Partial Carcass	1470	2.04	412	0.571
	Manganese	Partial Carcass	22.9	0.204	6.41	0.0571
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.286	0.286
	Nickel	Partial Carcass	0.949	0.511	0.266	0.143
	Lead	Partial Carcass	0.289	0.0482	0.0809	0.0135
	Selenium	Partial Carcass	1.81	0.0482	0.507	0.0135
	Strontium	Partial Carcass	131	0.0511	36.7	0.0143
	Vanadium	Partial Carcass	1.13	0.511	0.316	0.143
	Zinc	Partial Carcass	122	0.204	34.2	0.0571
4-08-3						
	Aluminum	Partial Carcass	111	0.508	32.2	0.147
	Arsenic	Partial Carcass	2.67	0.0480	0.774	0.0139
	Boron	Partial Carcass	< 0.508	0.508	< 0.147	0.147
	Barium	Partial Carcass	49.8	0.102	14.4	0.0296
	Beryllium	Partial Carcass	< 0.0508	0.0508	< 0.0147	0.0147
	Cadmium	Partial Carcass	< 0.0192	0.0192	< 0.00557	0.00557
	Chromium	Partial Carcass	< 0.508	0.508	< 0.147	0.147
	Copper	Partial Carcass	20.0	0.508	5.80	0.147
	Iron	Partial Carcass	141	1.02	40.9	0.296
	Mercury	Partial Carcass	0.106	0.00650	0.0307	0.00188
	Magnesium	Partial Carcass	1310	2.03	380.	0.589
	Manganese	Partial Carcass	13.0	0.203	3.77	0.0589
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.296	0.296
	Nickel	Partial Carcass	4.58	0.508	1.33	0.147
	Lead	Partial Carcass	0.305	0.0480	0.0884	0.0139
	Selenium	Partial Carcass	1.68	0.0480	0.487	0.0139
	Strontium	Partial Carcass	109	0.0508	31.6	0.0147

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Vanadium	Partial Carcass	0.705	0.508	0.204	0.147
	Zinc	Partial Carcass	119	0.203	34.5	0.0589
4-08-4						
	Aluminum	Partial Carcass	214	0.512	57.8	0.138
	Arsenic	Partial Carcass	2.78	0.0483	0.751	0.0130
	Boron	Partial Carcass	< 0.512	0.512	< 0.138	0.138
	Barium	Partial Carcass	55.3	0.102	14.9	0.0275
	Beryllium	Partial Carcass	< 0.0512	0.0512	< 0.0138	0.0138
	Cadmium	Partial Carcass	< 0.0193	0.0193	< 0.00521	0.00521
	Chromium	Partial Carcass	0.602	0.512	0.163	0.138
	Copper	Partial Carcass	14.9	0.512	4.02	0.138
	Iron	Partial Carcass	233	1.02	62.9	0.275
	Mercury	Partial Carcass	0.111	0.00560	0.0300	0.00151
	Magnesium	Partial Carcass	1460	2.05	394	0.554
	Manganese	Partial Carcass	17.2	0.205	4.64	0.0554
	Molybdenum	Partial Carcass	< 1.02	1.02	< 0.275	0.275
	Nickel	Partial Carcass	3.27	0.512	0.883	0.138
	Lead	Partial Carcass	0.293	0.0483	0.0791	0.0130
	Selenium	Partial Carcass	1.53	0.0483	0.413	0.0130
	Strontium	Partial Carcass	124	0.0512	33.5	0.0138
	Vanadium	Partial Carcass	0.940	0.512	0.254	0.138
	Zinc	Partial Carcass	127	0.205	34.3	0.0554
4-08-6						
	Aluminum	Partial Carcass	95.3	0.506	25.4	0.135
	Arsenic	Partial Carcass	2.67	0.0477	0.713	0.0127
	Boron	Partial Carcass	< 0.506	0.506	< 0.135	0.135
	Barium	Partial Carcass	30.6	0.101	8.17	0.0270
	Beryllium	Partial Carcass	< 0.0506	0.0506	< 0.0135	0.0135
	Cadmium	Partial Carcass	< 0.0191	0.0191	< 0.00510	0.00510

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Chromium	Partial Carcass	< 0.506	0.506	< 0.135	0.135
	Copper	Partial Carcass	3.01	0.506	0.804	0.135
	Iron	Partial Carcass	163	1.01	43.5	0.270
	Mercury	Partial Carcass	0.120	0.00600	0.0320	0.00160
	Magnesium	Partial Carcass	1450	2.02	387	0.539
	Manganese	Partial Carcass	15.8	0.202	4.22	0.0539
	Molybdenum	Partial Carcass	< 1.01	1.01	< 0.270	0.270
	Nickel	Partial Carcass	0.639	0.506	0.171	0.135
	Lead	Partial Carcass	0.120	0.0477	0.0320	0.0127
	Selenium	Partial Carcass	1.69	0.0477	0.451	0.0127
	Strontium	Partial Carcass	186	0.0506	49.7	0.0135
	Vanadium	Partial Carcass	0.789	0.506	0.211	0.135
	Zinc	Partial Carcass	133	0.202	35.5	0.0539
7-06-1						
	Aluminum	Partial Carcass	1150	0.522	308	0.140
	Arsenic	Partial Carcass	3.27	0.0493	0.876	0.0132
	Boron	Partial Carcass	1.05	0.522	0.281	0.140
	Barium	Partial Carcass	88.0	0.104	23.6	0.0279
	Beryllium	Partial Carcass	0.0578	0.0522	0.0155	0.0140
	Cadmium	Partial Carcass	0.115	0.0197	0.0308	0.00528
	Chromium	Partial Carcass	3.73	0.522	1.00	0.140
	Copper	Partial Carcass	31.7	0.522	8.50	0.140
	Iron	Partial Carcass	1180	1.04	316	0.279
	Mercury	Partial Carcass	0.277	0.00670	0.0742	0.00180
	Magnesium	Partial Carcass	2000	2.09	536	0.560
	Manganese	Partial Carcass	58.2	0.209	15.6	0.0560
	Molybdenum	Partial Carcass	< 1.04	1.04	< 0.279	0.279
	Nickel	Partial Carcass	13.3	0.522	3.56	0.140
	Lead	Partial Carcass	1.12	0.0493	0.300	0.0132

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Selenium	Partial Carcass	2.09	0.0493	0.560	0.0132
	Strontium	Partial Carcass	162	0.0522	43.4	0.0140
	Vanadium	Partial Carcass	3.15	0.522	0.844	0.140
	Zinc	Partial Carcass	156	0.209	41.8	0.0560
7-06-2						
	Aluminum	Partial Carcass	970.	0.517	267	0.142
	Arsenic	Partial Carcass	3.08	0.0488	0.847	0.0134
	Boron	Partial Carcass	0.705	0.517	0.194	0.142
	Barium	Partial Carcass	74.3	0.103	20.4	0.0283
	Beryllium	Partial Carcass	< 0.0517	0.0517	< 0.0142	0.0142
	Cadmium	Partial Carcass	0.0542	0.0195	0.0149	0.00536
	Chromium	Partial Carcass	2.63	0.517	0.723	0.142
	Copper	Partial Carcass	18.2	0.517	5.00	0.142
	Iron	Partial Carcass	1060	1.03	292	0.283
	Mercury	Partial Carcass	0.273	0.00640	0.0751	0.00176
	Magnesium	Partial Carcass	1700	2.07	468	0.569
	Manganese	Partial Carcass	52.4	0.207	14.4	0.0569
	Molybdenum	Partial Carcass	< 1.03	1.03	< 0.283	0.283
	Nickel	Partial Carcass	12.9	0.517	3.55	0.142
	Lead	Partial Carcass	1.47	0.0488	0.404	0.0134
	Selenium	Partial Carcass	2.42	0.0488	0.666	0.0134
	Strontium	Partial Carcass	123	0.0517	33.8	0.0142
	Vanadium	Partial Carcass	2.48	0.517	0.682	0.142
	Zinc	Partial Carcass	140.	0.207	38.5	0.0569
7-06-3						
	Aluminum	Partial Carcass	767	0.500	219	0.142
	Arsenic	Partial Carcass	2.68	0.0472	0.764	0.0135
	Boron	Partial Carcass	0.651	0.500	0.186	0.142
	Barium	Partial Carcass	59.8	0.100	17.0	0.0285

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Beryllium	Partial Carcass	< 0.0500	0.0500	< 0.0142	0.0142
	Cadmium	Partial Carcass	< 0.0189	0.0189	< 0.00539	0.00539
	Chromium	Partial Carcass	2.58	0.500	0.735	0.142
	Copper	Partial Carcass	3.28	0.500	0.935	0.142
	Iron	Partial Carcass	838	1.00	239	0.285
	Mercury	Partial Carcass	0.205	0.00830	0.0584	0.00237
	Magnesium	Partial Carcass	1780	2.00	507	0.570
	Manganese	Partial Carcass	39.9	0.200	11.4	0.0570
	Molybdenum	Partial Carcass	< 1.00	1.00	< 0.285	0.285
	Nickel	Partial Carcass	2.24	0.500	0.638	0.142
	Lead	Partial Carcass	0.763	0.0472	0.217	0.0135
	Selenium	Partial Carcass	1.74	0.0472	0.496	0.0135
	Strontium	Partial Carcass	164	0.0500	46.7	0.0142
	Vanadium	Partial Carcass	2.81	0.500	0.801	0.142
	Zinc	Partial Carcass	147	0.200	41.9	0.0570
7-06-4						
	Aluminum	Partial Carcass	626	0.524	210.	0.176
	Arsenic	Partial Carcass	2.68	0.0495	0.898	0.0166
	Boron	Partial Carcass	0.576	0.524	0.193	0.176
	Barium	Partial Carcass	65.3	0.105	21.9	0.0352
	Beryllium	Partial Carcass	< 0.0524	0.0524	< 0.0176	0.0176
	Cadmium	Partial Carcass	< 0.0198	0.0198	< 0.00663	0.00663
	Chromium	Partial Carcass	2.06	0.524	0.690	0.176
	Copper	Partial Carcass	2.81	0.524	0.941	0.176
	Iron	Partial Carcass	634	1.05	212	0.352
	Mercury	Partial Carcass	0.181	0.00720	0.0606	0.00241
	Magnesium	Partial Carcass	1690	2.10	566	0.704
	Manganese	Partial Carcass	36.8	0.210	12.3	0.0704
	Molybdenum	Partial Carcass	< 1.05	1.05	< 0.352	0.352

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Nickel	Partial Carcass	1.87	0.524	0.626	0.176
	Lead	Partial Carcass	0.668	0.0495	0.224	0.0166
	Selenium	Partial Carcass	1.81	0.0495	0.606	0.0166
	Strontium	Partial Carcass	154	0.0524	51.6	0.0176
	Vanadium	Partial Carcass	1.74	0.524	0.583	0.176
	Zinc	Partial Carcass	170.	0.210	57.0	0.0704
7-06-5						
	Aluminum	Partial Carcass	328	0.513	102	0.160
	Arsenic	Partial Carcass	2.80	0.0484	0.874	0.0151
	Boron	Partial Carcass	< 0.513	0.513	< 0.160	0.160
	Barium	Partial Carcass	54.4	0.103	17.0	0.0321
	Beryllium	Partial Carcass	< 0.0513	0.0513	< 0.0160	0.0160
	Cadmium	Partial Carcass	< 0.0194	0.0194	< 0.00605	0.00605
	Chromium	Partial Carcass	0.993	0.513	0.310	0.160
	Copper	Partial Carcass	2.31	0.513	0.721	0.160
	Iron	Partial Carcass	344	1.03	107	0.321
	Mercury	Partial Carcass	0.185	0.00630	0.0577	0.00197
	Magnesium	Partial Carcass	1640	2.05	512	0.640
	Manganese	Partial Carcass	49.9	0.205	15.6	0.0640
	Molybdenum	Partial Carcass	< 1.03	1.03	< 0.321	0.321
	Nickel	Partial Carcass	1.10	0.513	0.343	0.160
	Lead	Partial Carcass	0.392	0.0484	0.122	0.0151
	Selenium	Partial Carcass	1.42	0.0484	0.443	0.0151
	Strontium	Partial Carcass	187	0.0513	58.3	0.0160
	Vanadium	Partial Carcass	1.19	0.513	0.371	0.160
	Zinc	Partial Carcass	146	0.205	45.6	0.0640
7-06-6						
	Aluminum	Partial Carcass	289	0.521	87.3	0.157
	Arsenic	Partial Carcass	2.55	0.0492	0.770	0.0149

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Boron	Partial Carcass	< 0.521	0.521	< 0.157	0.157
	Barium	Partial Carcass	57.6	0.104	17.4	0.0314
	Beryllium	Partial Carcass	< 0.0521	0.0521	< 0.0157	0.0157
	Cadmium	Partial Carcass	< 0.0197	0.0197	< 0.00595	0.00595
	Chromium	Partial Carcass	0.728	0.521	0.220	0.157
	Copper	Partial Carcass	2.32	0.521	0.701	0.157
	Iron	Partial Carcass	309	1.04	93.3	0.314
	Mercury	Partial Carcass	0.232	0.00680	0.0701	0.00205
	Magnesium	Partial Carcass	1530	2.09	462	0.631
	Manganese	Partial Carcass	31.7	0.209	9.57	0.0631
	Molybdenum	Partial Carcass	< 1.04	1.04	< 0.314	0.314
	Nickel	Partial Carcass	1.19	0.521	0.359	0.157
	Lead	Partial Carcass	0.338	0.0492	0.102	0.0149
	Selenium	Partial Carcass	1.40	0.0492	0.423	0.0149
	Strontium	Partial Carcass	170.	0.0521	51.3	0.0157
	Vanadium	Partial Carcass	1.12	0.521	0.338	0.157
	Zinc	Partial Carcass	165	0.209	49.8	0.0631
7-08-1						
	Aluminum	Partial Carcass	542	0.468	154	0.133
	Arsenic	Partial Carcass	4.10	0.0468	1.16	0.0133
	Boron	Partial Carcass	0.562	0.468	0.160	0.133
	Barium	Partial Carcass	71.4	0.0940	20.3	0.0267
	Beryllium	Partial Carcass	< 0.0468	0.0468	< 0.0133	0.0133
	Cadmium	Partial Carcass	0.0776	0.00935	0.0220	0.00266
	Chromium	Partial Carcass	< 0.468	0.468	< 0.133	0.133
	Copper	Partial Carcass	3.72	0.468	1.06	0.133
	Iron	Partial Carcass	749	0.940	213	0.267
	Mercury	Partial Carcass	0.192	0.00440	0.0545	0.00125
	Magnesium	Partial Carcass	1570	1.87	446	0.531

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Manganese	Partial Carcass	56.6	0.187	16.1	0.0531
	Molybdenum	Partial Carcass	< 0.940	0.940	< 0.267	0.267
	Nickel	Partial Carcass	0.898	0.468	0.255	0.133
	Lead	Partial Carcass	0.959	0.0468	0.272	0.0133
	Selenium	Partial Carcass	1.74	0.0468	0.494	0.0133
	Strontium	Partial Carcass	120.	0.0468	34.1	0.0133
	Vanadium	Partial Carcass	1.47	0.468	0.417	0.133
	Zinc	Partial Carcass	149	0.187	42.3	0.0531
	Methyl Mercury	Partial Carcass	0.181	0.00571	0.0514	0.00162
7-08-2						
	Aluminum	Partial Carcass	419	0.463	119	0.132
	Arsenic	Partial Carcass	3.75	0.0463	1.07	0.0132
	Boron	Partial Carcass	< 0.463	0.463	< 0.132	0.132
	Barium	Partial Carcass	67.6	0.0930	19.3	0.0265
	Beryllium	Partial Carcass	< 0.0463	0.0463	< 0.0132	0.0132
	Cadmium	Partial Carcass	0.0230	0.00925	0.00656	0.00264
	Chromium	Partial Carcass	< 0.463	0.463	< 0.132	0.132
	Copper	Partial Carcass	3.19	0.463	0.909	0.132
	Iron	Partial Carcass	574	0.930	164	0.265
	Mercury	Partial Carcass	0.195	0.00440	0.0556	0.00125
	Magnesium	Partial Carcass	1440	1.85	410.	0.527
	Manganese	Partial Carcass	62.8	0.185	17.9	0.0527
	Molybdenum	Partial Carcass	< 0.930	0.930	< 0.265	0.265
	Nickel	Partial Carcass	0.866	0.463	0.247	0.132
	Lead	Partial Carcass	0.722	0.0463	0.206	0.0132
	Selenium	Partial Carcass	1.89	0.0463	0.539	0.0132
	Strontium	Partial Carcass	105	0.0463	29.9	0.0132
	Vanadium	Partial Carcass	1.22	0.463	0.348	0.132
	Zinc	Partial Carcass	143	0.185	40.8	0.0527

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Methyl Mercury	Partial Carcass	0.197	0.00908	0.0561	0.00259
7-08-3						
	Aluminum	Partial Carcass	81.0	0.484	22.0	0.132
	Arsenic	Partial Carcass	2.83	0.0484	0.770	0.0132
	Boron	Partial Carcass	< 0.484	0.484	< 0.132	0.132
	Barium	Partial Carcass	47.5	0.0970	12.9	0.0264
	Beryllium	Partial Carcass	< 0.0484	0.0484	< 0.0132	0.0132
	Cadmium	Partial Carcass	< 0.00968	0.00968	< 0.00263	0.00263
	Chromium	Partial Carcass	< 0.484	0.484	< 0.132	0.132
	Copper	Partial Carcass	2.56	0.484	0.696	0.132
	Iron	Partial Carcass	130.	0.970	35.4	0.264
	Mercury	Partial Carcass	0.136	0.00400	0.0370	0.00109
	Magnesium	Partial Carcass	1300	1.94	354	0.528
	Manganese	Partial Carcass	14.8	0.194	4.03	0.0528
	Molybdenum	Partial Carcass	< 0.970	0.970	< 0.264	0.264
	Nickel	Partial Carcass	< 0.484	0.484	< 0.132	0.132
	Lead	Partial Carcass	0.343	0.0484	0.0933	0.0132
	Selenium	Partial Carcass	1.69	0.0484	0.460	0.0132
	Strontium	Partial Carcass	104	0.0484	28.3	0.0132
	Vanadium	Partial Carcass	< 0.484	0.484	< 0.132	0.132
	Zinc	Partial Carcass	135	0.194	36.7	0.0528
	Methyl Mercury	Partial Carcass	0.143	0.00675	0.0389	0.00184
7-08-4						
	Aluminum	Partial Carcass	394	0.449	104	0.119
	Arsenic	Partial Carcass	3.43	0.0449	0.909	0.0119
	Boron	Partial Carcass	< 0.449	0.449	< 0.119	0.119
	Barium	Partial Carcass	64.2	0.0900	17.0	0.0238
	Beryllium	Partial Carcass	< 0.0449	0.0449	< 0.0119	0.0119
	Cadmium	Partial Carcass	< 0.00899	0.00899	< 0.00238	0.00238

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Chromium	Partial Carcass	< 0.449	0.449	< 0.119	0.119
	Copper	Partial Carcass	3.07	0.449	0.814	0.119
	Iron	Partial Carcass	590.	0.900	156	0.238
	Mercury	Partial Carcass	0.219	0.00440	0.0580	0.00117
	Magnesium	Partial Carcass	1520	1.80	403	0.477
	Manganese	Partial Carcass	35.6	0.180	9.43	0.0477
	Molybdenum	Partial Carcass	< 0.900	0.900	< 0.238	0.238
	Nickel	Partial Carcass	0.867	0.449	0.230	0.119
	Lead	Partial Carcass	0.908	0.0449	0.241	0.0119
	Selenium	Partial Carcass	1.84	0.0449	0.488	0.0119
	Strontium	Partial Carcass	121	0.0449	32.1	0.0119
	Vanadium	Partial Carcass	1.35	0.449	0.358	0.119
	Zinc	Partial Carcass	145	0.180	38.4	0.0477
	Methyl Mercury	Partial Carcass	0.199	0.00982	0.0527	0.00260
7-08-5						
	Aluminum	Partial Carcass	205	0.470	58.4	0.134
	Arsenic	Partial Carcass	3.20	0.0470	0.912	0.0134
	Boron	Partial Carcass	< 0.470	0.470	< 0.134	0.134
	Barium	Partial Carcass	42.0	0.0940	12.0	0.0268
	Beryllium	Partial Carcass	< 0.0470	0.0470	< 0.0134	0.0134
	Cadmium	Partial Carcass	< 0.00940	0.00940	< 0.00268	0.00268
	Chromium	Partial Carcass	< 0.470	0.470	< 0.134	0.134
	Copper	Partial Carcass	3.22	0.470	0.918	0.134
	Iron	Partial Carcass	314	0.940	89.5	0.268
	Mercury	Partial Carcass	0.204	0.00450	0.0581	0.00128
	Magnesium	Partial Carcass	1470	1.88	419	0.536
	Manganese	Partial Carcass	38.8	0.188	11.1	0.0536
	Molybdenum	Partial Carcass	< 0.940	0.940	< 0.268	0.268
	Nickel	Partial Carcass	0.622	0.470	0.177	0.134

Sample Number	Analyte	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	Lead	Partial Carcass	0.435	0.0470	0.124	0.0134
	Selenium	Partial Carcass	1.92	0.0470	0.547	0.0134
	Strontium	Partial Carcass	128	0.0470	36.5	0.0134
	Vanadium	Partial Carcass	0.708	0.470	0.202	0.134
	Zinc	Partial Carcass	153	0.188	43.6	0.0536
	Methyl Mercury	Partial Carcass	0.221	0.00742	0.0630	0.00211
7-08-6						
	Aluminum	Partial Carcass	30.8	0.456	8.84	0.131
	Arsenic	Partial Carcass	2.80	0.0456	0.804	0.0131
	Boron	Partial Carcass	< 0.456	0.456	< 0.131	0.131
	Barium	Partial Carcass	26.3	0.0910	7.55	0.0261
	Beryllium	Partial Carcass	< 0.0456	0.0456	< 0.0131	0.0131
	Cadmium	Partial Carcass	< 0.00913	0.00913	< 0.00262	0.00262
	Chromium	Partial Carcass	< 0.456	0.456	< 0.131	0.131
	Copper	Partial Carcass	2.78	0.456	0.798	0.131
	Iron	Partial Carcass	68.7	0.910	19.7	0.261
	Mercury	Partial Carcass	0.357	0.00410	0.102	0.00118
	Magnesium	Partial Carcass	1260	1.83	362	0.525
	Manganese	Partial Carcass	14.0	0.183	4.02	0.0525
	Molybdenum	Partial Carcass	< 0.910	0.910	< 0.261	0.261
	Nickel	Partial Carcass	< 0.456	0.456	< 0.131	0.131
	Lead	Partial Carcass	0.136	0.0456	0.0390	0.0131
	Selenium	Partial Carcass	1.92	0.0456	0.551	0.0131
	Strontium	Partial Carcass	98.8	0.0456	28.4	0.0131
	Vanadium	Partial Carcass	< 0.456	0.456	< 0.131	0.131
	Zinc	Partial Carcass	132	0.183	37.9	0.0525
	Methyl Mercury	Partial Carcass	0.360	0.00746	0.103	0.00214

6. Duplicates

Sample Number	Analyte	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average (ppm/%)	Relative Percent Diff.
06385810							
	Aluminum	Water	Wet	0.905	0.892	0.899	1.45
	Arsenic	Water	Wet	0.00440	0.00433	0.00436	1.60
	Boron	Water	Wet	0.116	0.115	0.116	0.870
	Barium	Water	Wet	0.106	0.105	0.106	0.950
	Beryllium	Water	Wet	< 0.000500	< 0.000500	0.000250	0.000
	Cadmium	Water	Wet	0.000260	0.000260	0.000260	0.000
	Chromium	Water	Wet	< 0.00500	< 0.00500	0.00250	0.000
	Copper	Water	Wet	0.0130	0.0130	0.0130	0.000
	Iron	Water	Wet	1.04	1.03	1.04	0.970
	Mercury	Water	Wet	0.00000300	0.00000300	0.00000300	0.000
	Magnesium	Water	Wet	4.48	4.42	4.45	1.35
	Manganese	Water	Wet	0.0750	0.0740	0.0745	1.34
	Molybdenum	Water	Wet	< 0.0100	< 0.0100	0.00500	0.000
	Nickel	Water	Wet	0.00800	0.00800	0.00800	0.000
	Lead	Water	Wet	0.0120	0.0119	0.0120	0.840
	Selenium	Water	Wet	0.000200	0.000300	0.000250	40.0
	Strontium	Water	Wet	0.264	0.263	0.264	0.380
	Vanadium	Water	Wet	0.00700	0.00600	0.00650	15.4
	Zinc	Water	Wet	0.0700	0.0680	0.0690	2.90
1-07-2							
	Methyl Mercury	Partial Carcass	Dry	0.221	0.249	0.235	11.9
4-07-1							
	Aluminum	Partial Carcass	Dry	382	359	370.	6.21
	Arsenic	Partial Carcass	Dry	3.02	3.28	3.15	8.25
	Boron	Partial Carcass	Dry	< 0.511	< 0.500	0.253	2.18
	Barium	Partial Carcass	Dry	79.0	73.2	76.1	7.62

Sample Number	Analyte	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average (ppm/%)	Relative Percent Diff.
	Beryllium	Partial Carcass	Dry	< 0.0511	< 0.0500	0.0253	2.18
	Cadmium	Partial Carcass	Dry	< 0.0193	< 0.0189	0.00955	2.09
	Chromium	Partial Carcass	Dry	0.618	0.533	0.576	14.8
	Copper	Partial Carcass	Dry	3.49	3.25	3.37	7.12
	Iron	Partial Carcass	Dry	457	401	429	13.0
	Magnesium	Partial Carcass	Dry	1730	1620	1680	6.57
	Manganese	Partial Carcass	Dry	32.4	28.6	30.5	12.5
	Molybdenum	Partial Carcass	Dry	< 1.02	< 1.00	0.505	1.98
	Nickel	Partial Carcass	Dry	1.24	1.12	1.18	10.2
	Lead	Partial Carcass	Dry	0.376	0.361	0.368	4.07
	Selenium	Partial Carcass	Dry	2.00	2.03	2.01	1.49
	Strontium	Partial Carcass	Dry	138	125	132	9.89
	Vanadium	Partial Carcass	Dry	1.32	1.32	1.32	0.000
	Zinc	Partial Carcass	Dry	140.	134	137	4.38
4-07-4							
	Mercury	Partial Carcass	Dry	0.161	0.155	0.158	3.80
4-07-5							
	Aluminum	Partial Carcass	Dry	201	210.	206	4.38
	Arsenic	Partial Carcass	Dry	2.79	3.05	2.92	8.90
	Boron	Partial Carcass	Dry	0.758	< 0.425	0.485	112
	Barium	Partial Carcass	Dry	29.7	30.5	30.1	2.66
	Beryllium	Partial Carcass	Dry	< 0.0493	< 0.0425	0.0229	14.8
	Cadmium	Partial Carcass	Dry	< 0.00987	< 0.00850	0.00459	14.9
	Chromium	Partial Carcass	Dry	< 0.493	< 0.425	0.229	14.8
	Copper	Partial Carcass	Dry	3.00	2.93	2.96	2.36
	Iron	Partial Carcass	Dry	294	305	300.	3.67
	Magnesium	Partial Carcass	Dry	1360	1400	1380	2.90
	Manganese	Partial Carcass	Dry	35.1	34.7	34.9	1.15
	Molybdenum	Partial Carcass	Dry	< 0.990	< 0.850	0.460	15.2

Sample Number	Analyte	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average (ppm/%)	Relative Percent Diff.
	Nickel	Partial Carcass	Dry	< 0.493	0.473	0.360	63.0
	Lead	Partial Carcass	Dry	0.562	0.475	0.518	16.8
	Selenium	Partial Carcass	Dry	2.33	2.38	2.36	2.12
	Strontium	Partial Carcass	Dry	155	156	156	0.640
	Vanadium	Partial Carcass	Dry	0.977	0.837	0.907	15.4
	Zinc	Partial Carcass	Dry	125	123	124	1.61
4-08-4							
	Mercury	Partial Carcass	Dry	0.111	0.109	0.110	1.82
7-06-2							
	Mercury	Partial Carcass	Dry	0.273	0.275	0.274	0.730
7-08-4							
	Mercury	Partial Carcass	Dry	0.219	0.214	0.216	2.31

7. Spike Recoveries

Sample Number	Analyte	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
06385810							
	Aluminum	Water	Wet	1.00	0.905	1.10	90.5
	Arsenic	Water	Wet	0.0200	0.0178	4.55	89.0
	Boron	Water	Wet	0.500	0.495	4.31	99.0
	Barium	Water	Wet	0.500	0.499	4.72	99.8
	Beryllium	Water	Wet	0.0100	0.00985	40.0	98.5
	Cadmium	Water	Wet	0.0200	0.0199	76.9	99.7
	Chromium	Water	Wet	0.100	0.0965	40.0	96.5
	Copper	Water	Wet	0.200	0.194	15.4	97.0
	Iron	Water	Wet	2.00	1.94	1.92	97.0
	Mercury	Water	Wet	0.0000500	0.0000450	16.7	90.0
	Magnesium	Water	Wet	10.0	9.32	2.23	93.2
	Manganese	Water	Wet	0.500	0.489	6.67	97.8
	Molybdenum	Water	Wet	0.500	0.475	100.	95.0
	Nickel	Water	Wet	0.100	0.0950	12.5	95.0
	Lead	Water	Wet	0.0200	0.0193	1.67	96.5
	Selenium	Water	Wet	0.0200	0.0182	100.	91.0
	Strontium	Water	Wet	0.200	0.174	0.760	87.0
	Vanadium	Water	Wet	0.100	0.0960	14.3	96.0
	Zinc	Water	Wet	1.00	0.990	14.3	99.0
4-07-4							
	Aluminum	Partial Carcass	Dry	191	184	0.430	96.1
	Arsenic	Partial Carcass	Dry	19.1	19.1	6.19	99.8
	Boron	Partial Carcass	Dry	47.8	53.6	189	112
	Barium	Partial Carcass	Dry	9.57	9.80	0.160	102
	Beryllium	Partial Carcass	Dry	0.957	1.05	37.8	110.
	Cadmium	Partial Carcass	Dry	4.78	5.43	501	114
	Chromium	Partial Carcass	Dry	19.1	21.4	25.4	112
	Copper	Partial Carcass	Dry	19.1	21.8	5.36	114

Sample Number	Analyte	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	Iron	Partial Carcass	Dry	191	202	0.370	106
	Magnesium	Partial Carcass	Dry	957	1030	0.560	108
	Manganese	Partial Carcass	Dry	47.8	54.3	1.68	113
	Molybdenum	Partial Carcass	Dry	9.57	10.7	19.0	112
	Nickel	Partial Carcass	Dry	9.57	10.5	7.59	110.
	Lead	Partial Carcass	Dry	9.57	9.44	17.2	98.7
	Selenium	Partial Carcass	Dry	4.78	4.96	2.31	104
	Strontium	Partial Carcass	Dry	19.1	18.0	0.150	94.0
	Vanadium	Partial Carcass	Dry	19.1	21.2	12.3	111
	Zinc	Partial Carcass	Dry	95.7	109	0.760	114
4-08-6							
	Mercury	Partial Carcass	Dry	0.703	0.739	5.86	105
7-06-3							
	Mercury	Partial Carcass	Dry	0.602	0.621	2.94	103
7-08-1							
	Mercury	Partial Carcass	Dry	0.942	0.978	4.91	104
7-08-2							
	Aluminum	Partial Carcass	Dry	246	291	0.590	118
	Arsenic	Partial Carcass	Dry	24.6	22.0	6.55	89.8
	Boron	Partial Carcass	Dry	61.4	60.7	265	98.8
	Barium	Partial Carcass	Dry	12.3	11.1	0.180	90.4
	Beryllium	Partial Carcass	Dry	1.23	1.33	53.0	108
	Cadmium	Partial Carcass	Dry	6.14	6.58	267	107
	Chromium	Partial Carcass	Dry	24.6	25.8	106	105
	Copper	Partial Carcass	Dry	24.6	26.6	7.70	108
	Iron	Partial Carcass	Dry	246	312	0.430	127
	Magnesium	Partial Carcass	Dry	1230	1360	0.850	111
	Manganese	Partial Carcass	Dry	61.4	65.2	0.980	106
	Molybdenum	Partial Carcass	Dry	12.3	12.3	26.4	100.
	Nickel	Partial Carcass	Dry	12.3	12.7	14.2	104
	Lead	Partial Carcass	Dry	12.3	13.7	17.0	111
	Selenium	Partial Carcass	Dry	6.14	6.78	3.25	110.

Sample Number	Analyte	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	Strontium	Partial Carcass	Dry	24.6	27.0	0.230	110.
	Vanadium	Partial Carcass	Dry	24.6	25.4	20.1	103
	Zinc	Partial Carcass	Dry	123	127	0.860	103
7-08-6							
	Mercury	Partial Carcass	Dry	0.553	0.483	1.55	87.3
	Methyl Mercury	Partial Carcass	Dry	1.55	1.33	4.32	85.5

*** For a spike to be a valid measure of method accuracy, this ratio must be higher than 1.0.

10. QAQC Summary

1. Procedural Blank Summary

Procedural Blank Summary of Blank Equivalent Concentration (BEC) Data

Within a lab sample matrix, there must be three or more Blank results for a given analyte in order to generate a report.

Analyte	Lab Sample Matrix	No. of Records	Basis	Lowest BEC (ppm)	Highest BEC (ppm)	BEC Mean (ppm)	BEC STDEV
Mercury	Animal Tissue	4	Dry	< 0.00430	< 0.00704	0.00280	0.000640

BEC = Blank Equivalent Concentration   STD = Standard Deviation

10.2. Duplicate Summary

Duplicate Summary of Relative Percent Difference (RPD) Data

Within a lab sample matrix and concentration range, there must be three or more Duplicate results for a given analyte in order to generate a report.

Mean concentration greater than 10 times the Limit of Detection						
Analyte	**Lab Sample Matrix**	**No. of Samples**	**Lowest RPD**	**Highest RPD**	**RPD Mean (ppm)**	**RPD STD**
Mercury	Animal Tissue	4	0.73	3.8	2.16	1.27

RPD = Relative Percent Difference STD = Standard Deviation

10.3. Spike Summary

Spike Summary of Percent Recovery (PR) Data

Within a lab sample matrix, there must be three or more Spike results for a given analyte in order to generate a report.

Analyte	Lab Sample Matrix	No. of Samples	Lowest PR	Highest PR	PR Mean	PR STD
Mercury	Animal Tissue	4	87.34	105.09	99.83	8.36

PR = Percent Recovery STD = Standard Deviation

10.4. SRM Summary

Standard Reference Material Summary of Percent Recovery (PR) Data

Within an SRM ID, there must be three or more Recoveries for a given analyte in order to generate a report.

SRM ID: NRCC DOLT-3 SRM Name: Dogfish Liver					
Analyte	**No. of Records**	**Lowest PR**	**Highest PR**	**PR Mean**	**PR STDEV**
Mercury	4	102.37	107.42	105.71	2.26

SRM ID: NRCC DORM-2 SRM Name: Dogfish Muscle					
Analyte	**No. of Records**	**Lowest PR**	**Highest PR**	**PR Mean**	**PR STDEV**
Mercury	4	93.32	104.31	97.58	4.87

PR = Percent Recovery STD = Standard Deviation

11. QA/QC Anomalies

1. Blank Frequency Anomalies

The required number of blank analyses were performed.

11.2. Duplicate Frequency Anomalies

The required number of duplicate analyses were performed.

11.3. Spike Frequency Anomalies

The required number of spike analyses were performed.

11.4. Reference Material Frequency Anomalies

The required number of Standard Reference Material analyses were performed.

11.5. Mass Spec Frequency Anomalies

No Carbamate, OC, or OP data exists in this set of results; therefore, the anomaly test was not performed.

11.6. Limit of Detection Anomalies

Limits of Detection were within the contract requirements.

11.7. Blank Anomalies

Procedural Blank analyses were acceptable.

11.8. Duplicate Anomalies

All duplicate results were within normal limits.

11.9. Spike Anomalies

All spike results were within normal limits.

11.10. S.R.M. Anomalies

All SRM results were within normal limits with the following exceptions.

Analyte	S.R.M. ID	Certified Value	95% Confidence Interval	LOD (ppm/%)	Result (ppm/%)	% Recovery	See QA/QC Note No.
Manganese	NIST 2976	33.0	2.00	0.207	38.6	117	1
Strontium	NIST 2976	93.0	2.00	0.0459	65.2	70.1	2

S.R.M Names

SRM ID	SRM Name
NIST 2976	Mussel Tissue

11.11. QA/QC Notes

QA/QC Note Number and Comments
Sample IDs 7-08-2, 7-08-3, 7-08-5 and 7-08-6 had ratios of greater than 1 for MeHg:Hg in the data report. To express MeHg as Hg you have to divide the MeHg based values by approx. 1.075. In doing this, the ratio values for all samples that were analyzed for MeHg and Hg are acceptable. 1-2 The values are acceptable for Mn and Sr in SRM Nist 2976 and should have no effect on the interpretation of the data. QA/QC Approved: Brenda Montgomery, Inorganic Analytical Chemist

12. Analytical Methods

Below are the analytical methods used by TERL to produce the results included in this report.

Method Codes:	001 005

Lab Matrix	Analyte
Animal Tissue	Aluminum
	Boron
	Barium
	Beryllium
	Chromium
	Copper
	Iron
	Magnesium
	Manganese
	Molybdenum
	Nickel
	Strontium
	Vanadium
	Zinc

Method Code: 001

LABORATORY: Trace Element Research Laboratory

Digestion of biological tissue.

Liquid or solid biological tissue samples are wet digested with nitric acid and converted into acidic digest solutions for analysis by various atomic spectroscopy methods. When possible, tissue is freeze dried in order to minimize loss of analytes and to facilitate subsequent sample preparation steps, and then homogenized to a fine powder by ball-milling in plastic containers. Approximately 0.20 to 0.25 g of powdered tissue is weighed into a Teflon reaction vessel and 3 ml of HNO3 are added. The closed reaction vessel is heated in a 130 C oven until digestion is complete. Samples are then diluted to a final volume of 20 ml with quartz distilled water and stored in 1 oz. polyethylene bottles for later analysis by instrumental techniques.

Method Code: 005

LABORATORY: Trace Element Research Laboratory

Analysis of trace metals by inductively coupled plasma
optical emission spectroscopy (ICP).

Liquid samples are nebulized and the resulting aerosol is transported to the plasma torch. Element-specific atomic-line emission spectra are produced by a inductively coupled argon plasma. The spectra are dispersed by a grating spectrometer, and the intensities of the lines are monitored by photomultiplier tubes or solid state detectors. Samples are quantitated by comparison with external standards. One or more internal standards may be incorporated to compensate for physical effects resulting from viscosity and varying levels of total dissolved solids in the samples. Background correction is required and is measured adjacent to analyte lines on samples during analysis.

Method Codes:	001 016

Lab Matrix	**Analyte**
Animal Tissue	Arsenic
	Cadmium
	Lead
	Selenium

Method Code: 001

LABORATORY: Trace Element Research Laboratory

Digestion of biological tissue.

Liquid or solid biological tissue samples are wet digested with nitric acid and converted into acidic digest solutions for analysis by various atomic spectroscopy methods. When possible, tissue is freeze dried in order to minimize loss of analytes and to facilitate subsequent sample preparation steps, and then homogenized to a fine powder by ball-milling in plastic containers. Approximately 0.20 to 0.25 g of powdered tissue is weighed into a Teflon reaction vessel and 3 ml of HNO3 are added. The closed reaction vessel is heated in a 130 C oven until digestion is complete. Samples are then diluted to a final volume of 20 ml with quartz distilled water and stored in 1 oz. polyethylene bottles for later analysis by instrumental techniques.

Method Code: 016

LABORATORY: Trace Element Research Laboratory

Analysis of trace metals in water samples by inductively
coupled plasma-mass spectroscopy (ICP-MS).

Concentrations of trace elements in water samples are determined with an atomic spectroscopy method that relies on ionization of sample constituents in a high temperature argon plasma and separation of positively-charged ions on the basis of their

mass:charge ratios (m:z) by a quadrupole mass spectrometer. The method offers extremely low detection limits but is subject to interferences from atomic and molecular ions having values within 1 AMU of the target ions. Sample preconcentration and matrix elimination can sometimes eliminate these problems, along with those resulting from high total dissolved solids.

Method Codes:	003 007

Lab Matrix	Analyte
Water	Mercury

Method Code: 003

LABORATORY: Trace Element Research Laboratory

Digestion of water, soil, sediment, and biological tissue
for mercury analysis.

Before samples are analyzed by the CVAAS method in use in this laboratory, the mercury is converted to the Hg2+ form. Mercury is digested by a modified version of EPA method 245.5 and 245.6. Sediment and tissue samples can be analyzed either freeze dried or on a wet basis. Sediment samples are homogenized by mixing before subsampling, while tissue samples are homogenized in the original sample containers either after freeze drying or with a Tekmar Tissumizer and subsampled. Samples are digested with nitric acid, sulfuric acid, potassium permanganate, and potassium persulfate in polypropylene tubes in a water bath at 90-95 C. Before analysis, hydroxylamine hydrochloride is added to reduce excess permanganate and the samples are brought to volume with distilled-deionized water.

Method Code: 007

LABORATORY: Trace Element Research Laboratory

Analysis of mercury by cold-vapor atomic absorption
spectroscopy (CVAAS).

In this procedure, divalent mercury (Hg++) in aqueous samples (digests of water, tissue or sediment samples) is reduced to the elemental state (Hgo) by a strong reducing agent (stannous chloride). Gaseous Hgo enters the sweep gas and is introduced into an atomic absorption cell, where light produced by a mercury vapor lamp is absorbed by the free Hg atoms. Mercury in the sample is determined by comparing light absorption of the sample with that of external calibration standards.

Method Codes:	004 005

Lab Matrix	Analyte
Water	Aluminum
	Boron
	Barium
	Beryllium
	Chromium
	Copper
	Iron
	Magnesium
	Manganese
	Molybdenum
	Nickel
	Strontium
	Vanadium
	Zinc

Method Code: 004

LABORATORY: Trace Element Research Laboratory

Digestion of water samples for "total recoverable" metals
(other than mercury).

Water samples are digested for two hours at 85 degrees Centigrade in polyethylene containers with ultrapure nitric and hydrochloric acids. Acid strength, on a vol:vol basis, is 1% HCl and 0.5% HNO3. Sample aliquots for digestion are taken after vigorous shaking to assure resuspension of solids that may have settled. The original sample must have had preservative added (usually HNO3) in order to ensure that metals do not adhere to the walls of the container.

Method Code: 005

LABORATORY: Trace Element Research Laboratory

Analysis of trace metals by inductively coupled plasma
optical emission spectroscopy (ICP).

Liquid samples are nebulized and the resulting aerosol is transported to the plasma torch. Element-specific atomic-line emission spectra are produced by a inductively coupled argon plasma. The spectra are dispersed by a grating spectrometer, and the intensities of the lines are monitored by photomultiplier tubes or solid state detectors. Samples are quantitated by comparison with external standards. One or more internal standards may be incorporated to compensate for physical effects resulting from viscosity

and varying levels of total dissolved solids in the samples. Background correction is required and is measured adjacent to analyte lines on samples during analysis.

Method Codes:	004 016

Lab Matrix	Analyte
Water	Arsenic
	Cadmium
	Lead
	Selenium

Method Code: 004

LABORATORY: Trace Element Research Laboratory

Digestion of water samples for "total recoverable" metals
(other than mercury).

Water samples are digested for two hours at 85 degrees Centigrade in polyethylene containers with ultrapure nitric and hydrochloric acids. Acid strength, on a vol:vol basis, is 1% HCl and 0.5% HNO3. Sample aliquots for digestion are taken after vigorous shaking to assure resuspension of solids that may have settled. The original sample must have had preservative added (usually HNO3) in order to ensure that metals do not adhere to the walls of the container.

Method Code: 016

LABORATORY: Trace Element Research Laboratory

Analysis of trace metals in water samples by inductively
coupled plasma-mass spectroscopy (ICP-MS).

Concentrations of trace elements in water samples are determined with an atomic spectroscopy method that relies on ionization of sample constituents in a high temperature argon plasma and separation of positively-charged ions on the basis of their mass:charge ratios (m:z) by a quadrupole mass spectrometer. The method offers extremely low detection limits but is subject to interferences from atomic and molecular ions having values within 1 AMU of the target ions. Sample preconcentration and matrix elimination can sometimes eliminate these problems, along with those resulting from high total dissolved solids.

Method Codes:	014

Lab Matrix	Analyte
Animal Tissue	% Moisture

Method Code: 014
LABORATORY: Trace Element Research Laboratory Moisture content of sediment, soil, and tissue samples. Moisture content is determined by weight loss upon freeze-drying, and is expressed as weight percent of the original wet sample. Depending upon sample size, either the whole sample or a representative aliquot is frozen and then dried under vacuum until a constant weight is attained. Samples are prepared and dried using plastic materials, whenever possible, in order to minimize potential contamination artifacts that might impact subsequent trace element analysis.

Method Codes:	024

Lab Matrix	Analyte
Animal Tissue	Mercury

Method Code: 024
LABORATORY: Trace Element Research Laboratory Determination of mercury in tissue and sediment Determination of mercury in tissue and sediment samples by decomposition, trapping, and atomic absorption. Total mercury is determined in wet or dry samples by combustion in a stream of oxygen, trapping on a gold column, release by electrothermal heating, and analysis by atomic absorption. Mercury is reported in ppm on either a wet or dry weight basis.

Method Codes:	026

Lab Matrix	Analyte
Animal Tissue	Methyl Mercury

Method Code: 026
LABORATORY:Trace Element Research Laboratory Determination of methyl mercury in water Determination of methyl mercury in water by distillation, ethylation, trapping, gas chromatography, and atomic fluorescence. Methyl mercury in water is distilled to separate it from interfering species and then ethylated with sodium tetraethyl borate. Methyl ethyl mercury is trapped on a Tenax column and then separated on an isothermal GC column. Following pyrolysis of the separated species, Hg is detected by atomic fluorescence.

Appendix 3B. Organic Analytical Results for Rio Grande Silvery Minnow Health Study Catalog 2020128 by TDI Brooks International, Inc., College Station, Texas.

Analytical Results Report TOC

1. ECDMS Analytical Results Report 4/12/2010

Catalog Number	Purchase Order Number	Lab ID	Catalog Submitter	ECDMS User ID
2020128	94420-09-Y005	TDI	Lusk, Joel - Albuquerque, NM	r2alfo

Catalog Title	Rio Grande Silvery Minnow Health Study
Lab Name:	TDI - Brooks International, Inc.

Notes, Symbols and Abbreviations Used
Based on the report options selected the report should be **printed in landscape mode**
Notes, Symbols and Abbreviations Used The following may appear before a reported result (e.g. < 1234). < - Less than symbol indicates that the actual result is less than the reported detection limit. > - Greater than symbol indicates that the actual result is greater than the reported result.
All results are reported as 3 significant digits.
All results are reported as parts per million (ppm), or percent, unless otherwise noted.

1. Integrity Report

Lab Receipt Date	09/03/2009	Lab Approval Date	09/03/2009

Catalog Problems
No problems reported
Problem Resolution

2. Bulk Data

Sample Number	Sample Matrix	Sample Weight (grams)	Percent Lipid	Percent Moisture
1-08-1	Partial Carcass	31.9	4.07	76.4
1-08-2	Partial Carcass	52.6	4.30	73.8
1-08-3	Partial Carcass	50.3	4.95	72.6
1-08-4	Partial Carcass	47.1	6.56	74.1
1-08-6	Partial Carcass	55.9	7.46	73.3
10-06-1	Partial Carcass	38.5	7.70	73.3
10-06-2	Partial Carcass	77.8	5.37	74.6
10-06-3	Partial Carcass	79	6.56	70.0
10-06-4	Partial Carcass	122	7.75	71.3
10-06-5	Partial Carcass	110.2	7.85	69.9
10-07-1	Partial Carcass	25.7	5.25	71.7
10-07-2	Partial Carcass	26.7	6.90	71.4
10-07-3	Partial Carcass	43.3	8.20	70.6
10-07-4	Partial Carcass	39.8	5.68	76.4
10-07-5	Partial Carcass	46.9	5.77	73.7
10-07-6	Partial Carcass	29	6.00	74.5
4-08-1	Partial Carcass	33.5	7.51	76.2
4-08-2	Partial Carcass	47.2	6.11	74.3
4-08-3	Partial Carcass	189.9	7.73	72.4
4-08-4	Partial Carcass	134.7	5.49	73.3
4-08-6	Partial Carcass	32.4	5.57	74.8
7-06-1	Partial Carcass	79.6	5.47	73.9
7-06-2	Partial Carcass	83.6	6.80	74.1
7-06-3	Partial Carcass	32.9	5.93	74.2
7-06-4	Partial Carcass	59	10.8	69.0
7-06-5	Partial Carcass	64.5	7.73	72.1
7-06-6	Partial Carcass	40.3	8.00	73.5
06385810	Water			

4. Contaminant Concentrations

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
1,2,3,4,6,7,8-HpCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,4,6,7,8-HpCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,4,7,8,9-HpCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
1,2,3,4,7,8-HxCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,4,7,8-HxCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,4-Tetrachlorobenzene						
	*10-06-1	Partial Carcass	< 0.000565	0.000565	< 0.000151	0.000151
	*10-06-2	Partial Carcass	< 0.000592	0.000592	< 0.000151	0.000151
	*10-06-3	Partial Carcass	< 0.000502	0.000502	< 0.000151	0.000151
	*10-06-4	Partial Carcass	< 0.000525	0.000525	< 0.000151	0.000151
	*10-06-5	Partial Carcass	< 0.000501	0.000501	< 0.000151	0.000151
	*10-07-1	Partial Carcass	< 0.000532	0.000532	< 0.000151	0.000151
	*10-07-2	Partial Carcass	< 0.000528	0.000528	< 0.000151	0.000151
	*10-07-3	Partial Carcass	< 0.000513	0.000513	< 0.000151	0.000151
	*10-07-4	Partial Carcass	< 0.000639	0.000639	< 0.000151	0.000151
	*10-07-5	Partial Carcass	< 0.000573	0.000573	< 0.000151	0.000151

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-6	Partial Carcass	< 0.000590	0.000590	< 0.000151	0.000151
	*4-08-6	Partial Carcass	< 0.000597	0.000597	< 0.000151	0.000151
1,2,3,6,7,8-HxCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,6,7,8-HxCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,7,8,9-HxCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
1,2,3,7,8,9-HxCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,7,8-PeCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,3,7,8-PeCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
1,2,4,5-Tetrachlorobenzene						
	*10-06-1	Partial Carcass	< 0.00101	0.00101	< 0.000269	0.000269

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-06-2	Partial Carcass	< 0.00106	0.00106	< 0.000269	0.000269
	*10-06-3	Partial Carcass	< 0.000898	0.000898	< 0.000269	0.000269
	*10-06-4	Partial Carcass	< 0.000938	0.000938	< 0.000269	0.000269
	*10-06-5	Partial Carcass	< 0.000895	0.000895	< 0.000269	0.000269
	*10-07-1	Partial Carcass	< 0.000951	0.000951	< 0.000269	0.000269
	*10-07-2	Partial Carcass	< 0.000943	0.000943	< 0.000269	0.000269
	*10-07-3	Partial Carcass	< 0.000917	0.000917	< 0.000269	0.000269
	*10-07-4	Partial Carcass	< 0.00114	0.00114	< 0.000269	0.000269
	*10-07-5	Partial Carcass	< 0.00102	0.00102	< 0.000269	0.000269
	*10-07-6	Partial Carcass	< 0.00106	0.00106	< 0.000269	0.000269
	*4-08-6	Partial Carcass	< 0.00107	0.00107	< 0.000269	0.000269
1,6,7-Trimethyl-naphthalene						
	1-08-1	Partial Carcass	< 0.00191	0.00191	< 0.000451	0.000451
	1-08-2	Partial Carcass	< 0.00170	0.00170	< 0.000446	0.000446
	1-08-3	Partial Carcass	< 0.00163	0.00163	< 0.000447	0.000447
	1-08-4	Partial Carcass	< 0.00173	0.00173	< 0.000450	0.000450
	1-08-6	Partial Carcass	< 0.00164	0.00164	< 0.000438	0.000438
1-methylnaphthalene						
	1-08-1	Partial Carcass	0.0110	0.00319	0.00260	0.000755
	1-08-2	Partial Carcass	0.0141	0.00285	0.00370	0.000747
	1-08-3	Partial Carcass	0.0124	0.00274	0.00340	0.000749
	1-08-4	Partial Carcass	0.0139	0.00291	0.00360	0.000754
	1-08-6	Partial Carcass	0.00787	0.00275	0.00210	0.000735
1-methylphenanthrene						
	1-08-1	Partial Carcass	< 0.00216	0.00216	< 0.000511	0.000511
	1-08-2	Partial Carcass	0.0298	0.00193	0.00780	0.000506
	1-08-3	Partial Carcass	0.00658	0.00185	0.00180	0.000507
	1-08-4	Partial Carcass	< 0.00197	0.00197	< 0.000510	0.000510
	1-08-6	Partial Carcass	0.0176	0.00187	0.00470	0.000498

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
2,3,4,6,7,8-HxCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
2,3,4,7,8-PeCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
2,3,7,8-TCDD						
	10-06-1	Partial Carcass	< 0.00000686	0.00000686	< 0.00000183	0.00000183
	10-06-2	Partial Carcass	< 0.00000743	0.00000743	< 0.00000189	0.00000189
	10-06-3	Partial Carcass	< 0.00000653	0.00000653	< 0.00000196	0.00000196
	10-06-4	Partial Carcass	< 0.00000690	0.00000690	< 0.00000198	0.00000198
	10-07-1	Partial Carcass	< 0.00000702	0.00000702	< 0.00000199	0.00000199
	10-07-2	Partial Carcass	< 0.00000690	0.00000690	< 0.00000197	0.00000197
	10-07-3	Partial Carcass	< 0.00000664	0.00000664	< 0.00000195	0.00000195
	10-07-4	Partial Carcass	< 0.00000797	0.00000797	< 0.00000188	0.00000188
2,3,7,8-TCDF						
	10-06-1	Partial Carcass	< 0.00000686	0.00000686	< 0.00000183	0.00000183

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-2	Partial Carcass	< 0.00000743	0.00000743	< 0.00000189	0.00000189
	10-06-3	Partial Carcass	< 0.00000653	0.00000653	< 0.00000196	0.00000196
	10-06-4	Partial Carcass	< 0.00000690	0.00000690	< 0.00000198	0.00000198
	10-07-1	Partial Carcass	< 0.00000702	0.00000702	< 0.00000199	0.00000199
	10-07-2	Partial Carcass	< 0.00000690	0.00000690	< 0.00000197	0.00000197
	10-07-3	Partial Carcass	< 0.00000664	0.00000664	< 0.00000195	0.00000195
	10-07-4	Partial Carcass	< 0.00000797	0.00000797	< 0.00000188	0.00000188
2,6-dimethylnaphthalene						
	1-08-1	Partial Carcass	0.00296	0.00278	0.000700	0.000658
	1-08-2	Partial Carcass	0.00382	0.00249	0.00100	0.000651
	1-08-3	Partial Carcass	0.00475	0.00238	0.00130	0.000652
	1-08-4	Partial Carcass	0.00386	0.00253	0.00100	0.000656
	1-08-6	Partial Carcass	< 0.00240	0.00240	< 0.000640	0.000640
2-methylnaphthalene						
	1-08-1	Partial Carcass	0.0216	0.00345	0.00510	0.000816
	1-08-2	Partial Carcass	0.0260	0.00308	0.00680	0.000808
	1-08-3	Partial Carcass	0.0216	0.00296	0.00590	0.000809
	1-08-4	Partial Carcass	0.0220	0.00314	0.00570	0.000814
	1-08-6	Partial Carcass	0.0142	0.00298	0.00380	0.000794
Aldrin						
	*10-06-1	Partial Carcass	< 0.000458	0.000458	< 0.000122	0.000122
	*10-06-2	Partial Carcass	< 0.000481	0.000481	< 0.000122	0.000122
	*10-06-3	Partial Carcass	< 0.000408	0.000408	< 0.000122	0.000122
	*10-06-4	Partial Carcass	< 0.000426	0.000426	< 0.000122	0.000122
	*10-06-5	Partial Carcass	< 0.000406	0.000406	< 0.000122	0.000122
	*10-07-1	Partial Carcass	< 0.000432	0.000432	< 0.000122	0.000122
	*10-07-2	Partial Carcass	< 0.000428	0.000428	< 0.000122	0.000122
	*10-07-3	Partial Carcass	< 0.000416	0.000416	< 0.000122	0.000122
	*10-07-4	Partial Carcass	< 0.000518	0.000518	< 0.000122	0.000122

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-5	Partial Carcass	< 0.000465	0.000465	< 0.000122	0.000122
	*10-07-6	Partial Carcass	< 0.000479	0.000479	< 0.000122	0.000122
	*4-08-6	Partial Carcass	< 0.000484	0.000484	< 0.000122	0.000122
BDE# 1						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 10						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 100						
	4-08-1	Partial Carcass	0.00420	0.00156	0.00100	0.000370

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-2	Partial Carcass	0.0121	0.00141	0.00310	0.000363
	4-08-3	Partial Carcass	0.0323	0.00132	0.00890	0.000364
	4-08-4	Partial Carcass	0.0604	0.00141	0.0161	0.000376
	4-08-6	Partial Carcass	0.0139	0.00142	0.00350	0.000360
	7-06-1	Partial Carcass	0.0233	0.00139	0.00610	0.000364
	7-06-2	Partial Carcass	0.0263	0.00142	0.00680	0.000367
	7-06-3	Partial Carcass	0.104	0.00144	0.0269	0.000370
	7-06-4	Partial Carcass	0.144	0.00114	0.0447	0.000355
	7-06-5	Partial Carcass	0.0555	0.00128	0.0155	0.000358
	7-06-6	Partial Carcass	0.0343	0.00130	0.00910	0.000345
BDE# 11						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 116						
	4-08-1	Partial Carcass	< 0.000797	0.000797	< 0.000190	0.000190
	4-08-2	Partial Carcass	< 0.000724	0.000724	< 0.000186	0.000186
	4-08-3	Partial Carcass	< 0.000676	0.000676	< 0.000186	0.000186
	4-08-4	Partial Carcass	< 0.000722	0.000722	< 0.000192	0.000192
	4-08-6	Partial Carcass	< 0.000730	0.000730	< 0.000184	0.000184
	7-06-1	Partial Carcass	< 0.000713	0.000713	< 0.000186	0.000186

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-2	Partial Carcass	< 0.000726	0.000726	< 0.000188	0.000188
	7-06-3	Partial Carcass	< 0.000736	0.000736	< 0.000190	0.000190
	7-06-4	Partial Carcass	< 0.000587	0.000587	< 0.000182	0.000182
	7-06-5	Partial Carcass	< 0.000657	0.000657	< 0.000183	0.000183
	7-06-6	Partial Carcass	< 0.000667	0.000667	< 0.000177	0.000177
BDE# 118						
	4-08-1	Partial Carcass	< 0.000797	0.000797	< 0.000190	0.000190
	4-08-2	Partial Carcass	< 0.000724	0.000724	< 0.000186	0.000186
	4-08-3	Partial Carcass	< 0.000676	0.000676	< 0.000186	0.000186
	4-08-4	Partial Carcass	< 0.000722	0.000722	< 0.000192	0.000192
	4-08-6	Partial Carcass	< 0.000730	0.000730	< 0.000184	0.000184
	7-06-1	Partial Carcass	< 0.000713	0.000713	< 0.000186	0.000186
	7-06-2	Partial Carcass	< 0.000726	0.000726	< 0.000188	0.000188
	7-06-3	Partial Carcass	< 0.000736	0.000736	< 0.000190	0.000190
	7-06-4	Partial Carcass	< 0.000587	0.000587	< 0.000182	0.000182
	7-06-5	Partial Carcass	< 0.000657	0.000657	< 0.000183	0.000183
	7-06-6	Partial Carcass	< 0.000667	0.000667	< 0.000177	0.000177
BDE# 119						
	4-08-1	Partial Carcass	< 0.000797	0.000797	< 0.000190	0.000190
	4-08-2	Partial Carcass	< 0.000724	0.000724	< 0.000186	0.000186
	4-08-3	Partial Carcass	0.00109	0.000676	0.000300	0.000186
	4-08-4	Partial Carcass	0.00300	0.000722	0.000800	0.000192
	4-08-6	Partial Carcass	< 0.000730	0.000730	< 0.000184	0.000184
	7-06-1	Partial Carcass	< 0.000713	0.000713	< 0.000186	0.000186
	7-06-2	Partial Carcass	0.000772	0.000726	0.000200	0.000188
	7-06-3	Partial Carcass	0.00543	0.000736	0.00140	0.000190
	7-06-4	Partial Carcass	0.00645	0.000587	0.00200	0.000182
	7-06-5	Partial Carcass	0.00287	0.000657	0.000800	0.000183
	7-06-6	Partial Carcass	0.00188	0.000667	0.000500	0.000177

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
BDE# 12						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 126						
	4-08-1	Partial Carcass	< 0.000797	0.000797	< 0.000190	0.000190
	4-08-2	Partial Carcass	< 0.000724	0.000724	< 0.000186	0.000186
	4-08-3	Partial Carcass	0.000725	0.000676	0.000200	0.000186
	4-08-4	Partial Carcass	0.00112	0.000722	0.000300	0.000192
	4-08-6	Partial Carcass	< 0.000730	0.000730	< 0.000184	0.000184
	7-06-1	Partial Carcass	< 0.000713	0.000713	< 0.000186	0.000186
	7-06-2	Partial Carcass	< 0.000726	0.000726	< 0.000188	0.000188
	7-06-3	Partial Carcass	0.00155	0.000736	0.000400	0.000190
	7-06-4	Partial Carcass	0.00258	0.000587	0.000800	0.000182
	7-06-5	Partial Carcass	< 0.000657	0.000657	< 0.000183	0.000183
	7-06-6	Partial Carcass	< 0.000667	0.000667	< 0.000177	0.000177
BDE# 13						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 138						
	4-08-1	Partial Carcass	0.000840	0.000593	0.000200	0.000141
	4-08-2	Partial Carcass	< 0.000538	0.000538	< 0.000138	0.000138
	4-08-3	Partial Carcass	< 0.000503	0.000503	< 0.000139	0.000139
	4-08-4	Partial Carcass	< 0.000537	0.000537	< 0.000143	0.000143
	4-08-6	Partial Carcass	< 0.000543	0.000543	< 0.000137	0.000137
	7-06-1	Partial Carcass	< 0.000531	0.000531	< 0.000139	0.000139
	7-06-2	Partial Carcass	< 0.000540	0.000540	< 0.000140	0.000140
	7-06-3	Partial Carcass	< 0.000548	0.000548	< 0.000141	0.000141
	7-06-4	Partial Carcass	< 0.000436	0.000436	< 0.000135	0.000135
	7-06-5	Partial Carcass	< 0.000488	0.000488	< 0.000136	0.000136
	7-06-6	Partial Carcass	< 0.000496	0.000496	< 0.000132	0.000132
BDE# 15						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	0.000750	0.000681	0.000200	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	0.00272	0.000695	0.000700	0.000179
	7-06-4	Partial Carcass	0.00258	0.000553	0.000800	0.000172

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-5	Partial Carcass	0.00108	0.000620	0.000300	0.000173
	7-06-6	Partial Carcass	0.000754	0.000629	0.000200	0.000167
BDE# 153						
	4-08-1	Partial Carcass	0.000840	0.000643	0.000200	0.000153
	4-08-2	Partial Carcass	0.00156	0.000584	0.000400	0.000150
	4-08-3	Partial Carcass	0.00471	0.000546	0.00130	0.000151
	4-08-4	Partial Carcass	0.00825	0.000583	0.00220	0.000155
	4-08-6	Partial Carcass	0.00198	0.000589	0.000500	0.000149
	7-06-1	Partial Carcass	0.00191	0.000576	0.000500	0.000151
	7-06-2	Partial Carcass	0.00232	0.000586	0.000600	0.000152
	7-06-3	Partial Carcass	0.0128	0.000594	0.00330	0.000153
	7-06-4	Partial Carcass	0.0135	0.000474	0.00420	0.000147
	7-06-5	Partial Carcass	0.00358	0.000530	0.00100	0.000148
	7-06-6	Partial Carcass	0.00264	0.000538	0.000700	0.000143
BDE# 154						
	4-08-1	Partial Carcass	0.00168	0.000736	0.000400	0.000175
	4-08-2	Partial Carcass	0.00389	0.000668	0.00100	0.000172
	4-08-3	Partial Carcass	0.00979	0.000624	0.00270	0.000172
	4-08-4	Partial Carcass	0.0184	0.000666	0.00490	0.000178
	4-08-6	Partial Carcass	0.00475	0.000674	0.00120	0.000170
	7-06-1	Partial Carcass	0.00612	0.000659	0.00160	0.000172
	7-06-2	Partial Carcass	0.00734	0.000670	0.00190	0.000173
	7-06-3	Partial Carcass	0.0272	0.000680	0.00700	0.000175
	7-06-4	Partial Carcass	0.0413	0.000542	0.0128	0.000168
	7-06-5	Partial Carcass	0.0150	0.000606	0.00420	0.000169
	7-06-6	Partial Carcass	0.00980	0.000616	0.00260	0.000163
BDE# 155						
	4-08-1	Partial Carcass	< 0.000593	0.000593	< 0.000141	0.000141
	4-08-2	Partial Carcass	< 0.000538	0.000538	< 0.000138	0.000138

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-3	Partial Carcass	< 0.000503	0.000503	< 0.000139	0.000139
	4-08-4	Partial Carcass	< 0.000537	0.000537	< 0.000143	0.000143
	4-08-6	Partial Carcass	< 0.000543	0.000543	< 0.000137	0.000137
	7-06-1	Partial Carcass	< 0.000531	0.000531	< 0.000139	0.000139
	7-06-2	Partial Carcass	< 0.000540	0.000540	< 0.000140	0.000140
	7-06-3	Partial Carcass	< 0.000548	0.000548	< 0.000141	0.000141
	7-06-4	Partial Carcass	< 0.000436	0.000436	< 0.000135	0.000135
	7-06-5	Partial Carcass	< 0.000488	0.000488	< 0.000136	0.000136
	7-06-6	Partial Carcass	< 0.000496	0.000496	< 0.000132	0.000132
BDE# 166						
	4-08-1	Partial Carcass	< 0.000593	0.000593	< 0.000141	0.000141
	4-08-2	Partial Carcass	< 0.000538	0.000538	< 0.000138	0.000138
	4-08-3	Partial Carcass	< 0.000503	0.000503	< 0.000139	0.000139
	4-08-4	Partial Carcass	< 0.000537	0.000537	< 0.000143	0.000143
	4-08-6	Partial Carcass	< 0.000543	0.000543	< 0.000137	0.000137
	7-06-1	Partial Carcass	< 0.000531	0.000531	< 0.000139	0.000139
	7-06-2	Partial Carcass	< 0.000540	0.000540	< 0.000140	0.000140
	7-06-3	Partial Carcass	< 0.000548	0.000548	< 0.000141	0.000141
	7-06-4	Partial Carcass	< 0.000436	0.000436	< 0.000135	0.000135
	7-06-5	Partial Carcass	< 0.000488	0.000488	< 0.000136	0.000136
	7-06-6	Partial Carcass	< 0.000496	0.000496	< 0.000132	0.000132
BDE# 17						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-3	Partial Carcass	0.00233	0.000695	0.000600	0.000179
	7-06-4	Partial Carcass	0.00323	0.000553	0.00100	0.000172
	7-06-5	Partial Carcass	0.00108	0.000620	0.000300	0.000173
	7-06-6	Partial Carcass	0.000754	0.000629	0.000200	0.000167
BDE# 181						
	4-08-1	Partial Carcass	< 0.000550	0.000550	< 0.000131	0.000131
	4-08-2	Partial Carcass	< 0.000500	0.000500	< 0.000128	0.000128
	4-08-3	Partial Carcass	< 0.000466	0.000466	< 0.000129	0.000129
	4-08-4	Partial Carcass	< 0.000498	0.000498	< 0.000133	0.000133
	4-08-6	Partial Carcass	< 0.000504	0.000504	< 0.000127	0.000127
	7-06-1	Partial Carcass	< 0.000492	0.000492	< 0.000129	0.000129
	7-06-2	Partial Carcass	< 0.000501	0.000501	< 0.000130	0.000130
	7-06-3	Partial Carcass	< 0.000508	0.000508	< 0.000131	0.000131
	7-06-4	Partial Carcass	< 0.000405	0.000405	< 0.000126	0.000126
	7-06-5	Partial Carcass	< 0.000453	0.000453	< 0.000127	0.000127
	7-06-6	Partial Carcass	< 0.000460	0.000460	< 0.000122	0.000122
BDE# 183						
	4-08-1	Partial Carcass	0.000840	0.000550	0.000200	0.000131
	4-08-2	Partial Carcass	0.000778	0.000500	0.000200	0.000128
	4-08-3	Partial Carcass	< 0.000466	0.000466	< 0.000129	0.000129
	4-08-4	Partial Carcass	0.00112	0.000498	0.000300	0.000133
	4-08-6	Partial Carcass	< 0.000504	0.000504	< 0.000127	0.000127
	7-06-1	Partial Carcass	< 0.000492	0.000492	< 0.000129	0.000129
	7-06-2	Partial Carcass	< 0.000501	0.000501	< 0.000130	0.000130
	7-06-3	Partial Carcass	< 0.000508	0.000508	< 0.000131	0.000131
	7-06-4	Partial Carcass	< 0.000405	0.000405	< 0.000126	0.000126
	7-06-5	Partial Carcass	< 0.000453	0.000453	< 0.000127	0.000127
	7-06-6	Partial Carcass	< 0.000460	0.000460	< 0.000122	0.000122
BDE# 190						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-1	Partial Carcass	0.00210	0.000417	0.000500	0.0000990
	4-08-2	Partial Carcass	0.00156	0.000378	0.000400	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	0.000750	0.000377	0.000200	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 194						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 195						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 196						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 197						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 198/199/203/200						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 2						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 201						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 202						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 204						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	< 0.000378	0.000378	< 0.0000970	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	< 0.000377	0.000377	< 0.000101	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	< 0.000373	0.000373	< 0.0000970	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-4	Partial Carcass	< 0.000307	0.000307	< 0.0000950	0.0000950
	7-06-5	Partial Carcass	< 0.000343	0.000343	< 0.0000960	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 205						
	4-08-1	Partial Carcass	0.00210	0.000417	0.000500	0.0000990
	4-08-2	Partial Carcass	0.00194	0.000378	0.000500	0.0000970
	4-08-3	Partial Carcass	0.00145	0.000353	0.000400	0.0000970
	4-08-4	Partial Carcass	0.00225	0.000377	0.000600	0.000101
	4-08-6	Partial Carcass	0.00277	0.000382	0.000700	0.0000960
	7-06-1	Partial Carcass	0.00153	0.000373	0.000400	0.0000970
	7-06-2	Partial Carcass	0.00193	0.000379	0.000500	0.0000980
	7-06-3	Partial Carcass	0.00233	0.000385	0.000600	0.0000990
	7-06-4	Partial Carcass	0.00129	0.000307	0.000400	0.0000950
	7-06-5	Partial Carcass	0.00179	0.000343	0.000500	0.0000960
	7-06-6	Partial Carcass	0.00188	0.000349	0.000500	0.0000920
BDE# 206						
	4-08-1	Partial Carcass	0.00294	0.000417	0.000700	0.0000990
	4-08-2	Partial Carcass	0.00233	0.000378	0.000600	0.0000970
	4-08-3	Partial Carcass	< 0.000353	0.000353	< 0.0000970	0.0000970
	4-08-4	Partial Carcass	0.00225	0.000377	0.000600	0.000101
	4-08-6	Partial Carcass	< 0.000382	0.000382	< 0.0000960	0.0000960
	7-06-1	Partial Carcass	0.00230	0.000373	0.000600	0.0000970
	7-06-2	Partial Carcass	< 0.000379	0.000379	< 0.0000980	0.0000980
	7-06-3	Partial Carcass	< 0.000385	0.000385	< 0.0000990	0.0000990
	7-06-4	Partial Carcass	0.00129	0.000307	0.000400	0.0000950
	7-06-5	Partial Carcass	0.00179	0.000343	0.000500	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 207						
	4-08-1	Partial Carcass	0.00336	0.000417	0.000800	0.0000990

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-2	Partial Carcass	0.00311	0.000378	0.000800	0.0000970
	4-08-3	Partial Carcass	0.00254	0.000353	0.000700	0.0000970
	4-08-4	Partial Carcass	0.00262	0.000377	0.000700	0.000101
	4-08-6	Partial Carcass	0.00356	0.000382	0.000900	0.0000960
	7-06-1	Partial Carcass	0.00268	0.000373	0.000700	0.0000970
	7-06-2	Partial Carcass	0.00270	0.000379	0.000700	0.0000980
	7-06-3	Partial Carcass	0.00233	0.000385	0.000600	0.0000990
	7-06-4	Partial Carcass	0.00161	0.000307	0.000500	0.0000950
	7-06-5	Partial Carcass	0.00287	0.000343	0.000800	0.0000960
	7-06-6	Partial Carcass	0.00226	0.000349	0.000600	0.0000920
BDE# 208						
	4-08-1	Partial Carcass	< 0.000417	0.000417	< 0.0000990	0.0000990
	4-08-2	Partial Carcass	0.00156	0.000378	0.000400	0.0000970
	4-08-3	Partial Carcass	0.00181	0.000353	0.000500	0.0000970
	4-08-4	Partial Carcass	0.00150	0.000377	0.000400	0.000101
	4-08-6	Partial Carcass	0.00238	0.000382	0.000600	0.0000960
	7-06-1	Partial Carcass	0.00153	0.000373	0.000400	0.0000970
	7-06-2	Partial Carcass	0.00116	0.000379	0.000300	0.0000980
	7-06-3	Partial Carcass	0.00194	0.000385	0.000500	0.0000990
	7-06-4	Partial Carcass	0.000968	0.000307	0.000300	0.0000950
	7-06-5	Partial Carcass	0.00143	0.000343	0.000400	0.0000960
	7-06-6	Partial Carcass	< 0.000349	0.000349	< 0.0000920	0.0000920
BDE# 209						
	4-08-1	Partial Carcass	< 0.00619	0.00619	< 0.00147	0.00147
	4-08-2	Partial Carcass	< 0.00562	0.00562	< 0.00144	0.00144
	4-08-3	Partial Carcass	< 0.00525	0.00525	< 0.00145	0.00145
	4-08-4	Partial Carcass	< 0.00560	0.00560	< 0.00149	0.00149
	4-08-6	Partial Carcass	< 0.00567	0.00567	< 0.00143	0.00143
	7-06-1	Partial Carcass	< 0.00554	0.00554	< 0.00145	0.00145

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-2	Partial Carcass	< 0.00563	0.00563	< 0.00146	0.00146
	7-06-3	Partial Carcass	< 0.00572	0.00572	< 0.00147	0.00147
	7-06-4	Partial Carcass	< 0.00455	0.00455	< 0.00141	0.00141
	7-06-5	Partial Carcass	< 0.00510	0.00510	< 0.00142	0.00142
	7-06-6	Partial Carcass	< 0.00518	0.00518	< 0.00137	0.00137
BDE# 25						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	0.000645	0.000553	0.000200	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 28						
	4-08-1	Partial Carcass	0.00126	0.000805	0.000300	0.000192
	4-08-2	Partial Carcass	0.00350	0.000731	0.000900	0.000188
	4-08-3	Partial Carcass	0.00616	0.000683	0.00170	0.000188
	4-08-4	Partial Carcass	0.00937	0.000729	0.00250	0.000194
	4-08-6	Partial Carcass	0.00238	0.000738	0.000600	0.000186
	7-06-1	Partial Carcass	0.0103	0.000721	0.00270	0.000188
	7-06-2	Partial Carcass	0.00850	0.000733	0.00220	0.000190
	7-06-3	Partial Carcass	0.0198	0.000744	0.00510	0.000192
	7-06-4	Partial Carcass	0.0274	0.000593	0.00850	0.000184
	7-06-5	Partial Carcass	0.0129	0.000663	0.00360	0.000185
	7-06-6	Partial Carcass	0.00792	0.000674	0.00210	0.000179

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
BDE# 3						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	0.00218	0.000638	0.000600	0.000176
	4-08-4	Partial Carcass	0.00262	0.000681	0.000700	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	0.0128	0.000695	0.00330	0.000179
	7-06-4	Partial Carcass	0.00516	0.000553	0.00160	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 30						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 32						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 33						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 35						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 37						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	0.000776	0.000695	0.000200	0.000179
	7-06-4	Partial Carcass	0.00161	0.000553	0.000500	0.000172
	7-06-5	Partial Carcass	0.00108	0.000620	0.000300	0.000173
	7-06-6	Partial Carcass	0.00188	0.000629	0.000500	0.000167
BDE# 47						
	4-08-1	Partial Carcass	0.0252	0.000876	0.00600	0.000209
	4-08-2	Partial Carcass	0.0856	0.000796	0.0220	0.000205
	4-08-3	Partial Carcass	0.272	0.000743	0.0751	0.000205
	4-08-4	Partial Carcass	0.484	0.000793	0.129	0.000212
	4-08-6	Partial Carcass	0.105	0.000803	0.0266	0.000203
	7-06-1	Partial Carcass	0.186	0.000784	0.0487	0.000205
	7-06-2	Partial Carcass	0.195	0.000798	0.0506	0.000207
	7-06-3	Partial Carcass	1.03	0.000809	0.266	0.000209
	7-06-4	Partial Carcass	1.24	0.000645	0.385	0.000200
	7-06-5	Partial Carcass	0.455	0.000722	0.127	0.000201
	7-06-6	Partial Carcass	0.265	0.000733	0.0704	0.000194
BDE# 49/71						
	4-08-1	Partial Carcass	< 0.000644	0.000644	< 0.000153	0.000153
	4-08-2	Partial Carcass	0.00350	0.000585	0.000900	0.000150

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-3	Partial Carcass	0.00725	0.000546	0.00200	0.000151
	4-08-4	Partial Carcass	0.0120	0.000583	0.00320	0.000155
	4-08-6	Partial Carcass	0.00396	0.000590	0.00100	0.000149
	7-06-1	Partial Carcass	0.00765	0.000576	0.00200	0.000151
	7-06-2	Partial Carcass	0.00811	0.000586	0.00210	0.000152
	7-06-3	Partial Carcass	0.0221	0.000595	0.00570	0.000153
	7-06-4	Partial Carcass	0.0403	0.000474	0.0125	0.000147
	7-06-5	Partial Carcass	0.0233	0.000530	0.00650	0.000148
	7-06-6	Partial Carcass	0.0151	0.000539	0.00400	0.000143
BDE# 66						
	4-08-1	Partial Carcass	< 0.000481	0.000481	< 0.000114	0.000114
	4-08-2	Partial Carcass	< 0.000436	0.000436	< 0.000112	0.000112
	4-08-3	Partial Carcass	< 0.000408	0.000408	< 0.000112	0.000112
	4-08-4	Partial Carcass	< 0.000435	0.000435	< 0.000116	0.000116
	4-08-6	Partial Carcass	< 0.000440	0.000440	< 0.000111	0.000111
	7-06-1	Partial Carcass	< 0.000430	0.000430	< 0.000112	0.000112
	7-06-2	Partial Carcass	< 0.000438	0.000438	< 0.000113	0.000113
	7-06-3	Partial Carcass	< 0.000444	0.000444	< 0.000114	0.000114
	7-06-4	Partial Carcass	< 0.000354	0.000354	< 0.000110	0.000110
	7-06-5	Partial Carcass	< 0.000396	0.000396	< 0.000111	0.000111
	7-06-6	Partial Carcass	< 0.000402	0.000402	< 0.000107	0.000107
BDE# 7						
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 75						
	4-08-1	Partial Carcass	< 0.000876	0.000876	< 0.000209	0.000209
	4-08-2	Partial Carcass	< 0.000796	0.000796	< 0.000205	0.000205
	4-08-3	Partial Carcass	< 0.000743	0.000743	< 0.000205	0.000205
	4-08-4	Partial Carcass	< 0.000793	0.000793	< 0.000212	0.000212
	4-08-6	Partial Carcass	< 0.000803	0.000803	< 0.000203	0.000203
	7-06-1	Partial Carcass	< 0.000784	0.000784	< 0.000205	0.000205
	7-06-2	Partial Carcass	< 0.000798	0.000798	< 0.000207	0.000207
	7-06-3	Partial Carcass	< 0.000809	0.000809	< 0.000209	0.000209
	7-06-4	Partial Carcass	< 0.000645	0.000645	< 0.000200	0.000200
	7-06-5	Partial Carcass	0.00143	0.000722	0.000400	0.000201
	7-06-6	Partial Carcass	< 0.000733	0.000733	< 0.000194	0.000194
BDE# 77						
	4-08-1	Partial Carcass	< 0.000876	0.000876	< 0.000209	0.000209
	4-08-2	Partial Carcass	0.00117	0.000796	0.000300	0.000205
	4-08-3	Partial Carcass	< 0.000743	0.000743	< 0.000205	0.000205
	4-08-4	Partial Carcass	0.00188	0.000793	0.000500	0.000212
	4-08-6	Partial Carcass	< 0.000803	0.000803	< 0.000203	0.000203
	7-06-1	Partial Carcass	0.00153	0.000784	0.000400	0.000205
	7-06-2	Partial Carcass	0.00270	0.000798	0.000700	0.000207
	7-06-3	Partial Carcass	< 0.000809	0.000809	< 0.000209	0.000209
	7-06-4	Partial Carcass	< 0.000645	0.000645	< 0.000200	0.000200
	7-06-5	Partial Carcass	< 0.000722	0.000722	< 0.000201	0.000201
	7-06-6	Partial Carcass	0.00377	0.000733	0.00100	0.000194
BDE# 8						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-1	Partial Carcass	< 0.000752	0.000752	< 0.000179	0.000179
	4-08-2	Partial Carcass	< 0.000683	0.000683	< 0.000176	0.000176
	4-08-3	Partial Carcass	< 0.000638	0.000638	< 0.000176	0.000176
	4-08-4	Partial Carcass	< 0.000681	0.000681	< 0.000182	0.000182
	4-08-6	Partial Carcass	< 0.000689	0.000689	< 0.000174	0.000174
	7-06-1	Partial Carcass	< 0.000673	0.000673	< 0.000176	0.000176
	7-06-2	Partial Carcass	< 0.000685	0.000685	< 0.000177	0.000177
	7-06-3	Partial Carcass	< 0.000695	0.000695	< 0.000179	0.000179
	7-06-4	Partial Carcass	< 0.000553	0.000553	< 0.000172	0.000172
	7-06-5	Partial Carcass	< 0.000620	0.000620	< 0.000173	0.000173
	7-06-6	Partial Carcass	< 0.000629	0.000629	< 0.000167	0.000167
BDE# 85						
	4-08-1	Partial Carcass	< 0.00164	0.00164	< 0.000391	0.000391
	4-08-2	Partial Carcass	< 0.00149	0.00149	< 0.000383	0.000383
	4-08-3	Partial Carcass	< 0.00139	0.00139	< 0.000384	0.000384
	4-08-4	Partial Carcass	0.00150	0.00148	0.000400	0.000396
	4-08-6	Partial Carcass	< 0.00150	0.00150	< 0.000380	0.000380
	7-06-1	Partial Carcass	< 0.00147	0.00147	< 0.000384	0.000384
	7-06-2	Partial Carcass	< 0.00149	0.00149	< 0.000387	0.000387
	7-06-3	Partial Carcass	0.00194	0.00152	0.000500	0.000391
	7-06-4	Partial Carcass	0.00355	0.00121	0.00110	0.000374
	7-06-5	Partial Carcass	< 0.00135	0.00135	< 0.000377	0.000377
	7-06-6	Partial Carcass	< 0.00137	0.00137	< 0.000364	0.000364
BDE# 99						
	4-08-1	Partial Carcass	< 0.000797	0.000797	< 0.000190	0.000190
	4-08-2	Partial Carcass	< 0.000724	0.000724	< 0.000186	0.000186
	4-08-3	Partial Carcass	< 0.000676	0.000676	< 0.000186	0.000186
	4-08-4	Partial Carcass	< 0.000722	0.000722	< 0.000192	0.000192
	4-08-6	Partial Carcass	< 0.000730	0.000730	< 0.000184	0.000184

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	7-06-1	Partial Carcass	< 0.000713	0.000713	< 0.000186	0.000186
	7-06-2	Partial Carcass	< 0.000726	0.000726	< 0.000188	0.000188
	7-06-3	Partial Carcass	< 0.000736	0.000736	< 0.000190	0.000190
	7-06-4	Partial Carcass	< 0.000587	0.000587	< 0.000182	0.000182
	7-06-5	Partial Carcass	< 0.000657	0.000657	< 0.000183	0.000183
	7-06-6	Partial Carcass	< 0.000667	0.000667	< 0.000177	0.000177
BDE-TOTAL						
	4-08-1	Partial Carcass	0.0462	0.00729	0.0110	0.00174
	4-08-2	Partial Carcass	0.123	0.00662	0.0315	0.00170
	4-08-3	Partial Carcass	0.343	0.00618	0.0946	0.00171
	4-08-4	Partial Carcass	0.614	0.00660	0.164	0.00176
	4-08-6	Partial Carcass	0.141	0.00668	0.0357	0.00169
	7-06-1	Partial Carcass	0.246	0.00653	0.0643	0.00171
	7-06-2	Partial Carcass	0.258	0.00664	0.0667	0.00172
	7-06-3	Partial Carcass	1.25	0.00674	0.323	0.00174
	7-06-4	Partial Carcass	1.54	0.00537	0.477	0.00166
	7-06-5	Partial Carcass	0.582	0.00601	0.162	0.00168
	7-06-6	Partial Carcass	0.348	0.00610	0.0924	0.00162
BHC (Total)						
	*10-06-1	Partial Carcass	< 0.00135	0.00135	< 0.000359	0.000359
	*10-06-2	Partial Carcass	< 0.00141	0.00141	< 0.000359	0.000359
	*10-06-3	Partial Carcass	< 0.00120	0.00120	< 0.000359	0.000359
	*10-06-4	Partial Carcass	< 0.00125	0.00125	< 0.000359	0.000359
	*10-06-5	Partial Carcass	< 0.00120	0.00120	< 0.000359	0.000359
	*10-07-1	Partial Carcass	< 0.00127	0.00127	< 0.000359	0.000359
	*10-07-2	Partial Carcass	< 0.00126	0.00126	< 0.000359	0.000359
	*10-07-3	Partial Carcass	0.00900	0.00122	0.00264	0.000359
	*10-07-4	Partial Carcass	< 0.00152	0.00152	< 0.000359	0.000359
	*10-07-5	Partial Carcass	< 0.00136	0.00136	< 0.000359	0.000359

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-6	Partial Carcass	< 0.00141	0.00141	< 0.000359	0.000359
	*4-08-6	Partial Carcass	< 0.00142	0.00142	< 0.000359	0.000359
Benzo(a)anthracene						
	1-08-1	Partial Carcass	< 0.00160	0.00160	< 0.000378	0.000378
	1-08-2	Partial Carcass	< 0.00143	0.00143	< 0.000374	0.000374
	1-08-3	Partial Carcass	< 0.00137	0.00137	< 0.000374	0.000374
	1-08-4	Partial Carcass	< 0.00145	0.00145	< 0.000377	0.000377
	1-08-6	Partial Carcass	< 0.00138	0.00138	< 0.000367	0.000367
C1-Fluoranthenes & Pyrenes						
	1-08-1	Partial Carcass	< 0.00541	0.00541	< 0.00128	0.00128
	1-08-2	Partial Carcass	< 0.00483	0.00483	< 0.00127	0.00127
	1-08-3	Partial Carcass	< 0.00464	0.00464	< 0.00127	0.00127
	1-08-4	Partial Carcass	< 0.00492	0.00492	< 0.00128	0.00128
	1-08-6	Partial Carcass	< 0.00466	0.00466	< 0.00124	0.00124
C1-Phenanthrenes & Anthracenes						
	1-08-1	Partial Carcass	< 0.00252	0.00252	< 0.000597	0.000597
	1-08-2	Partial Carcass	0.0302	0.00226	0.00790	0.000591
	1-08-3	Partial Carcass	0.00950	0.00216	0.00260	0.000592
	1-08-4	Partial Carcass	< 0.00230	0.00230	< 0.000596	0.000596
	1-08-6	Partial Carcass	0.0199	0.00218	0.00530	0.000581
C1-chrysenes						
	1-08-1	Partial Carcass	< 0.00397	0.00397	< 0.000938	0.000938
	1-08-2	Partial Carcass	< 0.00354	0.00354	< 0.000928	0.000928
	1-08-3	Partial Carcass	< 0.00340	0.00340	< 0.000930	0.000930
	1-08-4	Partial Carcass	< 0.00361	0.00361	< 0.000936	0.000936
	1-08-6	Partial Carcass	< 0.00342	0.00342	< 0.000912	0.000912
C1-dibenzothiophenes						
	1-08-1	Partial Carcass	< 0.00288	0.00288	< 0.000682	0.000682
	1-08-2	Partial Carcass	< 0.00258	0.00258	< 0.000675	0.000675

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-3	Partial Carcass	0.0113	0.00247	0.00310	0.000677
	1-08-4	Partial Carcass	0.0158	0.00262	0.00410	0.000681
	1-08-6	Partial Carcass	0.00600	0.00249	0.00160	0.000664
C1-fluorenes						
	1-08-1	Partial Carcass	0.00719	0.00350	0.00170	0.000828
	1-08-2	Partial Carcass	0.00993	0.00313	0.00260	0.000820
	1-08-3	Partial Carcass	< 0.00300	0.00300	< 0.000821	0.000821
	1-08-4	Partial Carcass	< 0.00319	0.00319	< 0.000826	0.000826
	1-08-6	Partial Carcass	< 0.00302	0.00302	< 0.000806	0.000806
C1-naphthalenes						
	1-08-1	Partial Carcass	0.0212	0.00664	0.00500	0.00157
	1-08-2	Partial Carcass	0.0260	0.00594	0.00680	0.00156
	1-08-3	Partial Carcass	0.0219	0.00570	0.00600	0.00156
	1-08-4	Partial Carcass	0.0231	0.00605	0.00600	0.00157
	1-08-6	Partial Carcass	0.0142	0.00573	0.00380	0.00153
C2-Phenanthrenes & Anthracenes						
	1-08-1	Partial Carcass	< 0.00252	0.00252	< 0.000597	0.000597
	1-08-2	Partial Carcass	< 0.00226	0.00226	< 0.000591	0.000591
	1-08-3	Partial Carcass	< 0.00216	0.00216	< 0.000592	0.000592
	1-08-4	Partial Carcass	< 0.00230	0.00230	< 0.000596	0.000596
	1-08-6	Partial Carcass	< 0.00218	0.00218	< 0.000581	0.000581
C2-chrysenes						
	1-08-1	Partial Carcass	< 0.00397	0.00397	< 0.000938	0.000938
	1-08-2	Partial Carcass	< 0.00354	0.00354	< 0.000928	0.000928
	1-08-3	Partial Carcass	< 0.00340	0.00340	< 0.000930	0.000930
	1-08-4	Partial Carcass	< 0.00361	0.00361	< 0.000936	0.000936
	1-08-6	Partial Carcass	< 0.00342	0.00342	< 0.000912	0.000912
C2-dibenzothiophenes						
	1-08-1	Partial Carcass	< 0.00288	0.00288	< 0.000682	0.000682

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-2	Partial Carcass	< 0.00258	0.00258	< 0.000675	0.000675
	1-08-3	Partial Carcass	< 0.00247	0.00247	< 0.000677	0.000677
	1-08-4	Partial Carcass	< 0.00262	0.00262	< 0.000681	0.000681
	1-08-6	Partial Carcass	< 0.00249	0.00249	< 0.000664	0.000664
C2-fluorenes						
	1-08-1	Partial Carcass	< 0.00350	0.00350	< 0.000828	0.000828
	1-08-2	Partial Carcass	< 0.00313	0.00313	< 0.000820	0.000820
	1-08-3	Partial Carcass	< 0.00300	0.00300	< 0.000821	0.000821
	1-08-4	Partial Carcass	< 0.00319	0.00319	< 0.000826	0.000826
	1-08-6	Partial Carcass	< 0.00302	0.00302	< 0.000806	0.000806
C2-naphthalenes						
	1-08-1	Partial Carcass	0.00973	0.00845	0.00230	0.00200
	1-08-2	Partial Carcass	0.0141	0.00755	0.00370	0.00198
	1-08-3	Partial Carcass	0.0139	0.00724	0.00380	0.00198
	1-08-4	Partial Carcass	0.0139	0.00769	0.00360	0.00199
	1-08-6	Partial Carcass	0.0101	0.00729	0.00270	0.00194
C3-Phenanthrenes & Anthracenes						
	1-08-1	Partial Carcass	< 0.00252	0.00252	< 0.000597	0.000597
	1-08-2	Partial Carcass	< 0.00226	0.00226	< 0.000591	0.000591
	1-08-3	Partial Carcass	< 0.00216	0.00216	< 0.000592	0.000592
	1-08-4	Partial Carcass	< 0.00230	0.00230	< 0.000596	0.000596
	1-08-6	Partial Carcass	< 0.00218	0.00218	< 0.000581	0.000581
C3-chrysenes						
	1-08-1	Partial Carcass	< 0.00397	0.00397	< 0.000938	0.000938
	1-08-2	Partial Carcass	< 0.00354	0.00354	< 0.000928	0.000928
	1-08-3	Partial Carcass	< 0.00340	0.00340	< 0.000930	0.000930
	1-08-4	Partial Carcass	< 0.00361	0.00361	< 0.000936	0.000936
	1-08-6	Partial Carcass	< 0.00342	0.00342	< 0.000912	0.000912
C3-dibenzothiophenes						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-1	Partial Carcass	< 0.00288	0.00288	< 0.000682	0.000682
	1-08-2	Partial Carcass	< 0.00258	0.00258	< 0.000675	0.000675
	1-08-3	Partial Carcass	< 0.00247	0.00247	< 0.000677	0.000677
	1-08-4	Partial Carcass	< 0.00262	0.00262	< 0.000681	0.000681
	1-08-6	Partial Carcass	< 0.00249	0.00249	< 0.000664	0.000664
C3-fluorenes						
	1-08-1	Partial Carcass	< 0.00350	0.00350	< 0.000828	0.000828
	1-08-2	Partial Carcass	< 0.00313	0.00313	< 0.000820	0.000820
	1-08-3	Partial Carcass	< 0.00300	0.00300	< 0.000821	0.000821
	1-08-4	Partial Carcass	< 0.00319	0.00319	< 0.000826	0.000826
	1-08-6	Partial Carcass	< 0.00302	0.00302	< 0.000806	0.000806
C3-naphthalenes						
	1-08-1	Partial Carcass	< 0.00845	0.00845	< 0.00200	0.00200
	1-08-2	Partial Carcass	< 0.00755	0.00755	< 0.00198	0.00198
	1-08-3	Partial Carcass	0.00950	0.00724	0.00260	0.00198
	1-08-4	Partial Carcass	0.0112	0.00769	0.00290	0.00199
	1-08-6	Partial Carcass	< 0.00729	0.00729	< 0.00194	0.00194
C4-Phenanthrenes & Anthracenes						
	1-08-1	Partial Carcass	< 0.00252	0.00252	< 0.000597	0.000597
	1-08-2	Partial Carcass	< 0.00226	0.00226	< 0.000591	0.000591
	1-08-3	Partial Carcass	< 0.00216	0.00216	< 0.000592	0.000592
	1-08-4	Partial Carcass	< 0.00230	0.00230	< 0.000596	0.000596
	1-08-6	Partial Carcass	< 0.00218	0.00218	< 0.000581	0.000581
C4-chrysenes						
	1-08-1	Partial Carcass	< 0.00397	0.00397	< 0.000938	0.000938
	1-08-2	Partial Carcass	< 0.00354	0.00354	< 0.000928	0.000928
	1-08-3	Partial Carcass	< 0.00340	0.00340	< 0.000930	0.000930
	1-08-4	Partial Carcass	< 0.00361	0.00361	< 0.000936	0.000936
	1-08-6	Partial Carcass	< 0.00342	0.00342	< 0.000912	0.000912

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
C4-naphthalenes						
	1-08-1	Partial Carcass	< 0.00845	0.00845	< 0.00200	0.00200
	1-08-2	Partial Carcass	< 0.00755	0.00755	< 0.00198	0.00198
	1-08-3	Partial Carcass	< 0.00724	0.00724	< 0.00198	0.00198
	1-08-4	Partial Carcass	< 0.00769	0.00769	< 0.00199	0.00199
	1-08-6	Partial Carcass	< 0.00729	0.00729	< 0.00194	0.00194
Cl4-PCDD						
	10-06-1	Partial Carcass	< 0.00000686	0.00000686	< 0.00000183	0.00000183
	10-06-2	Partial Carcass	< 0.00000743	0.00000743	< 0.00000189	0.00000189
	10-06-3	Partial Carcass	< 0.00000653	0.00000653	< 0.00000196	0.00000196
	10-06-4	Partial Carcass	< 0.00000690	0.00000690	< 0.00000198	0.00000198
	10-07-1	Partial Carcass	< 0.00000702	0.00000702	< 0.00000199	0.00000199
	10-07-2	Partial Carcass	< 0.00000690	0.00000690	< 0.00000197	0.00000197
	10-07-3	Partial Carcass	< 0.00000664	0.00000664	< 0.00000195	0.00000195
	10-07-4	Partial Carcass	< 0.00000797	0.00000797	< 0.00000188	0.00000188
Cl4-PCDF						
	10-06-1	Partial Carcass	< 0.00000686	0.00000686	< 0.00000183	0.00000183
	10-06-2	Partial Carcass	< 0.00000743	0.00000743	< 0.00000189	0.00000189
	10-06-3	Partial Carcass	< 0.00000653	0.00000653	< 0.00000196	0.00000196
	10-06-4	Partial Carcass	< 0.00000690	0.00000690	< 0.00000198	0.00000198
	10-07-1	Partial Carcass	< 0.00000702	0.00000702	< 0.00000199	0.00000199
	10-07-2	Partial Carcass	< 0.00000690	0.00000690	< 0.00000197	0.00000197
	10-07-3	Partial Carcass	< 0.00000664	0.00000664	< 0.00000195	0.00000195
	10-07-4	Partial Carcass	< 0.00000797	0.00000797	< 0.00000188	0.00000188
Cl5-PCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
Cl5-PCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
Cl6-PCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
Cl6-PCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
Cl7-PCDD						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
Cl7-PCDF						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
DDMU						
	*10-06-1	Partial Carcass	0.00264	0.000481	0.000703	0.000128
	*10-06-2	Partial Carcass	0.00168	0.000504	0.000427	0.000128
	*10-06-3	Partial Carcass	0.00213	0.000428	0.000638	0.000128
	*10-06-4	Partial Carcass	0.00263	0.000447	0.000754	0.000128
	*10-06-5	Partial Carcass	0.000735	0.000426	0.000221	0.000128
	*10-07-1	Partial Carcass	< 0.000453	0.000453	< 0.000128	0.000128
	*10-07-2	Partial Carcass	< 0.000449	0.000449	< 0.000128	0.000128
	*10-07-3	Partial Carcass	< 0.000437	0.000437	< 0.000128	0.000128

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-4	Partial Carcass	0.00258	0.000544	0.000609	0.000128
	*10-07-5	Partial Carcass	0.00129	0.000487	0.000340	0.000128
	*10-07-6	Partial Carcass	0.00425	0.000502	0.00108	0.000128
	*4-08-6	Partial Carcass	0.00253	0.000508	0.000639	0.000128
Dibenz(a,h)anthracene						
	1-08-1	Partial Carcass	< 0.00180	0.00180	< 0.000426	0.000426
	1-08-2	Partial Carcass	< 0.00161	0.00161	< 0.000422	0.000422
	1-08-3	Partial Carcass	< 0.00154	0.00154	< 0.000423	0.000423
	1-08-4	Partial Carcass	< 0.00164	0.00164	< 0.000425	0.000425
	1-08-6	Partial Carcass	< 0.00156	0.00156	< 0.000415	0.000415
HCB						
	*10-06-1	Partial Carcass	0.00220	0.000846	0.000586	0.000226
	*10-06-2	Partial Carcass	0.000988	0.000887	0.000251	0.000226
	*10-06-3	Partial Carcass	0.00238	0.000752	0.000714	0.000226
	*10-06-4	Partial Carcass	0.00154	0.000786	0.000442	0.000226
	*10-06-5	Partial Carcass	0.00132	0.000750	0.000398	0.000226
	*10-07-1	Partial Carcass	< 0.000796	0.000796	< 0.000226	0.000226
	*10-07-2	Partial Carcass	0.00102	0.000789	0.000290	0.000226
	*10-07-3	Partial Carcass	0.00274	0.000768	0.000805	0.000226
	*10-07-4	Partial Carcass	0.00154	0.000957	0.000363	0.000226
	*10-07-5	Partial Carcass	< 0.000857	0.000857	< 0.000226	0.000226
	*10-07-6	Partial Carcass	0.00121	0.000883	0.000308	0.000226
	*4-08-6	Partial Carcass	0.00155	0.000893	0.000391	0.000226
Heptachlor						
	*10-06-1	Partial Carcass	< 0.000498	0.000498	< 0.000133	0.000133
	*10-06-2	Partial Carcass	< 0.000522	0.000522	< 0.000133	0.000133
	*10-06-3	Partial Carcass	< 0.000443	0.000443	< 0.000133	0.000133
	*10-06-4	Partial Carcass	< 0.000463	0.000463	< 0.000133	0.000133
	*10-06-5	Partial Carcass	< 0.000442	0.000442	< 0.000133	0.000133

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-1	Partial Carcass	< 0.000469	0.000469	< 0.000133	0.000133
	*10-07-2	Partial Carcass	< 0.000465	0.000465	< 0.000133	0.000133
	*10-07-3	Partial Carcass	< 0.000452	0.000452	< 0.000133	0.000133
	*10-07-4	Partial Carcass	< 0.000563	0.000563	< 0.000133	0.000133
	*10-07-5	Partial Carcass	< 0.000505	0.000505	< 0.000133	0.000133
	*10-07-6	Partial Carcass	< 0.000520	0.000520	< 0.000133	0.000133
	*4-08-6	Partial Carcass	< 0.000526	0.000526	< 0.000133	0.000133
OCDD						
	10-06-1	Partial Carcass	< 0.0000686	0.0000686	< 0.0000183	0.0000183
	10-06-2	Partial Carcass	< 0.0000743	0.0000743	< 0.0000189	0.0000189
	10-06-3	Partial Carcass	< 0.0000653	0.0000653	< 0.0000196	0.0000196
	10-06-4	Partial Carcass	< 0.0000690	0.0000690	< 0.0000198	0.0000198
	10-07-1	Partial Carcass	< 0.0000702	0.0000702	< 0.0000199	0.0000199
	10-07-2	Partial Carcass	< 0.0000690	0.0000690	< 0.0000197	0.0000197
	10-07-3	Partial Carcass	< 0.0000664	0.0000664	< 0.0000195	0.0000195
	10-07-4	Partial Carcass	< 0.0000797	0.0000797	< 0.0000188	0.0000188
OCDF						
	10-06-1	Partial Carcass	< 0.0000686	0.0000686	< 0.0000183	0.0000183
	10-06-2	Partial Carcass	< 0.0000743	0.0000743	< 0.0000189	0.0000189
	10-06-3	Partial Carcass	< 0.0000653	0.0000653	< 0.0000196	0.0000196
	10-06-4	Partial Carcass	< 0.0000690	0.0000690	< 0.0000198	0.0000198
	10-07-1	Partial Carcass	< 0.0000702	0.0000702	< 0.0000199	0.0000199
	10-07-2	Partial Carcass	< 0.0000690	0.0000690	< 0.0000197	0.0000197
	10-07-3	Partial Carcass	< 0.0000664	0.0000664	< 0.0000195	0.0000195
	10-07-4	Partial Carcass	< 0.0000797	0.0000797	< 0.0000188	0.0000188
PCB# 1						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 10						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-06-5	Partial Carcass	< 0.0000271	0.0000271	< 0.00000815	0.00000815
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
	10-07-5	Partial Carcass	< 0.0000304	0.0000304	< 0.00000801	0.00000801
	10-07-6	Partial Carcass	< 0.0000304	0.0000304	< 0.00000777	0.00000777
	4-08-6	Partial Carcass	< 0.0000298	0.0000298	< 0.00000753	0.00000753
	06385810	Water			< 0.0000000649	0.0000000649
PCB# 103						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 104						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 105						
	10-06-1	Partial Carcass	0.000465	0.000138	0.000124	0.0000367
	10-06-2	Partial Carcass	0.000252	0.000148	0.0000642	0.0000377
	10-06-3	Partial Carcass	0.000373	0.000131	0.000112	0.0000392
	10-06-4	Partial Carcass	0.000676	0.000138	0.000194	0.0000395
	10-06-5	Partial Carcass	0.000416	0.000108	0.000125	0.0000326

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-1	Partial Carcass	0.000218	0.000140	0.0000617	0.0000398
	10-07-2	Partial Carcass	0.000431	0.000138	0.000123	0.0000393
	10-07-3	Partial Carcass	0.00214	0.000133	0.000628	0.0000390
	10-07-4	Partial Carcass	0.00151	0.000160	0.000355	0.0000377
	10-07-5	Partial Carcass	0.000483	0.000122	0.000127	0.0000320
	10-07-6	Partial Carcass	0.000205	0.000122	0.0000524	0.0000311
	4-08-6	Partial Carcass	0.000305	0.000119	0.0000770	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 106						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 107						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 108/124						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 11						
	10-06-1	Partial Carcass	0.00230	0.000138	0.000613	0.0000367
	10-06-2	Partial Carcass	0.00250	0.000148	0.000635	0.0000377
	10-06-3	Partial Carcass	0.00220	0.000131	0.000659	0.0000392
	10-06-4	Partial Carcass	0.00239	0.000138	0.000687	0.0000395
	10-06-5	Partial Carcass	0.00190	0.000108	0.000571	0.0000326
	10-07-1	Partial Carcass	0.00239	0.000140	0.000676	0.0000398
	10-07-2	Partial Carcass	0.00284	0.000138	0.000810	0.0000393

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-3	Partial Carcass	0.00247	0.000133	0.000725	0.0000390
	10-07-4	Partial Carcass	0.00277	0.000160	0.000653	0.0000377
	10-07-5	Partial Carcass	0.00248	0.000122	0.000652	0.0000320
	10-07-6	Partial Carcass	0.00233	0.000122	0.000594	0.0000311
	4-08-6	Partial Carcass	0.00209	0.000119	0.000527	0.0000301
	06385810	Water			0.00000330	0.000000260
PCB# 110/115						
	10-06-1	Partial Carcass	0.00129	0.000686	0.000344	0.000183
	10-06-2	Partial Carcass	0.00100	0.000743	0.000255	0.000189
	10-06-3	Partial Carcass	0.00145	0.000653	0.000434	0.000196
	10-06-4	Partial Carcass	0.00252	0.000690	0.000723	0.000198
	10-06-5	Partial Carcass	0.00141	0.000542	0.000423	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	0.00128	0.000690	0.000367	0.000197
	10-07-3	Partial Carcass	0.00664	0.000664	0.00195	0.000195
	10-07-4	Partial Carcass	0.00475	0.000797	0.00112	0.000188
	10-07-5	Partial Carcass	0.00168	0.000608	0.000441	0.000160
	10-07-6	Partial Carcass	0.000736	0.000607	0.000188	0.000155
	4-08-6	Partial Carcass	0.00120	0.000598	0.000303	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 111						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 112						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 114						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 117						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 118						
	10-06-1	Partial Carcass	0.00138	0.000344	0.000367	0.0000917
	10-06-2	Partial Carcass	0.000696	0.000371	0.000177	0.0000943
	10-06-3	Partial Carcass	0.00106	0.000327	0.000319	0.0000980
	10-06-4	Partial Carcass	0.00188	0.000344	0.000540	0.0000988
	10-06-5	Partial Carcass	0.00112	0.000271	0.000337	0.0000815
	10-07-1	Partial Carcass	0.000512	0.000351	0.000145	0.0000994
	10-07-2	Partial Carcass	0.00117	0.000344	0.000333	0.0000983
	10-07-3	Partial Carcass	0.00565	0.000332	0.00166	0.0000976
	10-07-4	Partial Carcass	0.00419	0.000399	0.000987	0.0000942
	10-07-5	Partial Carcass	0.00132	0.000304	0.000347	0.0000801

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-6	Partial Carcass	0.000572	0.000304	0.000146	0.0000777
	4-08-6	Partial Carcass	0.000931	0.000298	0.000235	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 12/13						
	10-06-1	Partial Carcass	< 0.0000686	0.0000686	< 0.0000183	0.0000183
	10-06-2	Partial Carcass	< 0.0000743	0.0000743	< 0.0000189	0.0000189
	10-06-3	Partial Carcass	< 0.0000653	0.0000653	< 0.0000196	0.0000196
	10-06-4	Partial Carcass	< 0.0000690	0.0000690	< 0.0000198	0.0000198
	10-06-5	Partial Carcass	< 0.0000542	0.0000542	< 0.0000163	0.0000163
	10-07-1	Partial Carcass	< 0.0000702	0.0000702	< 0.0000199	0.0000199
	10-07-2	Partial Carcass	< 0.0000690	0.0000690	< 0.0000197	0.0000197
	10-07-3	Partial Carcass	< 0.0000664	0.0000664	< 0.0000195	0.0000195
	10-07-4	Partial Carcass	< 0.0000797	0.0000797	< 0.0000188	0.0000188
	10-07-5	Partial Carcass	< 0.0000608	0.0000608	< 0.0000160	0.0000160
	10-07-6	Partial Carcass	< 0.0000607	0.0000607	< 0.0000155	0.0000155
	4-08-6	Partial Carcass	< 0.0000598	0.0000598	< 0.0000151	0.0000151
	06385810	Water			< 0.000000130	0.000000130
PCB# 120						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 121						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 122						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	06385810	Water			< 0.000000649	0.000000649
PCB# 123						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 126						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
PCB# 127						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 128/166						
	10-06-1	Partial Carcass	0.000397	0.000344	0.000106	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	0.000383	0.000327	0.000115	0.0000980
	10-06-4	Partial Carcass	0.000731	0.000344	0.000210	0.0000988
	10-06-5	Partial Carcass	0.000449	0.000271	0.000135	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.00152	0.000332	0.000446	0.0000976
	10-07-4	Partial Carcass	0.00124	0.000399	0.000293	0.0000942
	10-07-5	Partial Carcass	0.000441	0.000304	0.000116	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.000304	0.000298	0.0000767	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 129/138/163						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-1	Partial Carcass	0.00330	0.000344	0.000879	0.0000917
	10-06-2	Partial Carcass	0.00349	0.000371	0.000887	0.0000943
	10-06-3	Partial Carcass	0.00433	0.000327	0.00130	0.0000980
	10-06-4	Partial Carcass	0.00825	0.000344	0.00237	0.0000988
	10-06-5	Partial Carcass	0.00452	0.000271	0.00136	0.0000815
	10-07-1	Partial Carcass	0.000935	0.000351	0.000265	0.0000994
	10-07-2	Partial Carcass	0.00228	0.000344	0.000650	0.0000983
	10-07-3	Partial Carcass	0.0119	0.000332	0.00350	0.0000976
	10-07-4	Partial Carcass	0.0103	0.000399	0.00242	0.0000942
	10-07-5	Partial Carcass	0.00395	0.000304	0.00104	0.0000801
	10-07-6	Partial Carcass	0.00135	0.000304	0.000345	0.0000777
	4-08-6	Partial Carcass	0.00305	0.000298	0.000769	0.0000753
	06385810	Water			0.000000893	0.000000649
PCB# 130						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	0.000359	0.000344	0.000103	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000630	0.000332	0.000185	0.0000976
	10-07-4	Partial Carcass	0.000526	0.000399	0.000124	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 131						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 132						
	10-06-1	Partial Carcass	0.000547	0.000344	0.000146	0.0000917
	10-06-2	Partial Carcass	0.000853	0.000371	0.000217	0.0000943
	10-06-3	Partial Carcass	0.00102	0.000327	0.000307	0.0000980
	10-06-4	Partial Carcass	0.00184	0.000344	0.000529	0.0000988
	10-06-5	Partial Carcass	0.000964	0.000271	0.000290	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	0.000578	0.000344	0.000165	0.0000983
	10-07-3	Partial Carcass	0.00300	0.000332	0.000881	0.0000976
	10-07-4	Partial Carcass	0.00238	0.000399	0.000562	0.0000942
	10-07-5	Partial Carcass	0.000935	0.000304	0.000246	0.0000801
	10-07-6	Partial Carcass	0.000346	0.000304	0.0000884	0.0000777
	4-08-6	Partial Carcass	0.000689	0.000298	0.000174	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 133						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 134						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000416	0.000332	0.000122	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 135/151						
	10-06-1	Partial Carcass	0.000615	0.000344	0.000164	0.0000917
	10-06-2	Partial Carcass	0.00101	0.000371	0.000258	0.0000943
	10-06-3	Partial Carcass	0.00114	0.000327	0.000343	0.0000980

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-4	Partial Carcass	0.00202	0.000344	0.000579	0.0000988
	10-06-5	Partial Carcass	0.000954	0.000271	0.000287	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	0.000427	0.000344	0.000122	0.0000983
	10-07-3	Partial Carcass	0.00229	0.000332	0.000671	0.0000976
	10-07-4	Partial Carcass	0.00183	0.000399	0.000432	0.0000942
	10-07-5	Partial Carcass	0.000805	0.000304	0.000212	0.0000801
	10-07-6	Partial Carcass	0.000305	0.000304	0.0000778	0.0000777
	4-08-6	Partial Carcass	0.000717	0.000298	0.000181	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 136						
	10-06-1	Partial Carcass	0.000147	0.000138	0.0000391	0.0000367
	10-06-2	Partial Carcass	0.000285	0.000148	0.0000724	0.0000377
	10-06-3	Partial Carcass	0.000316	0.000131	0.0000948	0.0000392
	10-06-4	Partial Carcass	0.000529	0.000138	0.000152	0.0000395
	10-06-5	Partial Carcass	0.000247	0.000108	0.0000744	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	0.000153	0.000138	0.0000438	0.0000393
	10-07-3	Partial Carcass	0.000725	0.000133	0.000213	0.0000390
	10-07-4	Partial Carcass	0.000530	0.000160	0.000125	0.0000377
	10-07-5	Partial Carcass	0.000233	0.000122	0.0000612	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	0.000201	0.000119	0.0000508	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 137						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 139/140						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 14						
	10-06-1	Partial Carcass	< 0.0000686	0.0000686	< 0.0000183	0.0000183
	10-06-2	Partial Carcass	< 0.0000743	0.0000743	< 0.0000189	0.0000189
	10-06-3	Partial Carcass	< 0.0000653	0.0000653	< 0.0000196	0.0000196
	10-06-4	Partial Carcass	< 0.0000690	0.0000690	< 0.0000198	0.0000198
	10-06-5	Partial Carcass	< 0.0000542	0.0000542	< 0.0000163	0.0000163

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-1	Partial Carcass	< 0.0000702	0.0000702	< 0.0000199	0.0000199
	10-07-2	Partial Carcass	< 0.0000690	0.0000690	< 0.0000197	0.0000197
	10-07-3	Partial Carcass	< 0.0000664	0.0000664	< 0.0000195	0.0000195
	10-07-4	Partial Carcass	< 0.0000797	0.0000797	< 0.0000188	0.0000188
	10-07-5	Partial Carcass	< 0.0000608	0.0000608	< 0.0000160	0.0000160
	10-07-6	Partial Carcass	< 0.0000607	0.0000607	< 0.0000155	0.0000155
	4-08-6	Partial Carcass	< 0.0000598	0.0000598	< 0.0000151	0.0000151
	06385810	Water			< 0.000000130	0.000000130
PCB# 141						
	10-06-1	Partial Carcass	0.000351	0.000138	0.0000937	0.0000367
	10-06-2	Partial Carcass	0.000861	0.000148	0.000219	0.0000377
	10-06-3	Partial Carcass	0.00103	0.000131	0.000309	0.0000392
	10-06-4	Partial Carcass	0.00180	0.000138	0.000516	0.0000395
	10-06-5	Partial Carcass	0.000838	0.000108	0.000252	0.0000326
	10-07-1	Partial Carcass	0.000158	0.000140	0.0000448	0.0000398
	10-07-2	Partial Carcass	0.000396	0.000138	0.000113	0.0000393
	10-07-3	Partial Carcass	0.00221	0.000133	0.000649	0.0000390
	10-07-4	Partial Carcass	0.00191	0.000160	0.000450	0.0000377
	10-07-5	Partial Carcass	0.000748	0.000122	0.000197	0.0000320
	10-07-6	Partial Carcass	0.000228	0.000122	0.0000582	0.0000311
	4-08-6	Partial Carcass	0.000606	0.000119	0.000153	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 142						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 143						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 144						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-3	Partial Carcass	0.000371	0.000332	0.000109	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 145						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 146						
	10-06-1	Partial Carcass	0.000709	0.000344	0.000189	0.0000917
	10-06-2	Partial Carcass	0.000519	0.000371	0.000132	0.0000943
	10-06-3	Partial Carcass	0.000607	0.000327	0.000182	0.0000980
	10-06-4	Partial Carcass	0.00119	0.000344	0.000343	0.0000988
	10-06-5	Partial Carcass	0.000652	0.000271	0.000196	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.00144	0.000332	0.000424	0.0000976

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-4	Partial Carcass	0.00131	0.000399	0.000310	0.0000942
	10-07-5	Partial Carcass	0.000524	0.000304	0.000138	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.000436	0.000298	0.000110	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 147/149						
	10-06-1	Partial Carcass	0.00171	0.000344	0.000456	0.0000917
	10-06-2	Partial Carcass	0.00277	0.000371	0.000705	0.0000943
	10-06-3	Partial Carcass	0.00322	0.000327	0.000966	0.0000980
	10-06-4	Partial Carcass	0.00592	0.000344	0.00170	0.0000988
	10-06-5	Partial Carcass	0.00280	0.000271	0.000843	0.0000815
	10-07-1	Partial Carcass	0.000561	0.000351	0.000159	0.0000994
	10-07-2	Partial Carcass	0.00137	0.000344	0.000390	0.0000983
	10-07-3	Partial Carcass	0.00712	0.000332	0.00209	0.0000976
	10-07-4	Partial Carcass	0.00602	0.000399	0.00142	0.0000942
	10-07-5	Partial Carcass	0.00245	0.000304	0.000645	0.0000801
	10-07-6	Partial Carcass	0.000866	0.000304	0.000221	0.0000777
	4-08-6	Partial Carcass	0.00207	0.000298	0.000522	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 148						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 15						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 150						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 152						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 153/168						
	10-06-1	Partial Carcass	0.00364	0.000344	0.000970	0.0000917
	10-06-2	Partial Carcass	0.00401	0.000371	0.00102	0.0000943
	10-06-3	Partial Carcass	0.00480	0.000327	0.00144	0.0000980
	10-06-4	Partial Carcass	0.00902	0.000344	0.00259	0.0000988
	10-06-5	Partial Carcass	0.00475	0.000271	0.00143	0.0000815
	10-07-1	Partial Carcass	0.000875	0.000351	0.000248	0.0000994
	10-07-2	Partial Carcass	0.00200	0.000344	0.000571	0.0000983
	10-07-3	Partial Carcass	0.0104	0.000332	0.00305	0.0000976
	10-07-4	Partial Carcass	0.00954	0.000399	0.00225	0.0000942
	10-07-5	Partial Carcass	0.00380	0.000304	0.00100	0.0000801
	10-07-6	Partial Carcass	0.00137	0.000304	0.000351	0.0000777

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	0.00321	0.000298	0.000810	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 154						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 155						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	06385810	Water			< 0.00000130	0.00000130
PCB# 156/157						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	0.000623	0.000344	0.000179	0.0000988
	10-06-5	Partial Carcass	0.000349	0.000271	0.000105	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.00129	0.000332	0.000378	0.0000976
	10-07-4	Partial Carcass	0.00106	0.000399	0.000249	0.0000942
	10-07-5	Partial Carcass	0.000351	0.000304	0.0000923	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 158						
	10-06-1	Partial Carcass	0.000182	0.000138	0.0000486	0.0000367
	10-06-2	Partial Carcass	0.000289	0.000148	0.0000734	0.0000377
	10-06-3	Partial Carcass	0.000367	0.000131	0.000110	0.0000392
	10-06-4	Partial Carcass	0.000721	0.000138	0.000207	0.0000395
	10-06-5	Partial Carcass	0.000379	0.000108	0.000114	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	0.000227	0.000138	0.0000648	0.0000393
	10-07-3	Partial Carcass	0.00115	0.000133	0.000337	0.0000390
	10-07-4	Partial Carcass	0.000963	0.000160	0.000227	0.0000377
	10-07-5	Partial Carcass	0.000353	0.000122	0.0000930	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	0.000259	0.000119	0.0000654	0.0000301
	06385810	Water			< 0.000000260	0.000000260

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
PCB# 159						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 16						
	10-06-1	Partial Carcass	0.000157	0.0000686	0.0000420	0.0000183
	10-06-2	Partial Carcass	0.000133	0.0000743	0.0000338	0.0000189
	10-06-3	Partial Carcass	0.000153	0.0000653	0.0000458	0.0000196
	10-06-4	Partial Carcass	0.000176	0.0000690	0.0000505	0.0000198
	10-06-5	Partial Carcass	0.000100	0.0000542	0.0000302	0.0000163
	10-07-1	Partial Carcass	0.000173	0.0000702	0.0000490	0.0000199
	10-07-2	Partial Carcass	0.000135	0.0000690	0.0000385	0.0000197
	10-07-3	Partial Carcass	0.000156	0.0000664	0.0000459	0.0000195
	10-07-4	Partial Carcass	0.000146	0.0000797	0.0000345	0.0000188
	10-07-5	Partial Carcass	0.000119	0.0000608	0.0000314	0.0000160
	10-07-6	Partial Carcass	0.000112	0.0000607	0.0000285	0.0000155
	4-08-6	Partial Carcass	0.0000915	0.0000598	0.0000231	0.0000151
	06385810	Water			0.000000210	0.000000130
PCB# 160						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 161						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 162						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 164						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	0.000522	0.000344	0.000150	0.0000988
	10-06-5	Partial Carcass	0.000287	0.000271	0.0000862	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000729	0.000332	0.000214	0.0000976
	10-07-4	Partial Carcass	0.000619	0.000399	0.000146	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 165						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 167						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000470	0.000332	0.000138	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 169						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 17						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	0.000153	0.000138	0.0000438	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	0.000137	0.000133	0.0000403	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 170						
	10-06-1	Partial Carcass	0.000615	0.000344	0.000164	0.0000917
	10-06-2	Partial Carcass	0.00136	0.000371	0.000346	0.0000943
	10-06-3	Partial Carcass	0.00170	0.000327	0.000511	0.0000980
	10-06-4	Partial Carcass	0.00327	0.000344	0.000940	0.0000988

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-5	Partial Carcass	0.00157	0.000271	0.000471	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	0.000445	0.000344	0.000127	0.0000983
	10-07-3	Partial Carcass	0.00329	0.000332	0.000967	0.0000976
	10-07-4	Partial Carcass	0.00325	0.000399	0.000766	0.0000942
	10-07-5	Partial Carcass	0.00115	0.000304	0.000303	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.000962	0.000298	0.000243	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 171/173						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	0.00103	0.000690	0.000297	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	0.00101	0.000664	0.000296	0.000195
	10-07-4	Partial Carcass	0.000941	0.000797	0.000222	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 172						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	0.000693	0.000690	0.000199	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 174						
	10-06-1	Partial Carcass	0.000502	0.000344	0.000134	0.0000917
	10-06-2	Partial Carcass	0.00145	0.000371	0.000368	0.0000943
	10-06-3	Partial Carcass	0.00180	0.000327	0.000539	0.0000980
	10-06-4	Partial Carcass	0.00327	0.000344	0.000938	0.0000988
	10-06-5	Partial Carcass	0.00158	0.000271	0.000476	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	0.000417	0.000344	0.000119	0.0000983
	10-07-3	Partial Carcass	0.00305	0.000332	0.000895	0.0000976
	10-07-4	Partial Carcass	0.00285	0.000399	0.000671	0.0000942
	10-07-5	Partial Carcass	0.00118	0.000304	0.000311	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.00102	0.000298	0.000258	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 175						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 176						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 177						
	10-06-1	Partial Carcass	0.000367	0.000344	0.0000979	0.0000917
	10-06-2	Partial Carcass	0.000672	0.000371	0.000171	0.0000943
	10-06-3	Partial Carcass	0.000813	0.000327	0.000244	0.0000980
	10-06-4	Partial Carcass	0.00176	0.000344	0.000504	0.0000988
	10-06-5	Partial Carcass	0.000795	0.000271	0.000239	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-3	Partial Carcass	0.00148	0.000332	0.000435	0.0000976
	10-07-4	Partial Carcass	0.00162	0.000399	0.000383	0.0000942
	10-07-5	Partial Carcass	0.000604	0.000304	0.000159	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.000487	0.000298	0.000123	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 178						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	0.000627	0.000344	0.000180	0.0000988
	10-06-5	Partial Carcass	0.000321	0.000271	0.0000967	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000511	0.000332	0.000150	0.0000976
	10-07-4	Partial Carcass	0.000509	0.000399	0.000120	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 179						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	0.000425	0.000371	0.000108	0.0000943
	10-06-3	Partial Carcass	0.000487	0.000327	0.000146	0.0000980
	10-06-4	Partial Carcass	0.000878	0.000344	0.000252	0.0000988
	10-06-5	Partial Carcass	0.000422	0.000271	0.000127	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000773	0.000332	0.000227	0.0000976

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-4	Partial Carcass	0.000670	0.000399	0.000158	0.0000942
	10-07-5	Partial Carcass	0.000310	0.000304	0.0000815	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 18/30						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 180/193						
	10-06-1	Partial Carcass	0.00166	0.000344	0.000444	0.0000917
	10-06-2	Partial Carcass	0.00386	0.000371	0.000981	0.0000943
	10-06-3	Partial Carcass	0.00467	0.000327	0.00140	0.0000980
	10-06-4	Partial Carcass	0.00927	0.000344	0.00266	0.0000988
	10-06-5	Partial Carcass	0.00439	0.000271	0.00132	0.0000815
	10-07-1	Partial Carcass	0.000784	0.000351	0.000222	0.0000994
	10-07-2	Partial Carcass	0.00115	0.000344	0.000329	0.0000983
	10-07-3	Partial Carcass	0.00824	0.000332	0.00242	0.0000976
	10-07-4	Partial Carcass	0.00835	0.000399	0.00197	0.0000942

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-5	Partial Carcass	0.00312	0.000304	0.000821	0.0000801
	10-07-6	Partial Carcass	0.000705	0.000304	0.000180	0.0000777
	4-08-6	Partial Carcass	0.00270	0.000298	0.000682	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 181						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 182						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 183						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	0.000987	0.000743	0.000251	0.000189
	10-06-3	Partial Carcass	0.00116	0.000653	0.000348	0.000196
	10-06-4	Partial Carcass	0.00225	0.000690	0.000647	0.000198
	10-06-5	Partial Carcass	0.00108	0.000542	0.000324	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	0.00198	0.000664	0.000582	0.000195
	10-07-4	Partial Carcass	0.00202	0.000797	0.000476	0.000188
	10-07-5	Partial Carcass	0.000790	0.000608	0.000208	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	0.000685	0.000598	0.000173	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 184						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 185						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 186						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	06385810	Water			< 0.00000130	0.00000130
PCB# 187						
	10-06-1	Partial Carcass	0.00143	0.000344	0.000382	0.0000917
	10-06-2	Partial Carcass	0.00185	0.000371	0.000470	0.0000943
	10-06-3	Partial Carcass	0.00218	0.000327	0.000655	0.0000980
	10-06-4	Partial Carcass	0.00432	0.000344	0.00124	0.0000988
	10-06-5	Partial Carcass	0.00223	0.000271	0.000670	0.0000815
	10-07-1	Partial Carcass	0.000392	0.000351	0.000111	0.0000994
	10-07-2	Partial Carcass	0.000550	0.000344	0.000157	0.0000983
	10-07-3	Partial Carcass	0.00347	0.000332	0.00102	0.0000976
	10-07-4	Partial Carcass	0.00364	0.000399	0.000858	0.0000942
	10-07-5	Partial Carcass	0.00146	0.000304	0.000384	0.0000801
	10-07-6	Partial Carcass	0.000431	0.000304	0.000110	0.0000777
	4-08-6	Partial Carcass	0.00134	0.000298	0.000338	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 188						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
PCB# 189						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 19						
	10-06-1	Partial Carcass	0.000106	0.0000686	0.0000284	0.0000183
	10-06-2	Partial Carcass	< 0.0000743	0.0000743	< 0.0000189	0.0000189
	10-06-3	Partial Carcass	< 0.0000653	0.0000653	< 0.0000196	0.0000196
	10-06-4	Partial Carcass	< 0.0000690	0.0000690	< 0.0000198	0.0000198
	10-06-5	Partial Carcass	< 0.0000542	0.0000542	< 0.0000163	0.0000163
	10-07-1	Partial Carcass	0.0000995	0.0000702	0.0000282	0.0000199
	10-07-2	Partial Carcass	< 0.0000690	0.0000690	< 0.0000197	0.0000197
	10-07-3	Partial Carcass	< 0.0000664	0.0000664	< 0.0000195	0.0000195
	10-07-4	Partial Carcass	< 0.0000797	0.0000797	< 0.0000188	0.0000188
	10-07-5	Partial Carcass	< 0.0000608	0.0000608	< 0.0000160	0.0000160
	10-07-6	Partial Carcass	< 0.0000607	0.0000607	< 0.0000155	0.0000155
	4-08-6	Partial Carcass	< 0.0000598	0.0000598	< 0.0000151	0.0000151
	06385810	Water			< 0.000000130	0.000000130
PCB# 190						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	0.000383	0.000327	0.000115	0.0000980
	10-06-4	Partial Carcass	0.000724	0.000344	0.000208	0.0000988
	10-06-5	Partial Carcass	0.000376	0.000271	0.000113	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000664	0.000332	0.000195	0.0000976
	10-07-4	Partial Carcass	0.000700	0.000399	0.000165	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 191						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 192						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 194						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	0.000378	0.000371	0.0000962	0.0000943
	10-06-3	Partial Carcass	0.000543	0.000327	0.000163	0.0000980
	10-06-4	Partial Carcass	0.000996	0.000344	0.000286	0.0000988
	10-06-5	Partial Carcass	0.000419	0.000271	0.000126	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000845	0.000332	0.000248	0.0000976
	10-07-4	Partial Carcass	0.00110	0.000399	0.000260	0.0000942
	10-07-5	Partial Carcass	0.000335	0.000304	0.0000883	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 195						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 196						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	0.000770	0.000690	0.000221	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 197						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 198/199						
	10-06-1	Partial Carcass	0.000547	0.000344	0.000146	0.0000917
	10-06-2	Partial Carcass	0.000680	0.000371	0.000173	0.0000943
	10-06-3	Partial Carcass	0.000840	0.000327	0.000252	0.0000980
	10-06-4	Partial Carcass	0.00167	0.000344	0.000480	0.0000988
	10-06-5	Partial Carcass	0.000911	0.000271	0.000274	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.00137	0.000332	0.000403	0.0000976
	10-07-4	Partial Carcass	0.00152	0.000399	0.000358	0.0000942
	10-07-5	Partial Carcass	0.000581	0.000304	0.000153	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.000495	0.000298	0.000125	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 2						
	10-06-1	Partial Carcass	0.0000115	0.00000686	0.00000308	0.00000183
	10-06-2	Partial Carcass	0.0000112	0.00000743	0.00000286	0.00000189
	10-06-3	Partial Carcass	0.0000121	0.00000653	0.00000363	0.00000196
	10-06-4	Partial Carcass	0.0000140	0.00000690	0.00000401	0.00000198

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-5	Partial Carcass	0.00000894	0.00000542	0.00000269	0.00000163
	10-07-1	Partial Carcass	0.0000118	0.00000702	0.00000335	0.00000199
	10-07-2	Partial Carcass	0.0000163	0.00000690	0.00000467	0.00000197
	10-07-3	Partial Carcass	0.0000132	0.00000664	0.00000387	0.00000195
	10-07-4	Partial Carcass	0.0000168	0.00000797	0.00000397	0.00000188
	10-07-5	Partial Carcass	0.0000119	0.00000608	0.00000314	0.00000160
	10-07-6	Partial Carcass	0.0000113	0.00000607	0.00000289	0.00000155
	4-08-6	Partial Carcass	0.00000966	0.00000598	0.00000244	0.00000151
	06385810	Water			0.0000000264	0.0000000130
PCB# 20/28						
	10-06-1	Partial Carcass	0.000465	0.000344	0.000124	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	0.000540	0.000327	0.000162	0.0000980
	10-06-4	Partial Carcass	0.000724	0.000344	0.000208	0.0000988
	10-06-5	Partial Carcass	0.000300	0.000271	0.0000903	0.0000815
	10-07-1	Partial Carcass	0.000582	0.000351	0.000165	0.0000994
	10-07-2	Partial Carcass	0.000508	0.000344	0.000145	0.0000983
	10-07-3	Partial Carcass	0.000896	0.000332	0.000263	0.0000976
	10-07-4	Partial Carcass	0.000450	0.000399	0.000106	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.000331	0.000298	0.0000835	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 200						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 201						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 202						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 203						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	0.00113	0.000690	0.000323	0.000198
	10-06-5	Partial Carcass	0.000602	0.000542	0.000181	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	0.000903	0.000664	0.000265	0.000195
	10-07-4	Partial Carcass	0.00103	0.000797	0.000242	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 204						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 205						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 206						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 207						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 208						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 209						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 21/33						
	10-06-1	Partial Carcass	0.000248	0.000138	0.0000661	0.0000367
	10-06-2	Partial Carcass	0.000252	0.000148	0.0000641	0.0000377
	10-06-3	Partial Carcass	0.000257	0.000131	0.0000771	0.0000392
	10-06-4	Partial Carcass	0.000304	0.000138	0.0000873	0.0000395
	10-06-5	Partial Carcass	0.000164	0.000108	0.0000494	0.0000326
	10-07-1	Partial Carcass	0.000257	0.000140	0.0000727	0.0000398
	10-07-2	Partial Carcass	0.000207	0.000138	0.0000591	0.0000393
	10-07-3	Partial Carcass	0.000262	0.000133	0.0000770	0.0000390
	10-07-4	Partial Carcass	0.000227	0.000160	0.0000535	0.0000377
	10-07-5	Partial Carcass	0.000180	0.000122	0.0000473	0.0000320

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-6	Partial Carcass	0.000154	0.000122	0.0000392	0.0000311
	4-08-6	Partial Carcass	0.000135	0.000119	0.0000342	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 22						
	10-06-1	Partial Carcass	0.000252	0.000138	0.0000673	0.0000367
	10-06-2	Partial Carcass	0.000203	0.000148	0.0000516	0.0000377
	10-06-3	Partial Carcass	0.000271	0.000131	0.0000814	0.0000392
	10-06-4	Partial Carcass	0.000390	0.000138	0.000112	0.0000395
	10-06-5	Partial Carcass	0.000164	0.000108	0.0000493	0.0000326
	10-07-1	Partial Carcass	0.000279	0.000140	0.0000790	0.0000398
	10-07-2	Partial Carcass	0.000159	0.000138	0.0000454	0.0000393
	10-07-3	Partial Carcass	0.000324	0.000133	0.0000952	0.0000390
	10-07-4	Partial Carcass	0.000207	0.000160	0.0000487	0.0000377
	10-07-5	Partial Carcass	0.000144	0.000122	0.0000380	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	0.000125	0.000119	0.0000316	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 23						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 24						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 25						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	06385810	Water			< 0.000000260	0.000000260
PCB# 26/29						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 27						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
PCB# 3						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 31						
	10-06-1	Partial Carcass	0.000454	0.000344	0.000121	0.0000917
	10-06-2	Partial Carcass	0.000387	0.000371	0.0000985	0.0000943
	10-06-3	Partial Carcass	0.000517	0.000327	0.000155	0.0000980
	10-06-4	Partial Carcass	0.000665	0.000344	0.000191	0.0000988
	10-06-5	Partial Carcass	0.000311	0.000271	0.0000934	0.0000815
	10-07-1	Partial Carcass	0.000480	0.000351	0.000136	0.0000994
	10-07-2	Partial Carcass	0.000361	0.000344	0.000103	0.0000983
	10-07-3	Partial Carcass	0.000599	0.000332	0.000176	0.0000976
	10-07-4	Partial Carcass	0.000405	0.000399	0.0000954	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 32						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 34						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 35						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 36						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 37						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 38						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 39						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 4						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 40/41/70						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 42						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	0.000193	0.000138	0.0000553	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	0.000224	0.000133	0.0000657	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 43/73						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 44/47/65						
	10-06-1	Partial Carcass	0.000517	0.000344	0.000138	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	0.000520	0.000327	0.000156	0.0000980
	10-06-4	Partial Carcass	0.000759	0.000344	0.000218	0.0000988
	10-06-5	Partial Carcass	0.000376	0.000271	0.000113	0.0000815
	10-07-1	Partial Carcass	0.000434	0.000351	0.000123	0.0000994
	10-07-2	Partial Carcass	0.000392	0.000344	0.000112	0.0000983
	10-07-3	Partial Carcass	0.00119	0.000332	0.000348	0.0000976
	10-07-4	Partial Carcass	0.000683	0.000399	0.000161	0.0000942
	10-07-5	Partial Carcass	0.000350	0.000304	0.0000920	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	0.000301	0.000298	0.0000761	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 45/51						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 46						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 48						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 49/69						
	10-06-1	Partial Carcass	0.000346	0.000344	0.0000922	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	0.000509	0.000344	0.000146	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000749	0.000332	0.000220	0.0000976
	10-07-4	Partial Carcass	0.000428	0.000399	0.000101	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 5						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-06-5	Partial Carcass	< 0.0000271	0.0000271	< 0.00000815	0.00000815
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
	10-07-5	Partial Carcass	< 0.0000304	0.0000304	< 0.00000801	0.00000801
	10-07-6	Partial Carcass	< 0.0000304	0.0000304	< 0.00000777	0.00000777
	4-08-6	Partial Carcass	< 0.0000298	0.0000298	< 0.00000753	0.00000753
	06385810	Water			< 0.0000000649	0.0000000649
PCB# 50/53						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 52						
	10-06-1	Partial Carcass	0.000686	0.000344	0.000183	0.0000917
	10-06-2	Partial Carcass	0.000483	0.000371	0.000123	0.0000943
	10-06-3	Partial Carcass	0.000760	0.000327	0.000228	0.0000980
	10-06-4	Partial Carcass	0.00103	0.000344	0.000296	0.0000988
	10-06-5	Partial Carcass	0.000515	0.000271	0.000155	0.0000815
	10-07-1	Partial Carcass	0.000537	0.000351	0.000152	0.0000994
	10-07-2	Partial Carcass	0.000623	0.000344	0.000178	0.0000983
	10-07-3	Partial Carcass	0.00235	0.000332	0.000691	0.0000976
	10-07-4	Partial Carcass	0.00124	0.000399	0.000292	0.0000942

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-5	Partial Carcass	0.000570	0.000304	0.000150	0.0000801
	10-07-6	Partial Carcass	0.000356	0.000304	0.0000908	0.0000777
	4-08-6	Partial Carcass	0.000459	0.000298	0.000116	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 54						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 55						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 56						
	10-06-1	Partial Carcass	0.000154	0.000138	0.0000411	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	0.000193	0.000131	0.0000580	0.0000392
	10-06-4	Partial Carcass	0.000333	0.000138	0.0000955	0.0000395
	10-06-5	Partial Carcass	0.000137	0.000108	0.0000413	0.0000326
	10-07-1	Partial Carcass	0.000160	0.000140	0.0000454	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	0.000422	0.000133	0.000124	0.0000390
	10-07-4	Partial Carcass	0.000251	0.000160	0.0000593	0.0000377
	10-07-5	Partial Carcass	0.000122	0.000122	0.0000321	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 57						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 58						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 59/62/75						
	10-06-1	Partial Carcass	< 0.000138	0.000138	< 0.0000367	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	< 0.000131	0.000131	< 0.0000392	0.0000392
	10-06-4	Partial Carcass	< 0.000138	0.000138	< 0.0000395	0.0000395
	10-06-5	Partial Carcass	< 0.000108	0.000108	< 0.0000326	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	< 0.000138	0.000138	< 0.0000393	0.0000393
	10-07-3	Partial Carcass	< 0.000133	0.000133	< 0.0000390	0.0000390
	10-07-4	Partial Carcass	< 0.000160	0.000160	< 0.0000377	0.0000377
	10-07-5	Partial Carcass	< 0.000122	0.000122	< 0.0000320	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	< 0.000119	0.000119	< 0.0000301	0.0000301

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	06385810	Water			< 0.000000260	0.000000260
PCB# 6						
	10-06-1	Partial Carcass	0.0000607	0.0000344	0.0000162	0.00000917
	10-06-2	Partial Carcass	0.0000456	0.0000371	0.0000116	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-06-5	Partial Carcass	< 0.0000271	0.0000271	< 0.00000815	0.00000815
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	0.0000484	0.0000332	0.0000142	0.00000976
	10-07-4	Partial Carcass	0.0000505	0.0000399	0.0000119	0.00000942
	10-07-5	Partial Carcass	0.0000456	0.0000304	0.0000120	0.00000801
	10-07-6	Partial Carcass	0.0000462	0.0000304	0.0000118	0.00000777
	4-08-6	Partial Carcass	< 0.0000298	0.0000298	< 0.00000753	0.00000753
	06385810	Water			0.0000000769	0.0000000649
PCB# 60						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
PCB# 63						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 64						
	10-06-1	Partial Carcass	0.000207	0.000138	0.0000552	0.0000367
	10-06-2	Partial Carcass	0.000154	0.000148	0.0000391	0.0000377
	10-06-3	Partial Carcass	0.000255	0.000131	0.0000766	0.0000392
	10-06-4	Partial Carcass	0.000418	0.000138	0.000120	0.0000395
	10-06-5	Partial Carcass	0.000177	0.000108	0.0000532	0.0000326
	10-07-1	Partial Carcass	0.000218	0.000140	0.0000617	0.0000398
	10-07-2	Partial Carcass	0.000162	0.000138	0.0000462	0.0000393
	10-07-3	Partial Carcass	0.000552	0.000133	0.000162	0.0000390
	10-07-4	Partial Carcass	0.000322	0.000160	0.0000760	0.0000377
	10-07-5	Partial Carcass	0.000155	0.000122	0.0000409	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	0.000133	0.000119	0.0000337	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 66						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-1	Partial Carcass	0.000401	0.000344	0.000107	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	0.000393	0.000327	0.000118	0.0000980
	10-06-4	Partial Carcass	0.000648	0.000344	0.000186	0.0000988
	10-06-5	Partial Carcass	0.000292	0.000271	0.0000877	0.0000815
	10-07-1	Partial Carcass	0.000455	0.000351	0.000129	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.00101	0.000332	0.000296	0.0000976
	10-07-4	Partial Carcass	0.000585	0.000399	0.000138	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 67						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 68						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 7						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-06-5	Partial Carcass	< 0.0000271	0.0000271	< 0.00000815	0.00000815
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
	10-07-5	Partial Carcass	< 0.0000304	0.0000304	< 0.00000801	0.00000801
	10-07-6	Partial Carcass	< 0.0000304	0.0000304	< 0.00000777	0.00000777
	4-08-6	Partial Carcass	< 0.0000298	0.0000298	< 0.00000753	0.00000753
	06385810	Water			< 0.0000000649	0.0000000649
PCB# 70/61/74/76						
	10-06-1	Partial Carcass	0.000799	0.000344	0.000213	0.0000917
	10-06-2	Partial Carcass	0.000531	0.000371	0.000135	0.0000943

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-3	Partial Carcass	0.000820	0.000327	0.000246	0.0000980
	10-06-4	Partial Carcass	0.00133	0.000344	0.000383	0.0000988
	10-06-5	Partial Carcass	0.000602	0.000271	0.000181	0.0000815
	10-07-1	Partial Carcass	0.000805	0.000351	0.000228	0.0000994
	10-07-2	Partial Carcass	0.000669	0.000344	0.000191	0.0000983
	10-07-3	Partial Carcass	0.00264	0.000332	0.000776	0.0000976
	10-07-4	Partial Carcass	0.00147	0.000399	0.000347	0.0000942
	10-07-5	Partial Carcass	0.000616	0.000304	0.000162	0.0000801
	10-07-6	Partial Carcass	0.000327	0.000304	0.0000834	0.0000777
	4-08-6	Partial Carcass	0.000467	0.000298	0.000118	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 72						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 77						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 78						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 79						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 8						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 80						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 81						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 82						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000463	0.000332	0.000136	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 83/99						
	10-06-1	Partial Carcass	0.00105	0.000344	0.000281	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	0.000570	0.000327	0.000171	0.0000980
	10-06-4	Partial Carcass	0.000975	0.000344	0.000280	0.0000988
	10-06-5	Partial Carcass	0.000598	0.000271	0.000180	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	0.000599	0.000344	0.000171	0.0000983
	10-07-3	Partial Carcass	0.00281	0.000332	0.000826	0.0000976
	10-07-4	Partial Carcass	0.00195	0.000399	0.000460	0.0000942
	10-07-5	Partial Carcass	0.000661	0.000304	0.000174	0.0000801
	10-07-6	Partial Carcass	0.000370	0.000304	0.0000944	0.0000777
	4-08-6	Partial Carcass	0.000511	0.000298	0.000129	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 84						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-3	Partial Carcass	0.000937	0.000332	0.000275	0.0000976
	10-07-4	Partial Carcass	0.000594	0.000399	0.000140	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 85/116						
	10-06-1	Partial Carcass	0.000191	0.000138	0.0000509	0.0000367
	10-06-2	Partial Carcass	< 0.000148	0.000148	< 0.0000377	0.0000377
	10-06-3	Partial Carcass	0.000157	0.000131	0.0000471	0.0000392
	10-06-4	Partial Carcass	0.000279	0.000138	0.0000801	0.0000395
	10-06-5	Partial Carcass	0.000164	0.000108	0.0000492	0.0000326
	10-07-1	Partial Carcass	< 0.000140	0.000140	< 0.0000398	0.0000398
	10-07-2	Partial Carcass	0.000162	0.000138	0.0000462	0.0000393
	10-07-3	Partial Carcass	0.000804	0.000133	0.000236	0.0000390
	10-07-4	Partial Carcass	0.000547	0.000160	0.000129	0.0000377
	10-07-5	Partial Carcass	0.000196	0.000122	0.0000517	0.0000320
	10-07-6	Partial Carcass	< 0.000122	0.000122	< 0.0000311	0.0000311
	4-08-6	Partial Carcass	0.000123	0.000119	0.0000311	0.0000301
	06385810	Water			< 0.000000260	0.000000260
PCB# 86/87/97/109/119/125						
	10-06-1	Partial Carcass	0.000776	0.000344	0.000207	0.0000917
	10-06-2	Partial Carcass	0.000511	0.000371	0.000130	0.0000943
	10-06-3	Partial Carcass	0.000743	0.000327	0.000223	0.0000980
	10-06-4	Partial Carcass	0.00118	0.000344	0.000339	0.0000988
	10-06-5	Partial Carcass	0.000701	0.000271	0.000211	0.0000815
	10-07-1	Partial Carcass	0.000385	0.000351	0.000109	0.0000994
	10-07-2	Partial Carcass	0.000781	0.000344	0.000223	0.0000983
	10-07-3	Partial Carcass	0.00371	0.000332	0.00109	0.0000976

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-4	Partial Carcass	0.00250	0.000399	0.000590	0.0000942
	10-07-5	Partial Carcass	0.000881	0.000304	0.000232	0.0000801
	10-07-6	Partial Carcass	0.000423	0.000304	0.000108	0.0000777
	4-08-6	Partial Carcass	0.000614	0.000298	0.000155	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 88/91						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.000548	0.000332	0.000161	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 89						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 9						
	10-06-1	Partial Carcass	< 0.0000344	0.0000344	< 0.00000917	0.00000917
	10-06-2	Partial Carcass	< 0.0000371	0.0000371	< 0.00000943	0.00000943
	10-06-3	Partial Carcass	< 0.0000327	0.0000327	< 0.00000980	0.00000980
	10-06-4	Partial Carcass	< 0.0000344	0.0000344	< 0.00000988	0.00000988
	10-06-5	Partial Carcass	< 0.0000271	0.0000271	< 0.00000815	0.00000815
	10-07-1	Partial Carcass	< 0.0000351	0.0000351	< 0.00000994	0.00000994
	10-07-2	Partial Carcass	< 0.0000344	0.0000344	< 0.00000983	0.00000983
	10-07-3	Partial Carcass	< 0.0000332	0.0000332	< 0.00000976	0.00000976
	10-07-4	Partial Carcass	< 0.0000399	0.0000399	< 0.00000942	0.00000942
	10-07-5	Partial Carcass	< 0.0000304	0.0000304	< 0.00000801	0.00000801
	10-07-6	Partial Carcass	< 0.0000304	0.0000304	< 0.00000777	0.00000777
	4-08-6	Partial Carcass	< 0.0000298	0.0000298	< 0.00000753	0.00000753
	06385810	Water			< 0.0000000649	0.0000000649
PCB# 90/101/113						
	10-06-1	Partial Carcass	0.00160	0.000686	0.000428	0.000183
	10-06-2	Partial Carcass	0.00147	0.000743	0.000373	0.000189
	10-06-3	Partial Carcass	0.00184	0.000653	0.000551	0.000196
	10-06-4	Partial Carcass	0.00296	0.000690	0.000849	0.000198
	10-06-5	Partial Carcass	0.00154	0.000542	0.000463	0.000163
	10-07-1	Partial Carcass	< 0.000702	0.000702	< 0.000199	0.000199
	10-07-2	Partial Carcass	0.00138	0.000690	0.000393	0.000197
	10-07-3	Partial Carcass	0.00674	0.000664	0.00198	0.000195
	10-07-4	Partial Carcass	0.00483	0.000797	0.00114	0.000188
	10-07-5	Partial Carcass	0.00172	0.000608	0.000452	0.000160

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-07-6	Partial Carcass	0.000846	0.000607	0.000216	0.000155
	4-08-6	Partial Carcass	0.00137	0.000598	0.000346	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB# 92						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	0.000421	0.000344	0.000121	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	0.00104	0.000332	0.000304	0.0000976
	10-07-4	Partial Carcass	0.000708	0.000399	0.000167	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 94						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 95						
	10-06-1	Partial Carcass	0.000675	0.000344	0.000180	0.0000917
	10-06-2	Partial Carcass	0.000727	0.000371	0.000185	0.0000943
	10-06-3	Partial Carcass	0.000900	0.000327	0.000270	0.0000980
	10-06-4	Partial Carcass	0.00131	0.000344	0.000375	0.0000988
	10-06-5	Partial Carcass	0.000622	0.000271	0.000187	0.0000815
	10-07-1	Partial Carcass	0.000364	0.000351	0.000103	0.0000994
	10-07-2	Partial Carcass	0.000700	0.000344	0.000200	0.0000983
	10-07-3	Partial Carcass	0.00307	0.000332	0.000900	0.0000976
	10-07-4	Partial Carcass	0.00196	0.000399	0.000461	0.0000942
	10-07-5	Partial Carcass	0.000802	0.000304	0.000211	0.0000801
	10-07-6	Partial Carcass	0.000423	0.000304	0.000108	0.0000777
	4-08-6	Partial Carcass	0.000681	0.000298	0.000172	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 96						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	06385810	Water			< 0.000000649	0.000000649
PCB# 98/100						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649
PCB# 98/102						
	10-06-1	Partial Carcass	< 0.000344	0.000344	< 0.0000917	0.0000917
	10-06-2	Partial Carcass	< 0.000371	0.000371	< 0.0000943	0.0000943
	10-06-3	Partial Carcass	< 0.000327	0.000327	< 0.0000980	0.0000980
	10-06-4	Partial Carcass	< 0.000344	0.000344	< 0.0000988	0.0000988
	10-06-5	Partial Carcass	< 0.000271	0.000271	< 0.0000815	0.0000815
	10-07-1	Partial Carcass	< 0.000351	0.000351	< 0.0000994	0.0000994
	10-07-2	Partial Carcass	< 0.000344	0.000344	< 0.0000983	0.0000983
	10-07-3	Partial Carcass	< 0.000332	0.000332	< 0.0000976	0.0000976
	10-07-4	Partial Carcass	< 0.000399	0.000399	< 0.0000942	0.0000942
	10-07-5	Partial Carcass	< 0.000304	0.000304	< 0.0000801	0.0000801
	10-07-6	Partial Carcass	< 0.000304	0.000304	< 0.0000777	0.0000777
	4-08-6	Partial Carcass	< 0.000298	0.000298	< 0.0000753	0.0000753
	06385810	Water			< 0.000000649	0.000000649

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
PCB-1242						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	0.00202	0.000702	0.000571	0.000199
	10-07-2	Partial Carcass	< 0.000690	0.000690	< 0.000197	0.000197
	10-07-3	Partial Carcass	< 0.000664	0.000664	< 0.000195	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	0.00186	0.000607	0.000475	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			0.00000180	0.00000130
PCB-1248						
	10-06-1	Partial Carcass	< 0.000686	0.000686	< 0.000183	0.000183
	10-06-2	Partial Carcass	< 0.000743	0.000743	< 0.000189	0.000189
	10-06-3	Partial Carcass	< 0.000653	0.000653	< 0.000196	0.000196
	10-06-4	Partial Carcass	< 0.000690	0.000690	< 0.000198	0.000198
	10-06-5	Partial Carcass	< 0.000542	0.000542	< 0.000163	0.000163
	10-07-1	Partial Carcass	0.00403	0.000702	0.00114	0.000199
	10-07-2	Partial Carcass	0.00883	0.000690	0.00252	0.000197
	10-07-3	Partial Carcass	0.0260	0.000664	0.00764	0.000195
	10-07-4	Partial Carcass	< 0.000797	0.000797	< 0.000188	0.000188
	10-07-5	Partial Carcass	< 0.000608	0.000608	< 0.000160	0.000160
	10-07-6	Partial Carcass	0.00372	0.000607	0.000950	0.000155
	4-08-6	Partial Carcass	< 0.000598	0.000598	< 0.000151	0.000151
	06385810	Water			0.00000360	0.00000130
PCB-1254						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-1	Partial Carcass	0.0157	0.000686	0.00420	0.000183
	10-06-2	Partial Carcass	0.0131	0.000743	0.00333	0.000189
	10-06-3	Partial Carcass	0.0162	0.000653	0.00486	0.000196
	10-06-4	Partial Carcass	0.0288	0.000690	0.00828	0.000198
	10-06-5	Partial Carcass	0.0150	0.000542	0.00450	0.000163
	10-07-1	Partial Carcass	0.00907	0.000702	0.00257	0.000199
	10-07-2	Partial Carcass	0.0177	0.000690	0.00505	0.000197
	10-07-3	Partial Carcass	0.0911	0.000664	0.0267	0.000195
	10-07-4	Partial Carcass	0.0651	0.000797	0.0154	0.000188
	10-07-5	Partial Carcass	0.0220	0.000608	0.00580	0.000160
	10-07-6	Partial Carcass	0.0130	0.000607	0.00332	0.000155
	4-08-6	Partial Carcass	0.0125	0.000598	0.00316	0.000151
	06385810	Water			0.0000126	0.00000130
PCB-1260						
	10-06-1	Partial Carcass	0.0236	0.000686	0.00630	0.000183
	10-06-2	Partial Carcass	0.0305	0.000743	0.00777	0.000189
	10-06-3	Partial Carcass	0.0378	0.000653	0.0113	0.000196
	10-06-4	Partial Carcass	0.0673	0.000690	0.0193	0.000198
	10-06-5	Partial Carcass	0.0349	0.000542	0.0105	0.000163
	10-07-1	Partial Carcass	0.00504	0.000702	0.00143	0.000199
	10-07-2	Partial Carcass	0.00294	0.000690	0.000841	0.000197
	10-07-3	Partial Carcass	0.0130	0.000664	0.00382	0.000195
	10-07-4	Partial Carcass	0.0434	0.000797	0.0102	0.000188
	10-07-5	Partial Carcass	0.0220	0.000608	0.00580	0.000160
	10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	4-08-6	Partial Carcass	0.0233	0.000598	0.00588	0.000151
	06385810	Water			< 0.00000130	0.00000130
PCB-TOTAL						
	10-06-1	Partial Carcass	0.0394	0.000686	0.0105	0.000183

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	10-06-2	Partial Carcass	0.0436	0.000743	0.0111	0.000189
	10-06-3	Partial Carcass	0.0540	0.000653	0.0162	0.000196
	10-06-4	Partial Carcass	0.0961	0.000690	0.0276	0.000198
	10-06-5	Partial Carcass	0.0499	0.000542	0.0150	0.000163
	10-07-1	Partial Carcass	0.0202	0.000702	0.00571	0.000199
	10-07-2	Partial Carcass	0.0294	0.000690	0.00841	0.000197
	10-07-3	Partial Carcass	0.130	0.000664	0.0382	0.000195
	10-07-4	Partial Carcass	0.109	0.000797	0.0256	0.000188
	10-07-5	Partial Carcass	0.0441	0.000608	0.0116	0.000160
	10-07-6	Partial Carcass	0.0186	0.000607	0.00475	0.000155
	4-08-6	Partial Carcass	0.0358	0.000598	0.00904	0.000151
	06385810	Water			0.0000180	0.00000130
Pentachlorobenzene						
	*10-06-1	Partial Carcass	0.00976	0.000418	0.00260	0.000111
	*10-06-2	Partial Carcass	0.0102	0.000438	0.00259	0.000111
	*10-06-3	Partial Carcass	0.00910	0.000372	0.00273	0.000111
	*10-06-4	Partial Carcass	0.0109	0.000388	0.00313	0.000111
	*10-06-5	Partial Carcass	0.0126	0.000371	0.00379	0.000111
	*10-07-1	Partial Carcass	0.00880	0.000394	0.00249	0.000111
	*10-07-2	Partial Carcass	0.00764	0.000390	0.00218	0.000111
	*10-07-3	Partial Carcass	0.0107	0.000380	0.00315	0.000111
	*10-07-4	Partial Carcass	0.0121	0.000473	0.00286	0.000111
	*10-07-5	Partial Carcass	0.00944	0.000424	0.00249	0.000111
	*10-07-6	Partial Carcass	0.0118	0.000437	0.00301	0.000111
	*4-08-6	Partial Carcass	0.0119	0.000442	0.00300	0.000111
Total DDT's						
	*10-06-1	Partial Carcass	0.0548	0.00308	0.0146	0.000822
	*10-06-2	Partial Carcass	0.0671	0.00323	0.0171	0.000822
	*10-06-3	Partial Carcass	0.0756	0.00274	0.0227	0.000822

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-06-4	Partial Carcass	0.107	0.00286	0.0307	0.000822
	*10-06-5	Partial Carcass	0.0579	0.00273	0.0174	0.000822
	*10-07-1	Partial Carcass	0.0311	0.00290	0.00880	0.000822
	*10-07-2	Partial Carcass	0.0326	0.00288	0.00932	0.000822
	*10-07-3	Partial Carcass	0.0723	0.00280	0.0212	0.000822
	*10-07-4	Partial Carcass	0.121	0.00349	0.0284	0.000822
	*10-07-5	Partial Carcass	0.0438	0.00312	0.0115	0.000822
	*10-07-6	Partial Carcass	0.0609	0.00322	0.0155	0.000822
	*4-08-6	Partial Carcass	0.0514	0.00326	0.0130	0.000822
acenaphthalene						
	1-08-1	Partial Carcass	0.00212	0.00170	0.000500	0.000402
	1-08-2	Partial Carcass	0.00153	0.00152	0.000400	0.000398
	1-08-3	Partial Carcass	0.00366	0.00146	0.00100	0.000399
	1-08-4	Partial Carcass	0.00501	0.00155	0.00130	0.000401
	1-08-6	Partial Carcass	0.00262	0.00147	0.000700	0.000391
acenaphthene						
	1-08-1	Partial Carcass	< 0.00175	0.00175	< 0.000414	0.000414
	1-08-2	Partial Carcass	< 0.00156	0.00156	< 0.000410	0.000410
	1-08-3	Partial Carcass	0.00256	0.00150	0.000700	0.000411
	1-08-4	Partial Carcass	0.00270	0.00159	0.000700	0.000413
	1-08-6	Partial Carcass	0.00188	0.00151	0.000500	0.000403
alpha BHC						
	*10-06-1	Partial Carcass	< 0.000785	0.000785	< 0.000209	0.000209
	*10-06-2	Partial Carcass	< 0.000823	0.000823	< 0.000209	0.000209
	*10-06-3	Partial Carcass	< 0.000698	0.000698	< 0.000209	0.000209
	*10-06-4	Partial Carcass	< 0.000729	0.000729	< 0.000209	0.000209
	*10-06-5	Partial Carcass	< 0.000696	0.000696	< 0.000209	0.000209
	*10-07-1	Partial Carcass	< 0.000739	0.000739	< 0.000209	0.000209
	*10-07-2	Partial Carcass	< 0.000733	0.000733	< 0.000209	0.000209

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-3	Partial Carcass	< 0.000713	0.000713	< 0.000209	0.000209
	*10-07-4	Partial Carcass	< 0.000888	0.000888	< 0.000209	0.000209
	*10-07-5	Partial Carcass	< 0.000795	0.000795	< 0.000209	0.000209
	*10-07-6	Partial Carcass	< 0.000820	0.000820	< 0.000209	0.000209
	*4-08-6	Partial Carcass	< 0.000829	0.000829	< 0.000209	0.000209
alpha chlordane						
	*10-06-1	Partial Carcass	0.00642	0.000709	0.00171	0.000189
	*10-06-2	Partial Carcass	0.00998	0.000743	0.00254	0.000189
	*10-06-3	Partial Carcass	0.0189	0.000630	0.00566	0.000189
	*10-06-4	Partial Carcass	0.0226	0.000658	0.00649	0.000189
	*10-06-5	Partial Carcass	0.0105	0.000628	0.00317	0.000189
	*10-07-1	Partial Carcass	0.000951	0.000667	0.000269	0.000189
	*10-07-2	Partial Carcass	0.00477	0.000661	0.00136	0.000189
	*10-07-3	Partial Carcass	0.0320	0.000644	0.00938	0.000189
	*10-07-4	Partial Carcass	0.0286	0.000801	0.00675	0.000189
	*10-07-5	Partial Carcass	0.0123	0.000718	0.00323	0.000189
	*10-07-6	Partial Carcass	0.00164	0.000740	0.000419	0.000189
	*4-08-6	Partial Carcass	0.00862	0.000748	0.00218	0.000189
anthracene						
	1-08-1	Partial Carcass	0.00127	0.00103	0.000300	0.000244
	1-08-2	Partial Carcass	0.00114	0.000921	0.000300	0.000241
	1-08-3	Partial Carcass	0.00146	0.000883	0.000400	0.000242
	1-08-4	Partial Carcass	0.00193	0.000937	0.000500	0.000243
	1-08-6	Partial Carcass	0.00112	0.000889	0.000300	0.000237
benzo(a)pyrene						
	1-08-1	Partial Carcass	< 0.00191	0.00191	< 0.000451	0.000451
	1-08-2	Partial Carcass	< 0.00170	0.00170	< 0.000446	0.000446
	1-08-3	Partial Carcass	< 0.00163	0.00163	< 0.000447	0.000447
	1-08-4	Partial Carcass	< 0.00173	0.00173	< 0.000450	0.000450

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-6	Partial Carcass	< 0.00164	0.00164	< 0.000438	0.000438
benzo(b)fluoranthene						
	1-08-1	Partial Carcass	0.00338	0.00139	0.000800	0.000329
	1-08-2	Partial Carcass	0.00153	0.00124	0.000400	0.000326
	1-08-3	Partial Carcass	0.00256	0.00119	0.000700	0.000326
	1-08-4	Partial Carcass	0.00501	0.00127	0.00130	0.000328
	1-08-6	Partial Carcass	< 0.00120	0.00120	< 0.000320	0.000320
benzo(e)pyrene						
	1-08-1	Partial Carcass	< 0.00201	0.00201	< 0.000475	0.000475
	1-08-2	Partial Carcass	< 0.00180	0.00180	< 0.000470	0.000470
	1-08-3	Partial Carcass	< 0.00172	0.00172	< 0.000471	0.000471
	1-08-4	Partial Carcass	0.00231	0.00183	0.000600	0.000474
	1-08-6	Partial Carcass	< 0.00173	0.00173	< 0.000462	0.000462
benzo(g,h,i)perylene						
	1-08-1	Partial Carcass	< 0.00227	0.00227	< 0.000536	0.000536
	1-08-2	Partial Carcass	< 0.00202	0.00202	< 0.000530	0.000530
	1-08-3	Partial Carcass	< 0.00194	0.00194	< 0.000532	0.000532
	1-08-4	Partial Carcass	< 0.00206	0.00206	< 0.000535	0.000535
	1-08-6	Partial Carcass	< 0.00196	0.00196	< 0.000521	0.000521
benzo(k)fluoranthene						
	1-08-1	Partial Carcass	< 0.00175	0.00175	< 0.000414	0.000414
	1-08-2	Partial Carcass	< 0.00156	0.00156	< 0.000410	0.000410
	1-08-3	Partial Carcass	< 0.00150	0.00150	< 0.000411	0.000411
	1-08-4	Partial Carcass	< 0.00159	0.00159	< 0.000413	0.000413
	1-08-6	Partial Carcass	< 0.00151	0.00151	< 0.000403	0.000403
beta BHC						
	*10-06-1	Partial Carcass	< 0.000841	0.000841	< 0.000224	0.000224
	*10-06-2	Partial Carcass	< 0.000882	0.000882	< 0.000224	0.000224
	*10-06-3	Partial Carcass	< 0.000748	0.000748	< 0.000224	0.000224

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-06-4	Partial Carcass	< 0.000781	0.000781	< 0.000224	0.000224
	*10-06-5	Partial Carcass	< 0.000746	0.000746	< 0.000224	0.000224
	*10-07-1	Partial Carcass	< 0.000792	0.000792	< 0.000224	0.000224
	*10-07-2	Partial Carcass	< 0.000785	0.000785	< 0.000224	0.000224
	*10-07-3	Partial Carcass	< 0.000764	0.000764	< 0.000224	0.000224
	*10-07-4	Partial Carcass	< 0.000951	0.000951	< 0.000224	0.000224
	*10-07-5	Partial Carcass	< 0.000852	0.000852	< 0.000224	0.000224
	*10-07-6	Partial Carcass	< 0.000879	0.000879	< 0.000224	0.000224
	*4-08-6	Partial Carcass	< 0.000888	0.000888	< 0.000224	0.000224
biphenyl						
	1-08-1	Partial Carcass	0.00465	0.00314	0.00110	0.000743
	1-08-2	Partial Carcass	0.00687	0.00281	0.00180	0.000735
	1-08-3	Partial Carcass	0.0205	0.00269	0.00560	0.000737
	1-08-4	Partial Carcass	0.0108	0.00286	0.00280	0.000741
	1-08-6	Partial Carcass	0.00637	0.00271	0.00170	0.000723
chlorpyrifos						
	*10-06-1	Partial Carcass	< 0.00104	0.00104	< 0.000277	0.000277
	*10-06-2	Partial Carcass	< 0.00109	0.00109	< 0.000277	0.000277
	*10-06-3	Partial Carcass	< 0.000923	0.000923	< 0.000277	0.000277
	*10-06-4	Partial Carcass	< 0.000965	0.000965	< 0.000277	0.000277
	*10-06-5	Partial Carcass	< 0.000921	0.000921	< 0.000277	0.000277
	*10-07-1	Partial Carcass	< 0.000978	0.000978	< 0.000277	0.000277
	*10-07-2	Partial Carcass	< 0.000969	0.000969	< 0.000277	0.000277
	*10-07-3	Partial Carcass	< 0.000943	0.000943	< 0.000277	0.000277
	*10-07-4	Partial Carcass	< 0.00118	0.00118	< 0.000277	0.000277
	*10-07-5	Partial Carcass	< 0.00105	0.00105	< 0.000277	0.000277
	*10-07-6	Partial Carcass	< 0.00108	0.00108	< 0.000277	0.000277
	*4-08-6	Partial Carcass	< 0.00110	0.00110	< 0.000277	0.000277
chrysene						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-1	Partial Carcass	0.00212	0.00201	0.000500	0.000475
	1-08-2	Partial Carcass	0.00191	0.00180	0.000500	0.000470
	1-08-3	Partial Carcass	0.00183	0.00172	0.000500	0.000471
	1-08-4	Partial Carcass	0.00347	0.00183	0.000900	0.000474
	1-08-6	Partial Carcass	< 0.00173	0.00173	< 0.000462	0.000462
cis-nonachlor						
	*10-06-1	Partial Carcass	0.00365	0.000526	0.000973	0.000140
	*10-06-2	Partial Carcass	0.00455	0.000551	0.00116	0.000140
	*10-06-3	Partial Carcass	0.00736	0.000467	0.00221	0.000140
	*10-06-4	Partial Carcass	0.00965	0.000488	0.00277	0.000140
	*10-06-5	Partial Carcass	0.00529	0.000466	0.00159	0.000140
	*10-07-1	Partial Carcass	0.000555	0.000495	0.000157	0.000140
	*10-07-2	Partial Carcass	0.00230	0.000491	0.000656	0.000140
	*10-07-3	Partial Carcass	0.0117	0.000478	0.00344	0.000140
	*10-07-4	Partial Carcass	0.0143	0.000595	0.00337	0.000140
	*10-07-5	Partial Carcass	0.00661	0.000533	0.00174	0.000140
	*10-07-6	Partial Carcass	0.00101	0.000549	0.000259	0.000140
	*4-08-6	Partial Carcass	0.00387	0.000555	0.000977	0.000140
delta BHC						
	*10-06-1	Partial Carcass	< 0.000409	0.000409	< 0.000109	0.000109
	*10-06-2	Partial Carcass	< 0.000429	0.000429	< 0.000109	0.000109
	*10-06-3	Partial Carcass	< 0.000363	0.000363	< 0.000109	0.000109
	*10-06-4	Partial Carcass	< 0.000380	0.000380	< 0.000109	0.000109
	*10-06-5	Partial Carcass	< 0.000362	0.000362	< 0.000109	0.000109
	*10-07-1	Partial Carcass	< 0.000385	0.000385	< 0.000109	0.000109
	*10-07-2	Partial Carcass	< 0.000382	0.000382	< 0.000109	0.000109
	*10-07-3	Partial Carcass	< 0.000371	0.000371	< 0.000109	0.000109
	*10-07-4	Partial Carcass	< 0.000462	0.000462	< 0.000109	0.000109
	*10-07-5	Partial Carcass	< 0.000414	0.000414	< 0.000109	0.000109

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-6	Partial Carcass	< 0.000427	0.000427	< 0.000109	0.000109
	*4-08-6	Partial Carcass	< 0.000432	0.000432	< 0.000109	0.000109
dibenzothiophene						
	1-08-1	Partial Carcass	< 0.00144	0.00144	< 0.000341	0.000341
	1-08-2	Partial Carcass	0.00153	0.00129	0.000400	0.000338
	1-08-3	Partial Carcass	0.00146	0.00124	0.000400	0.000338
	1-08-4	Partial Carcass	0.00193	0.00131	0.000500	0.000340
	1-08-6	Partial Carcass	< 0.00124	0.00124	< 0.000332	0.000332
dieldrin						
	*10-06-1	Partial Carcass	0.00273	0.000707	0.000727	0.000189
	*10-06-2	Partial Carcass	0.00336	0.000741	0.000855	0.000189
	*10-06-3	Partial Carcass	0.0138	0.000629	0.00415	0.000189
	*10-06-4	Partial Carcass	0.0133	0.000657	0.00382	0.000189
	*10-06-5	Partial Carcass	0.00926	0.000627	0.00278	0.000189
	*10-07-1	Partial Carcass	< 0.000666	0.000666	< 0.000189	0.000189
	*10-07-2	Partial Carcass	0.00155	0.000660	0.000442	0.000189
	*10-07-3	Partial Carcass	0.00970	0.000642	0.00285	0.000189
	*10-07-4	Partial Carcass	0.0104	0.000800	0.00246	0.000189
	*10-07-5	Partial Carcass	0.00519	0.000717	0.00137	0.000189
	*10-07-6	Partial Carcass	0.00164	0.000739	0.000419	0.000189
	*4-08-6	Partial Carcass	0.00537	0.000747	0.00136	0.000189
endosulfan I						
	*10-06-1	Partial Carcass	0.00597	0.000569	0.00159	0.000152
	*10-06-2	Partial Carcass	0.00652	0.000596	0.00166	0.000152
	*10-06-3	Partial Carcass	0.00831	0.000506	0.00249	0.000152
	*10-06-4	Partial Carcass	0.0120	0.000528	0.00345	0.000152
	*10-06-5	Partial Carcass	0.00782	0.000504	0.00235	0.000152
	*10-07-1	Partial Carcass	0.00123	0.000536	0.000347	0.000152
	*10-07-2	Partial Carcass	0.00225	0.000531	0.000642	0.000152

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-3	Partial Carcass	0.0142	0.000517	0.00418	0.000152
	*10-07-4	Partial Carcass	0.0172	0.000643	0.00405	0.000152
	*10-07-5	Partial Carcass	0.00815	0.000576	0.00214	0.000152
	*10-07-6	Partial Carcass	0.00226	0.000594	0.000577	0.000152
	*4-08-6	Partial Carcass	0.00502	0.000601	0.00127	0.000152
endosulfan II						
	*10-06-1	Partial Carcass	0.000879	0.000557	0.000234	0.000148
	*10-06-2	Partial Carcass	< 0.000584	0.000584	< 0.000148	0.000148
	*10-06-3	Partial Carcass	< 0.000495	0.000495	< 0.000148	0.000148
	*10-06-4	Partial Carcass	< 0.000517	0.000517	< 0.000148	0.000148
	*10-06-5	Partial Carcass	0.00216	0.000494	0.000648	0.000148
	*10-07-1	Partial Carcass	< 0.000524	0.000524	< 0.000148	0.000148
	*10-07-2	Partial Carcass	< 0.000520	0.000520	< 0.000148	0.000148
	*10-07-3	Partial Carcass	0.00106	0.000506	0.000312	0.000148
	*10-07-4	Partial Carcass	< 0.000630	0.000630	< 0.000148	0.000148
	*10-07-5	Partial Carcass	< 0.000564	0.000564	< 0.000148	0.000148
	*10-07-6	Partial Carcass	< 0.000582	0.000582	< 0.000148	0.000148
	*4-08-6	Partial Carcass	< 0.000588	0.000588	< 0.000148	0.000148
endosulfan sulfate						
	*10-06-1	Partial Carcass	0.00528	0.000579	0.00141	0.000154
	*10-06-2	Partial Carcass	0.00944	0.000607	0.00240	0.000154
	*10-06-3	Partial Carcass	0.0145	0.000515	0.00434	0.000154
	*10-06-4	Partial Carcass	0.0160	0.000538	0.00460	0.000154
	*10-06-5	Partial Carcass	0.00945	0.000513	0.00284	0.000154
	*10-07-1	Partial Carcass	0.00150	0.000545	0.000427	0.000154
	*10-07-2	Partial Carcass	0.00384	0.000541	0.00110	0.000154
	*10-07-3	Partial Carcass	0.0110	0.000526	0.00322	0.000154
	*10-07-4	Partial Carcass	0.0183	0.000655	0.00432	0.000154
	*10-07-5	Partial Carcass	0.00753	0.000587	0.00198	0.000154

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-6	Partial Carcass	0.00188	0.000605	0.000481	0.000154
	*4-08-6	Partial Carcass	0.00604	0.000612	0.00152	0.000154
endrin						
	*10-06-1	Partial Carcass	< 0.000759	0.000759	< 0.000202	0.000202
	*10-06-2	Partial Carcass	< 0.000795	0.000795	< 0.000202	0.000202
	*10-06-3	Partial Carcass	< 0.000675	0.000675	< 0.000202	0.000202
	*10-06-4	Partial Carcass	< 0.000705	0.000705	< 0.000202	0.000202
	*10-06-5	Partial Carcass	< 0.000673	0.000673	< 0.000202	0.000202
	*10-07-1	Partial Carcass	< 0.000714	0.000714	< 0.000202	0.000202
	*10-07-2	Partial Carcass	< 0.000708	0.000708	< 0.000202	0.000202
	*10-07-3	Partial Carcass	< 0.000689	0.000689	< 0.000202	0.000202
	*10-07-4	Partial Carcass	< 0.000858	0.000858	< 0.000202	0.000202
	*10-07-5	Partial Carcass	< 0.000769	0.000769	< 0.000202	0.000202
	*10-07-6	Partial Carcass	< 0.000793	0.000793	< 0.000202	0.000202
	*4-08-6	Partial Carcass	< 0.000801	0.000801	< 0.000202	0.000202
fluoranthene						
	1-08-1	Partial Carcass	0.00423	0.00129	0.00100	0.000304
	1-08-2	Partial Carcass	0.00687	0.00115	0.00180	0.000301
	1-08-3	Partial Carcass	0.00439	0.00110	0.00120	0.000302
	1-08-4	Partial Carcass	0.0108	0.00117	0.00280	0.000304
	1-08-6	Partial Carcass	0.00188	0.00111	0.000500	0.000296
fluorene						
	1-08-1	Partial Carcass	0.00508	0.00175	0.00120	0.000414
	1-08-2	Partial Carcass	0.00573	0.00156	0.00150	0.000410
	1-08-3	Partial Carcass	0.00804	0.00150	0.00220	0.000411
	1-08-4	Partial Carcass	0.00733	0.00159	0.00190	0.000413
	1-08-6	Partial Carcass	0.00525	0.00151	0.00140	0.000403
gamma BHC						
	*10-06-1	Partial Carcass	< 0.000351	0.000351	< 0.0000940	0.0000940

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-06-2	Partial Carcass	< 0.000368	0.000368	< 0.0000940	0.0000940
	*10-06-3	Partial Carcass	< 0.000312	0.000312	< 0.0000940	0.0000940
	*10-06-4	Partial Carcass	< 0.000326	0.000326	< 0.0000940	0.0000940
	*10-06-5	Partial Carcass	< 0.000311	0.000311	< 0.0000940	0.0000940
	*10-07-1	Partial Carcass	< 0.000330	0.000330	< 0.0000940	0.0000940
	*10-07-2	Partial Carcass	< 0.000327	0.000327	< 0.0000940	0.0000940
	*10-07-3	Partial Carcass	0.00900	0.000318	0.00264	0.0000940
	*10-07-4	Partial Carcass	0.000497	0.000397	0.000117	0.0000940
	*10-07-5	Partial Carcass	< 0.000355	0.000355	< 0.0000940	0.0000940
	*10-07-6	Partial Carcass	< 0.000366	0.000366	< 0.0000940	0.0000940
	*4-08-6	Partial Carcass	< 0.000370	0.000370	< 0.0000940	0.0000940
gamma chlordane						
	*10-06-1	Partial Carcass	0.00321	0.000581	0.000856	0.000155
	*10-06-2	Partial Carcass	0.00484	0.000609	0.00123	0.000155
	*10-06-3	Partial Carcass	0.0107	0.000516	0.00322	0.000155
	*10-06-4	Partial Carcass	0.0114	0.000539	0.00328	0.000155
	*10-06-5	Partial Carcass	0.00500	0.000515	0.00150	0.000155
	*10-07-1	Partial Carcass	< 0.000547	0.000547	< 0.000155	0.000155
	*10-07-2	Partial Carcass	0.00331	0.000542	0.000946	0.000155
	*10-07-3	Partial Carcass	0.0186	0.000528	0.00546	0.000155
	*10-07-4	Partial Carcass	0.0138	0.000657	0.00324	0.000155
	*10-07-5	Partial Carcass	0.00601	0.000588	0.00158	0.000155
	*10-07-6	Partial Carcass	0.000676	0.000607	0.000173	0.000155
	*4-08-6	Partial Carcass	0.00449	0.000613	0.00113	0.000155
heptachlor epoxide						
	*10-06-1	Partial Carcass	0.000703	0.000598	0.000188	0.000160
	*10-06-2	Partial Carcass	< 0.000627	0.000627	< 0.000160	0.000160
	*10-06-3	Partial Carcass	0.00102	0.000532	0.000306	0.000160
	*10-06-4	Partial Carcass	0.00150	0.000556	0.000429	0.000160

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-06-5	Partial Carcass	0.00108	0.000531	0.000324	0.000160
	*10-07-1	Partial Carcass	< 0.000563	0.000563	< 0.000160	0.000160
	*10-07-2	Partial Carcass	< 0.000559	0.000559	< 0.000160	0.000160
	*10-07-3	Partial Carcass	0.00106	0.000544	0.000312	0.000160
	*10-07-4	Partial Carcass	< 0.000677	0.000677	< 0.000160	0.000160
	*10-07-5	Partial Carcass	0.000941	0.000606	0.000248	0.000160
	*10-07-6	Partial Carcass	< 0.000625	0.000625	< 0.000160	0.000160
	*4-08-6	Partial Carcass	< 0.000632	0.000632	< 0.000160	0.000160
indeno(1,2,3-cd)pyrene						
	1-08-1	Partial Carcass	< 0.00294	0.00294	< 0.000694	0.000694
	1-08-2	Partial Carcass	< 0.00262	0.00262	< 0.000687	0.000687
	1-08-3	Partial Carcass	< 0.00252	0.00252	< 0.000689	0.000689
	1-08-4	Partial Carcass	< 0.00267	0.00267	< 0.000693	0.000693
	1-08-6	Partial Carcass	< 0.00253	0.00253	< 0.000675	0.000675
mirex						
	*10-06-1	Partial Carcass	< 0.000462	0.000462	< 0.000123	0.000123
	*10-06-2	Partial Carcass	< 0.000485	0.000485	< 0.000123	0.000123
	*10-06-3	Partial Carcass	< 0.000411	0.000411	< 0.000123	0.000123
	*10-06-4	Partial Carcass	0.000453	0.000430	0.000130	0.000123
	*10-06-5	Partial Carcass	< 0.000410	0.000410	< 0.000123	0.000123
	*10-07-1	Partial Carcass	< 0.000435	0.000435	< 0.000123	0.000123
	*10-07-2	Partial Carcass	< 0.000432	0.000432	< 0.000123	0.000123
	*10-07-3	Partial Carcass	< 0.000420	0.000420	< 0.000123	0.000123
	*10-07-4	Partial Carcass	< 0.000523	0.000523	< 0.000123	0.000123
	*10-07-5	Partial Carcass	< 0.000469	0.000469	< 0.000123	0.000123
	*10-07-6	Partial Carcass	< 0.000483	0.000483	< 0.000123	0.000123
	*4-08-6	Partial Carcass	< 0.000488	0.000488	< 0.000123	0.000123
n-decane						
	1-08-1	Partial Carcass	0.841	0.0548	0.199	0.0130

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-2	Partial Carcass	0.183	0.0490	0.0479	0.0128
	1-08-3	Partial Carcass	0.241	0.0470	0.0660	0.0128
	1-08-4	Partial Carcass	0.268	0.0499	0.0695	0.0129
	1-08-6	Partial Carcass	0.338	0.0473	0.0901	0.0126
n-docosane						
	1-08-1	Partial Carcass	< 0.0693	0.0693	< 0.0164	0.0164
	1-08-2	Partial Carcass	< 0.0619	0.0619	< 0.0162	0.0162
	1-08-3	Partial Carcass	3.25	0.0594	0.889	0.0163
	1-08-4	Partial Carcass	< 0.0631	0.0631	< 0.0164	0.0164
	1-08-6	Partial Carcass	< 0.0598	0.0598	< 0.0159	0.0159
n-dodecane						
	1-08-1	Partial Carcass	0.0966	0.0401	0.0228	0.00948
	1-08-2	Partial Carcass	0.125	0.0358	0.0327	0.00939
	1-08-3	Partial Carcass	0.208	0.0344	0.0569	0.00940
	1-08-4	Partial Carcass	0.203	0.0365	0.0526	0.00946
	1-08-6	Partial Carcass	< 0.0346	0.0346	< 0.00922	0.00922
n-dotriacontane						
	1-08-1	Partial Carcass	< 0.0631	0.0631	< 0.0149	0.0149
	1-08-2	Partial Carcass	< 0.0564	0.0564	< 0.0148	0.0148
	1-08-3	Partial Carcass	< 0.0541	0.0541	< 0.0148	0.0148
	1-08-4	Partial Carcass	< 0.0574	0.0574	< 0.0149	0.0149
	1-08-6	Partial Carcass	< 0.0544	0.0544	< 0.0145	0.0145
n-eicosane						
	*1-08-1	Partial Carcass	5.13	0.0706	1.21	0.0167
	1-08-2	Partial Carcass	< 0.0631	0.0631	< 0.0165	0.0165
	1-08-3	Partial Carcass	0.675	0.0605	0.185	0.0166
	1-08-4	Partial Carcass	1.26	0.0642	0.327	0.0167
	1-08-6	Partial Carcass	< 0.0609	0.0609	< 0.0162	0.0162
n-heneicosane						

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-1	Partial Carcass	0.297	0.0742	0.0702	0.0175
	1-08-2	Partial Carcass	< 0.0663	0.0663	< 0.0174	0.0174
	1-08-3	Partial Carcass	5.28	0.0636	1.45	0.0174
	1-08-4	Partial Carcass	0.397	0.0675	0.103	0.0175
	1-08-6	Partial Carcass	< 0.0640	0.0640	< 0.0171	0.0171
n-hentriacontane						
	1-08-1	Partial Carcass	0.468	0.0645	0.111	0.0152
	1-08-2	Partial Carcass	< 0.0576	0.0576	< 0.0151	0.0151
	1-08-3	Partial Carcass	21.4	0.0553	5.85	0.0151
	1-08-4	Partial Carcass	< 0.0587	0.0587	< 0.0152	0.0152
	1-08-6	Partial Carcass	< 0.0556	0.0556	< 0.0148	0.0148
n-heptacosane						
	1-08-1	Partial Carcass	1.24	0.0615	0.294	0.0145
	1-08-2	Partial Carcass	0.388	0.0550	0.102	0.0144
	1-08-3	Partial Carcass	1.69	0.0527	0.463	0.0144
	*1-08-4	Partial Carcass	4.65	0.0560	1.21	0.0145
	1-08-6	Partial Carcass	0.494	0.0530	0.132	0.0141
n-heptadecane						
	1-08-1	Partial Carcass	11.5	0.0564	2.73	0.0133
	*1-08-2	Partial Carcass	24.8	0.0504	6.49	0.0132
	*1-08-3	Partial Carcass	28.0	0.0484	7.66	0.0132
	1-08-4	Partial Carcass	15.9	0.0514	4.12	0.0133
	*1-08-6	Partial Carcass	66.1	0.0487	17.6	0.0130
n-hexacosane						
	1-08-1	Partial Carcass	0.914	0.0630	0.216	0.0149
	1-08-2	Partial Carcass	0.346	0.0563	0.0907	0.0148
	1-08-3	Partial Carcass	1.14	0.0540	0.312	0.0148
	1-08-4	Partial Carcass	2.10	0.0573	0.545	0.0149
	1-08-6	Partial Carcass	2.13	0.0544	0.567	0.0145

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
n-hexadecane						
	1-08-1	Partial Carcass	1.15	0.0694	0.271	0.0164
	1-08-2	Partial Carcass	1.29	0.0620	0.337	0.0162
	1-08-3	Partial Carcass	2.85	0.0595	0.780	0.0163
	1-08-4	Partial Carcass	3.27	0.0631	0.849	0.0164
	1-08-6	Partial Carcass	1.94	0.0598	0.517	0.0160
n-nonacosane						
	1-08-1	Partial Carcass	0.248	0.0564	0.0587	0.0133
	1-08-2	Partial Carcass	< 0.0504	0.0504	< 0.0132	0.0132
	*1-08-3	Partial Carcass	25.2	0.0484	6.88	0.0132
	*1-08-4	Partial Carcass	31.9	0.0514	8.26	0.0133
	1-08-6	Partial Carcass	< 0.0487	0.0487	< 0.0130	0.0130
n-nonadecane						
	1-08-1	Partial Carcass	0.925	0.0578	0.219	0.0137
	1-08-2	Partial Carcass	1.60	0.0517	0.419	0.0135
	*1-08-3	Partial Carcass	19.4	0.0495	5.32	0.0136
	1-08-4	Partial Carcass	0.668	0.0526	0.173	0.0136
	1-08-6	Partial Carcass	3.09	0.0499	0.825	0.0133
n-octacosane						
	1-08-1	Partial Carcass	0.223	0.0610	0.0528	0.0144
	1-08-2	Partial Carcass	< 0.0545	0.0545	< 0.0143	0.0143
	1-08-3	Partial Carcass	0.689	0.0523	0.188	0.0143
	1-08-4	Partial Carcass	< 0.0555	0.0555	< 0.0144	0.0144
	1-08-6	Partial Carcass	< 0.0526	0.0526	< 0.0140	0.0140
n-octadecane						
	*1-08-1	Partial Carcass	11.6	0.0681	2.75	0.0161
	*1-08-2	Partial Carcass	12.8	0.0608	3.36	0.0159
	*1-08-3	Partial Carcass	25.3	0.0584	6.92	0.0160
	*1-08-4	Partial Carcass	18.5	0.0620	4.79	0.0161

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*1-08-6	Partial Carcass	17.5	0.0587	4.66	0.0157
n-pentacosane						
	1-08-1	Partial Carcass	5.20	0.0712	1.23	0.0168
	1-08-2	Partial Carcass	1.25	0.0636	0.327	0.0167
	1-08-3	Partial Carcass	3.09	0.0610	0.846	0.0167
	1-08-4	Partial Carcass	1.67	0.0648	0.434	0.0168
	1-08-6	Partial Carcass	1.32	0.0614	0.353	0.0164
n-pentadecane						
	*1-08-1	Partial Carcass	13.3	0.0878	3.15	0.0208
	*1-08-2	Partial Carcass	21.4	0.0785	5.61	0.0205
	*1-08-3	Partial Carcass	40.9	0.0753	11.2	0.0206
	*1-08-4	Partial Carcass	25.8	0.0799	6.69	0.0207
	*1-08-6	Partial Carcass	10.8	0.0757	2.87	0.0202
n-tetracosane						
	1-08-1	Partial Carcass	2.38	0.0560	0.562	0.0132
	1-08-2	Partial Carcass	0.807	0.0500	0.211	0.0131
	1-08-3	Partial Carcass	0.245	0.0480	0.0671	0.0131
	1-08-4	Partial Carcass	0.348	0.0510	0.0902	0.0132
	1-08-6	Partial Carcass	0.502	0.0483	0.134	0.0129
n-tetradecane						
	1-08-1	Partial Carcass	0.430	0.0696	0.102	0.0164
	1-08-2	Partial Carcass	0.506	0.0622	0.132	0.0163
	1-08-3	Partial Carcass	1.25	0.0596	0.343	0.0163
	1-08-4	Partial Carcass	0.334	0.0633	0.0867	0.0164
	1-08-6	Partial Carcass	0.512	0.0600	0.137	0.0160
n-tetratriacontane						
	1-08-1	Partial Carcass	< 0.0618	0.0618	< 0.0146	0.0146
	1-08-2	Partial Carcass	< 0.0552	0.0552	< 0.0145	0.0145
	1-08-3	Partial Carcass	< 0.0529	0.0529	< 0.0145	0.0145

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-4	Partial Carcass	< 0.0562	0.0562	< 0.0146	0.0146
	1-08-6	Partial Carcass	< 0.0533	0.0533	< 0.0142	0.0142
n-triacontane						
	1-08-1	Partial Carcass	0.252	0.0598	0.0596	0.0141
	1-08-2	Partial Carcass	< 0.0535	0.0535	< 0.0140	0.0140
	1-08-3	Partial Carcass	0.989	0.0513	0.270	0.0140
	1-08-4	Partial Carcass	0.986	0.0544	0.256	0.0141
	1-08-6	Partial Carcass	< 0.0516	0.0516	< 0.0138	0.0138
n-tricosane						
	*1-08-1	Partial Carcass	46.1	0.0674	10.9	0.0159
	*1-08-2	Partial Carcass	24.5	0.0602	6.42	0.0158
	*1-08-3	Partial Carcass	11.2	0.0577	3.07	0.0158
	1-08-4	Partial Carcass	< 0.0613	0.0613	< 0.0159	0.0159
	*1-08-6	Partial Carcass	14.4	0.0581	3.84	0.0155
n-tridecane						
	1-08-1	Partial Carcass	0.713	0.0695	0.168	0.0164
	1-08-2	Partial Carcass	1.27	0.0621	0.333	0.0163
	1-08-3	Partial Carcass	3.33	0.0596	0.912	0.0163
	1-08-4	Partial Carcass	2.23	0.0632	0.577	0.0164
	1-08-6	Partial Carcass	4.09	0.0599	1.09	0.0160
n-tritriacontane						
	1-08-1	Partial Carcass	< 0.0537	0.0537	< 0.0127	0.0127
	1-08-2	Partial Carcass	< 0.0480	0.0480	< 0.0126	0.0126
	1-08-3	Partial Carcass	< 0.0460	0.0460	< 0.0126	0.0126
	1-08-4	Partial Carcass	< 0.0488	0.0488	< 0.0127	0.0127
	1-08-6	Partial Carcass	< 0.0463	0.0463	< 0.0123	0.0123
n-undecane						
	*1-08-1	Partial Carcass	78.4	0.0606	18.5	0.0143
	1-08-2	Partial Carcass	0.157	0.0541	0.0410	0.0142

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-3	Partial Carcass	0.265	0.0519	0.0725	0.0142
	1-08-4	Partial Carcass	0.360	0.0551	0.0933	0.0143
	1-08-6	Partial Carcass	0.341	0.0523	0.0908	0.0139
naphthalene						
	1-08-1	Partial Carcass	0.0550	0.00422	0.0130	0.000999
	1-08-2	Partial Carcass	0.0531	0.00378	0.0139	0.000989
	1-08-3	Partial Carcass	0.0541	0.00362	0.0148	0.000991
	1-08-4	Partial Carcass	0.0594	0.00384	0.0154	0.000997
	1-08-6	Partial Carcass	0.0649	0.00364	0.0173	0.000972
o,p'-DDD						
	*10-06-1	Partial Carcass	0.00220	0.000773	0.000586	0.000206
	*10-06-2	Partial Carcass	0.00193	0.000810	0.000490	0.000206
	*10-06-3	Partial Carcass	0.00191	0.000687	0.000574	0.000206
	*10-06-4	Partial Carcass	0.00290	0.000718	0.000832	0.000206
	*10-06-5	Partial Carcass	0.00220	0.000685	0.000663	0.000206
	*10-07-1	Partial Carcass	< 0.000728	0.000728	< 0.000206	0.000206
	*10-07-2	Partial Carcass	< 0.000721	0.000721	< 0.000206	0.000206
	*10-07-3	Partial Carcass	0.00295	0.000702	0.000865	0.000206
	*10-07-4	Partial Carcass	0.00452	0.000874	0.00106	0.000206
	*10-07-5	Partial Carcass	0.00152	0.000783	0.000399	0.000206
	*10-07-6	Partial Carcass	< 0.000807	0.000807	< 0.000206	0.000206
	*4-08-6	Partial Carcass	0.00119	0.000816	0.000300	0.000206
o,p'-DDE						
	*10-06-1	Partial Carcass	0.00154	0.000445	0.000410	0.000119
	*10-06-2	Partial Carcass	0.00183	0.000466	0.000465	0.000119
	*10-06-3	Partial Carcass	0.00140	0.000395	0.000421	0.000119
	*10-06-4	Partial Carcass	0.00258	0.000413	0.000741	0.000119
	*10-06-5	Partial Carcass	0.00294	0.000394	0.000884	0.000119
	*10-07-1	Partial Carcass	< 0.000419	0.000419	< 0.000119	0.000119

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-2	Partial Carcass	0.000574	0.000415	0.000164	0.000119
	*10-07-3	Partial Carcass	0.00135	0.000404	0.000396	0.000119
	*10-07-4	Partial Carcass	0.00223	0.000503	0.000527	0.000119
	*10-07-5	Partial Carcass	0.00142	0.000451	0.000375	0.000119
	*10-07-6	Partial Carcass	0.00121	0.000465	0.000308	0.000119
	*4-08-6	Partial Carcass	0.000826	0.000470	0.000209	0.000119
o,p'-DDT						
	*10-06-1	Partial Carcass	< 0.000582	0.000582	< 0.000155	0.000155
	*10-06-2	Partial Carcass	< 0.000610	0.000610	< 0.000155	0.000155
	*10-06-3	Partial Carcass	< 0.000517	0.000517	< 0.000155	0.000155
	*10-06-4	Partial Carcass	< 0.000540	0.000540	< 0.000155	0.000155
	*10-06-5	Partial Carcass	< 0.000516	0.000516	< 0.000155	0.000155
	*10-07-1	Partial Carcass	< 0.000547	0.000547	< 0.000155	0.000155
	*10-07-2	Partial Carcass	< 0.000543	0.000543	< 0.000155	0.000155
	*10-07-3	Partial Carcass	< 0.000528	0.000528	< 0.000155	0.000155
	*10-07-4	Partial Carcass	< 0.000658	0.000658	< 0.000155	0.000155
	*10-07-5	Partial Carcass	< 0.000589	0.000589	< 0.000155	0.000155
	*10-07-6	Partial Carcass	< 0.000607	0.000607	< 0.000155	0.000155
	*4-08-6	Partial Carcass	< 0.000614	0.000614	< 0.000155	0.000155
oxychlordane						
	*10-06-1	Partial Carcass	0.00114	0.000455	0.000305	0.000121
	*10-06-2	Partial Carcass	0.000988	0.000478	0.000251	0.000121
	*10-06-3	Partial Carcass	0.00111	0.000405	0.000332	0.000121
	*10-06-4	Partial Carcass	0.00181	0.000423	0.000520	0.000121
	*10-06-5	Partial Carcass	0.00108	0.000404	0.000324	0.000121
	*10-07-1	Partial Carcass	0.000713	0.000429	0.000202	0.000121
	*10-07-2	Partial Carcass	0.000928	0.000425	0.000265	0.000121
	*10-07-3	Partial Carcass	0.00168	0.000414	0.000493	0.000121
	*10-07-4	Partial Carcass	0.00169	0.000515	0.000398	0.000121

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-07-5	Partial Carcass	0.00119	0.000462	0.000314	0.000121
	*10-07-6	Partial Carcass	< 0.000476	0.000476	< 0.000121	0.000121
	*4-08-6	Partial Carcass	0.00108	0.000481	0.000274	0.000121
p,p'-DDD						
	*10-06-1	Partial Carcass	0.00668	0.000502	0.00178	0.000134
	*10-06-2	Partial Carcass	0.00628	0.000526	0.00160	0.000134
	*10-06-3	Partial Carcass	0.00583	0.000446	0.00175	0.000134
	*10-06-4	Partial Carcass	0.0103	0.000466	0.00295	0.000134
	*10-06-5	Partial Carcass	0.00519	0.000445	0.00156	0.000134
	*10-07-1	Partial Carcass	0.00246	0.000472	0.000696	0.000134
	*10-07-2	Partial Carcass	0.00292	0.000468	0.000833	0.000134
	*10-07-3	Partial Carcass	0.00663	0.000456	0.00195	0.000134
	*10-07-4	Partial Carcass	0.0122	0.000568	0.00287	0.000134
	*10-07-5	Partial Carcass	0.00556	0.000508	0.00146	0.000134
	*10-07-6	Partial Carcass	0.00295	0.000524	0.000752	0.000134
	*4-08-6	Partial Carcass	0.00418	0.000530	0.00106	0.000134
p,p'-DDE						
	*10-06-1	Partial Carcass	0.0409	0.000539	0.0109	0.000144
	*10-06-2	Partial Carcass	0.0546	0.000565	0.0139	0.000144
	*10-06-3	Partial Carcass	0.0621	0.000479	0.0186	0.000144
	*10-06-4	Partial Carcass	0.0858	0.000501	0.0246	0.000144
	*10-06-5	Partial Carcass	0.0457	0.000478	0.0137	0.000144
	*10-07-1	Partial Carcass	0.0275	0.000508	0.00778	0.000144
	*10-07-2	Partial Carcass	0.0281	0.000503	0.00802	0.000144
	*10-07-3	Partial Carcass	0.0591	0.000490	0.0174	0.000144
	*10-07-4	Partial Carcass	0.0959	0.000610	0.0226	0.000144
	*10-07-5	Partial Carcass	0.0329	0.000546	0.00865	0.000144
	*10-07-6	Partial Carcass	0.0525	0.000563	0.0134	0.000144
	*4-08-6	Partial Carcass	0.0421	0.000570	0.0106	0.000144

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
p,p'-DDT						
	*10-06-1	Partial Carcass	0.000879	0.000623	0.000234	0.000166
	*10-06-2	Partial Carcass	0.000790	0.000653	0.000201	0.000166
	*10-06-3	Partial Carcass	0.00221	0.000554	0.000663	0.000166
	*10-06-4	Partial Carcass	0.00285	0.000579	0.000819	0.000166
	*10-06-5	Partial Carcass	0.00113	0.000552	0.000339	0.000166
	*10-07-1	Partial Carcass	0.000594	0.000586	0.000168	0.000166
	*10-07-2	Partial Carcass	0.00106	0.000581	0.000303	0.000166
	*10-07-3	Partial Carcass	0.00229	0.000566	0.000673	0.000166
	*10-07-4	Partial Carcass	0.00323	0.000704	0.000761	0.000166
	*10-07-5	Partial Carcass	0.00110	0.000631	0.000290	0.000166
	*10-07-6	Partial Carcass	< 0.000651	0.000651	< 0.000166	0.000166
	*4-08-6	Partial Carcass	< 0.000658	0.000658	< 0.000166	0.000166
pentachloro-anisole						
	*10-06-1	Partial Carcass	0.00180	0.000569	0.000481	0.000152
	*10-06-2	Partial Carcass	0.000890	0.000596	0.000226	0.000152
	*10-06-3	Partial Carcass	0.00328	0.000506	0.000982	0.000152
	*10-06-4	Partial Carcass	0.00367	0.000528	0.00105	0.000152
	*10-06-5	Partial Carcass	0.00456	0.000504	0.00137	0.000152
	*10-07-1	Partial Carcass	0.000594	0.000535	0.000168	0.000152
	*10-07-2	Partial Carcass	< 0.000531	0.000531	< 0.000152	0.000152
	*10-07-3	Partial Carcass	0.00295	0.000517	0.000865	0.000152
	*10-07-4	Partial Carcass	0.00164	0.000643	0.000386	0.000152
	*10-07-5	Partial Carcass	0.00119	0.000576	0.000314	0.000152
	*10-07-6	Partial Carcass	0.00116	0.000594	0.000296	0.000152
	*4-08-6	Partial Carcass	0.00103	0.000601	0.000261	0.000152
perylene						
	1-08-1	Partial Carcass	< 0.00273	0.00273	< 0.000645	0.000645
	1-08-2	Partial Carcass	< 0.00244	0.00244	< 0.000639	0.000639

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	1-08-3	Partial Carcass	< 0.00234	0.00234	< 0.000640	0.000640
	1-08-4	Partial Carcass	< 0.00248	0.00248	< 0.000644	0.000644
	1-08-6	Partial Carcass	< 0.00236	0.00236	< 0.000628	0.000628
phenanthrene						
	1-08-1	Partial Carcass	0.0123	0.00124	0.00290	0.000292
	1-08-2	Partial Carcass	0.0179	0.00110	0.00470	0.000289
	1-08-3	Partial Carcass	0.0161	0.00106	0.00440	0.000290
	1-08-4	Partial Carcass	0.0193	0.00112	0.00500	0.000292
	1-08-6	Partial Carcass	0.00712	0.00107	0.00190	0.000284
phytane						
	1-08-1	Partial Carcass	0.864	0.0630	0.204	0.0149
	1-08-2	Partial Carcass	0.669	0.0563	0.175	0.0148
	1-08-3	Partial Carcass	1.32	0.0540	0.362	0.0148
	1-08-4	Partial Carcass	1.65	0.0573	0.428	0.0149
	1-08-6	Partial Carcass	1.04	0.0544	0.278	0.0145
pristane						
	*1-08-1	Partial Carcass	0.993	0.0610	0.235	0.0144
	1-08-2	Partial Carcass	0.503	0.0545	0.132	0.0143
	1-08-3	Partial Carcass	0.917	0.0523	0.251	0.0143
	1-08-4	Partial Carcass	1.19	0.0555	0.308	0.0144
	1-08-6	Partial Carcass	0.788	0.0526	0.210	0.0140
pyrene						
	1-08-1	Partial Carcass	0.00254	0.00170	0.000600	0.000402
	1-08-2	Partial Carcass	0.00306	0.00152	0.000800	0.000398
	1-08-3	Partial Carcass	0.00256	0.00146	0.000700	0.000399
	1-08-4	Partial Carcass	0.00694	0.00155	0.00180	0.000401
	1-08-6	Partial Carcass	< 0.00147	0.00147	< 0.000391	0.000391
toxaphene						
	*10-06-1	Partial Carcass	< 0.0396	0.0396	< 0.0105	0.0105

Analyte	Sample Number	Sample Matrix	Dry Weight (ppm)	DL Dry Weight (ppm)	Wet Weight (ppm)	DL Wet Weight (ppm)
	*10-06-2	Partial Carcass	< 0.0415	0.0415	< 0.0105	0.0105
	*10-06-3	Partial Carcass	< 0.0352	0.0352	< 0.0105	0.0105
	*10-06-4	Partial Carcass	< 0.0367	0.0367	< 0.0105	0.0105
	*10-06-5	Partial Carcass	< 0.0351	0.0351	< 0.0105	0.0105
	*10-07-1	Partial Carcass	< 0.0372	0.0372	< 0.0105	0.0105
	*10-07-2	Partial Carcass	< 0.0369	0.0369	< 0.0105	0.0105
	*10-07-3	Partial Carcass	< 0.0359	0.0359	< 0.0105	0.0105
	*10-07-4	Partial Carcass	< 0.0447	0.0447	< 0.0105	0.0105
	*10-07-5	Partial Carcass	< 0.0401	0.0401	< 0.0105	0.0105
	*10-07-6	Partial Carcass	< 0.0413	0.0413	< 0.0105	0.0105
	*4-08-6	Partial Carcass	< 0.0418	0.0418	< 0.0105	0.0105
trans-nonachlor						
	*10-06-1	Partial Carcass	0.00857	0.000571	0.00229	0.000152
	*10-06-2	Partial Carcass	0.0135	0.000599	0.00344	0.000152
	*10-06-3	Partial Carcass	0.0240	0.000508	0.00721	0.000152
	*10-06-4	Partial Carcass	0.0280	0.000531	0.00805	0.000152
	*10-06-5	Partial Carcass	0.0130	0.000507	0.00390	0.000152
	*10-07-1	Partial Carcass	0.00154	0.000538	0.000438	0.000152
	*10-07-2	Partial Carcass	0.00768	0.000533	0.00220	0.000152
	*10-07-3	Partial Carcass	0.0366	0.000519	0.0107	0.000152
	*10-07-4	Partial Carcass	0.0371	0.000646	0.00876	0.000152
	*10-07-5	Partial Carcass	0.0156	0.000579	0.00410	0.000152
	*10-07-6	Partial Carcass	0.00222	0.000597	0.000567	0.000152
	*4-08-6	Partial Carcass	0.00981	0.000604	0.00248	0.000152

* See "Laboratory Notes" section.

5. Procedural Blanks

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
1,2,3,4,6,7,8-HpCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,4,6,7,8-HpCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,4,7,8,9-HpCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,4,7,8-HxCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,4,7,8-HxCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,4-Tetrachlorobenzene					
	ENV2176A	Animal Tissue	0.000700	< 0.000151	Wet
1,2,3,6,7,8-HxCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,6,7,8-HxCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,7,8,9-HxCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,7,8,9-HxCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,7,8-PeCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,3,7,8-PeCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
1,2,4,5-Tetrachlorobenzene					
	ENV2176A	Animal Tissue	0.00130	< 0.000269	Wet
1,6,7-Trimethyl-naphthalene					
	ENV2175A	Animal Tissue	0.00230	< 0.000370	Wet
1-methylnaphthalene					
	ENV2175A	Animal Tissue	0.00380	< 0.000620	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
1-methylphenanthrene					
	ENV2175A	Animal Tissue	0.00260	< 0.000420	Wet
2,3,4,6,7,8-HxCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
2,3,4,7,8-PeCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
2,3,7,8-TCDD					
	EQ0900410-01	Water	0.000200	< 0.00000200	Wet
2,3,7,8-TCDF					
	EQ0900410-01	Water	0.000200	< 0.00000200	Wet
2,6-dimethylnaphthalene					
	ENV2175A	Animal Tissue	0.00330	< 0.000540	Wet
2-methylnaphthalene					
	ENV2175A	Animal Tissue	0.00410	< 0.000670	Wet
Aldrin					
	ENV2176A	Animal Tissue	0.000600	< 0.000122	Wet
BDE# 1					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 10					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 100					
	ENV2179A	Animal Tissue	0.00190	< 0.000144	Wet
BDE# 11					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 116					
	ENV2179A	Animal Tissue	0.00100	< 0.0000740	Wet
BDE# 118					
	ENV2179A	Animal Tissue	0.00100	< 0.0000740	Wet
BDE# 119					
	ENV2179A	Animal Tissue	0.00100	< 0.0000740	Wet
BDE# 12					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 126					

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	ENV2179A	Animal Tissue	0.00100	< 0.0000740	Wet
BDE# 13					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 138					
	ENV2179A	Animal Tissue	0.000700	< 0.0000550	Wet
BDE# 15					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 153					
	ENV2179A	Animal Tissue	0.000800	< 0.0000600	Wet
BDE# 154					
	ENV2179A	Animal Tissue	0.000900	< 0.0000680	Wet
BDE# 155					
	ENV2179A	Animal Tissue	0.000700	< 0.0000550	Wet
BDE# 166					
	ENV2179A	Animal Tissue	0.000700	< 0.0000550	Wet
BDE# 17					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 181					
	ENV2179A	Animal Tissue	0.000700	< 0.0000510	Wet
BDE# 183					
	ENV2179A	Animal Tissue	0.000700	< 0.0000510	Wet
BDE# 190					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 194					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 195					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 196					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 197					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 198/199/203/200					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
BDE# 2					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 201					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 202					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 204					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 205					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 206					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 207					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 208					
	ENV2179A	Animal Tissue	0.000500	< 0.0000390	Wet
BDE# 209					
	ENV2179A	Animal Tissue	0.00750	< 0.000575	Wet
BDE# 25					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 28					
	ENV2179A	Animal Tissue	0.00100	< 0.0000750	Wet
BDE# 3					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 30					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 32					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 33					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 35					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 37					

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 47					
	ENV2179A	Animal Tissue	0.00110	< 0.0000810	Wet
BDE# 49/71					
	ENV2179A	Animal Tissue	0.000800	< 0.0000600	Wet
BDE# 66					
	ENV2179A	Animal Tissue	0.000600	< 0.0000450	Wet
BDE# 7					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 75					
	ENV2179A	Animal Tissue	0.00110	< 0.0000810	Wet
BDE# 77					
	ENV2179A	Animal Tissue	0.00110	< 0.0000810	Wet
BDE# 8					
	ENV2179A	Animal Tissue	0.000900	< 0.0000700	Wet
BDE# 85					
	ENV2179A	Animal Tissue	0.00200	< 0.000152	Wet
BDE# 99					
	ENV2179A	Animal Tissue	0.00100	< 0.0000740	Wet
BDE-TOTAL					
	ENV2179A	Animal Tissue	0.00880	< 0.000677	Wet
BHC (Total)					
	ENV2176A	Animal Tissue	0.00170	< 0.000359	Wet
Benzo(a)anthracene					
	ENV2175A	Animal Tissue	0.00190	< 0.000310	Wet
C1-Fluoranthenes & Pyrenes					
	ENV2175A	Animal Tissue	0.00640	< 0.00105	Wet
C1-Phenanthrenes & Anthracenes					
	ENV2175A	Animal Tissue	0.00300	< 0.000490	Wet
C1-chrysenes					
	ENV2175A	Animal Tissue	0.00470	< 0.000770	Wet
C1-dibenzothiophenes					
	ENV2175A	Animal Tissue	0.00340	< 0.000560	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
C1-fluorenes					
	ENV2175A	Animal Tissue	0.00410	< 0.000680	Wet
C1-naphthalenes					
	ENV2175A	Animal Tissue	0.00790	< 0.00129	Wet
C2-Phenanthrenes & Anthracenes					
	ENV2175A	Animal Tissue	0.00300	< 0.000490	Wet
C2-chrysenes					
	ENV2175A	Animal Tissue	0.00470	< 0.000770	Wet
C2-dibenzothiophenes					
	ENV2175A	Animal Tissue	0.00340	< 0.000560	Wet
C2-fluorenes					
	ENV2175A	Animal Tissue	0.00410	< 0.000680	Wet
C2-naphthalenes					
	ENV2175A	Animal Tissue	0.0100	< 0.00164	Wet
C3-Phenanthrenes & Anthracenes					
	ENV2175A	Animal Tissue	0.00300	< 0.000490	Wet
C3-chrysenes					
	ENV2175A	Animal Tissue	0.00470	< 0.000770	Wet
C3-dibenzothiophenes					
	ENV2175A	Animal Tissue	0.00340	< 0.000560	Wet
C3-fluorenes					
	ENV2175A	Animal Tissue	0.00410	< 0.000680	Wet
C3-naphthalenes					
	ENV2175A	Animal Tissue	0.0100	< 0.00164	Wet
C4-Phenanthrenes & Anthracenes					
	ENV2175A	Animal Tissue	0.00300	< 0.000490	Wet
C4-chrysenes					
	ENV2175A	Animal Tissue	0.00470	< 0.000770	Wet
C4-naphthalenes					
	ENV2175A	Animal Tissue	0.0100	< 0.00164	Wet
Cl4-PCDD					
	EQ0900410-01	Water	0.000200	< 0.00000200	Wet
Cl4-PCDF					

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900410-01	Water	0.000200	< 0.00000200	Wet
Cl5-PCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
Cl5-PCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
Cl6-PCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
Cl6-PCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
Cl7-PCDD					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
Cl7-PCDF					
	EQ0900410-01	Water	0.000200	< 0.0000100	Wet
DDMU					
	ENV2176A	Animal Tissue	0.000600	< 0.000128	Wet
Dibenz(a,h)anthracene					
	ENV2175A	Animal Tissue	0.00210	< 0.000350	Wet
HCB					
	ENV2176A	Animal Tissue	0.00110	< 0.000226	Wet
Heptachlor					
	ENV2176A	Animal Tissue	0.000600	< 0.000133	Wet
OCDD					
	EQ0900410-01	Water	0.000200	< 0.0000200	Wet
OCDF					
	EQ0900410-01	Water	0.000200	< 0.0000200	Wet
PCB# 1					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 10					
	EQ0900409-01	Water	0.000200	< 0.0000100	Wet
	EQ0900415-01	Water	0.0000500	< 0.0000000500	Wet
PCB# 103					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 104					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 105					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 106					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 107					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 108/124					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 11					
	EQ0900409-01	Water	0.000200	0.000581	Wet
	EQ0900415-01	Water	0.000832	0.000000832	Wet
PCB# 110/115					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 111					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 112					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 114					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 117					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 118					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 12/13					
	EQ0900409-01	Water	0.000200	< 0.0000200	Wet
	EQ0900415-01	Water	0.000100	< 0.000000100	Wet
PCB# 120					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 121					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 122					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 123					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 126					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 127					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 128/166					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 129/138/163					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 130					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 131					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 132					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 133					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 134					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 135/151					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 136					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 137					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 139/140					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 14					
	EQ0900409-01	Water	0.000200	< 0.0000200	Wet
	EQ0900415-01	Water	0.000100	< 0.000000100	Wet
PCB# 141					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 142					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 143					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 144					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 145					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 146					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 147/149					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 148					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 15					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 150					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 152					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 153/168					
	EQ0900409-01	Water	0.000200	0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 154					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 155					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 156/157					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 158					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 159					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 16					
	EQ0900409-01	Water	0.000200	0.0000216	Wet
	EQ0900415-01	Water	0.000100	< 0.000000100	Wet
PCB# 160					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 161					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 162					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 164					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 165					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 167					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 169					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 17					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 170					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 171/173					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 172					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 174					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 175					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 176					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 177					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 178					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 179					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 18/30					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 180/193					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 181					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 182					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 183					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 184					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 185					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 186					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 187					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 188					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 189					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 19					
	EQ0900409-01	Water	0.000200	< 0.0000200	Wet
	EQ0900415-01	Water	0.000100	< 0.000000100	Wet
PCB# 190					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 191					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 192					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 194					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 195					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 196					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 197					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 198/199					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 2					
	EQ0900409-01	Water	0.000200	< 0.00000200	Wet
	EQ0900415-01	Water	0.0000100	< 0.0000000100	Wet
PCB# 20/28					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 200					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 201					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 202					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 203					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 204					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 205					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 206					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 207					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 208					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 209					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 21/33					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 22					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 23					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 24					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 25					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 26/29					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 27					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 3					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 31					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 32					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 34					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 35					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 36					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 37					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 38					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 39					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 4					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 40/41/70					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 42					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 43/73					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 44/47/65					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 45/51					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 46					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 48					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 49/69					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 5					
	EQ0900409-01	Water	0.000200	< 0.0000100	Wet
	EQ0900415-01	Water	0.0000500	< 0.0000000500	Wet
PCB# 50/53					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 52					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 54					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 55					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 56					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 57					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 58					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 59/62/75					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 6					
	EQ0900409-01	Water	0.000200	< 0.0000100	Wet
	EQ0900415-01	Water	0.0000500	< 0.0000000500	Wet
PCB# 60					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 63					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 64					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 66					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 67					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 68					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 7					
	EQ0900409-01	Water	0.000200	< 0.0000100	Wet
	EQ0900415-01	Water	0.0000500	< 0.0000000500	Wet
PCB# 70/61/74/76					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 72					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 77					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 78					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 79					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 8					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 80					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 81					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 82					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 83/99					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 84					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 85/116					
	EQ0900409-01	Water	0.000200	< 0.0000400	Wet
	EQ0900415-01	Water	0.000200	< 0.000000200	Wet
PCB# 86/87/97/109/119/125					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 88/91					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 89					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 9					
	EQ0900409-01	Water	0.000200	< 0.0000100	Wet
	EQ0900415-01	Water	0.0000500	< 0.0000000500	Wet
PCB# 90/101/113					
	EQ0900409-01	Water	0.000200	< 0.000200	Wet
	EQ0900415-01	Water	0.00100	< 0.00000100	Wet
PCB# 92					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 94					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 95					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 96					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 98/100					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB# 98/102					
	EQ0900409-01	Water	0.000200	< 0.000100	Wet
	EQ0900415-01	Water	0.000500	< 0.000000500	Wet
PCB-TOTAL					
	EQ0900409-01	Water	0.000200	0.00240	Wet
	EQ0900415-01	Water	0.00386	0.00000386	Wet
Pentachlorobenzene					
	ENV2176A	Animal Tissue	0.000500	< 0.000111	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
Total DDT's					
	ENV2176A	Animal Tissue	0.00390	< 0.000822	Wet
acenaphthalene					
	ENV2175A	Animal Tissue	0.00200	< 0.000330	Wet
acenaphthene					
	ENV2175A	Animal Tissue	0.00210	< 0.000340	Wet
alpha BHC					
	ENV2176A	Animal Tissue	0.00100	< 0.000209	Wet
alpha chlordane					
	ENV2176A	Animal Tissue	0.000900	< 0.000189	Wet
anthracene					
	ENV2175A	Animal Tissue	0.00120	< 0.000200	Wet
benzo(a)pyrene					
	ENV2175A	Animal Tissue	0.00230	< 0.000370	Wet
benzo(b)fluoranthene					
	ENV2175A	Animal Tissue	0.00160	< 0.000270	Wet
benzo(e)pyrene					
	ENV2175A	Animal Tissue	0.00240	< 0.000390	Wet
benzo(g,h,i)perylene					
	ENV2175A	Animal Tissue	0.00270	< 0.000440	Wet
benzo(k)fluoranthene					
	ENV2175A	Animal Tissue	0.00210	< 0.000340	Wet
beta BHC					
	ENV2176A	Animal Tissue	0.00110	< 0.000224	Wet
biphenyl					
	ENV2175A	Animal Tissue	0.00370	< 0.000610	Wet
chlorpyrifos					
	ENV2176A	Animal Tissue	0.00130	< 0.000277	Wet
chrysene					
	ENV2175A	Animal Tissue	0.00240	< 0.000390	Wet
cis-nonachlor					
	ENV2176A	Animal Tissue	0.000700	< 0.000140	Wet
delta BHC					

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	ENV2176A	Animal Tissue	0.000500	< 0.000109	Wet
dibenzothiophene					
	ENV2175A	Animal Tissue	0.00170	< 0.000280	Wet
dieldrin					
	ENV2176A	Animal Tissue	0.000900	< 0.000189	Wet
endosulfan I					
	ENV2176A	Animal Tissue	0.000700	< 0.000152	Wet
endosulfan II					
	ENV2176A	Animal Tissue	0.000700	< 0.000148	Wet
endosulfan sulfate					
	ENV2176A	Animal Tissue	0.000700	< 0.000154	Wet
endrin					
	ENV2176A	Animal Tissue	0.00100	< 0.000202	Wet
fluoranthene					
	ENV2175A	Animal Tissue	0.00150	< 0.000250	Wet
fluorene					
	ENV2175A	Animal Tissue	0.00210	< 0.000340	Wet
gamma BHC					
	ENV2176A	Animal Tissue	0.000400	< 0.0000940	Wet
gamma chlordane					
	ENV2176A	Animal Tissue	0.000700	< 0.000155	Wet
heptachlor epoxide					
	ENV2176A	Animal Tissue	0.000800	< 0.000160	Wet
indeno(1,2,3-cd)pyrene					
	ENV2175A	Animal Tissue	0.00350	< 0.000570	Wet
mirex					
	ENV2176A	Animal Tissue	0.000600	< 0.000123	Wet
n-decane					
	ENV2175A	Animal Tissue	0.0649	< 0.0106	Wet
n-docosane					
	ENV2175A	Animal Tissue	0.0821	< 0.0135	Wet
n-dodecane					
	ENV2175A	Animal Tissue	0.0475	< 0.00779	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
n-dotriacontane					
	ENV2175A	Animal Tissue	0.0748	< 0.0123	Wet
n-eicosane					
	ENV2175A	Animal Tissue	0.0836	< 0.0137	Wet
n-heneicosane					
	ENV2175A	Animal Tissue	0.0879	< 0.0144	Wet
n-hentriacontane					
	ENV2175A	Animal Tissue	0.0764	< 0.0125	Wet
n-heptacosane					
	ENV2175A	Animal Tissue	0.0728	< 0.0119	Wet
n-heptadecane					
	ENV2175A	Animal Tissue	0.0669	< 0.0110	Wet
n-hexacosane					
	ENV2175A	Animal Tissue	0.0746	< 0.0122	Wet
n-hexadecane					
	ENV2175A	Animal Tissue	0.0822	< 0.0135	Wet
n-nonacosane					
	ENV2175A	Animal Tissue	0.0669	< 0.0110	Wet
n-nonadecane					
	ENV2175A	Animal Tissue	0.0685	< 0.0112	Wet
n-octacosane					
	ENV2175A	Animal Tissue	0.0723	< 0.0118	Wet
n-octadecane					
	ENV2175A	Animal Tissue	0.0806	< 0.0132	Wet
n-pentacosane					
	ENV2175A	Animal Tissue	0.0843	< 0.0138	Wet
n-pentadecane					
	ENV2175A	Animal Tissue	0.104	< 0.0170	Wet
n-tetracosane					
	ENV2175A	Animal Tissue	0.0663	< 0.0109	Wet
n-tetradecane					
	ENV2175A	Animal Tissue	0.0824	< 0.0135	Wet
n-tetratriacontane					

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
	ENV2175A	Animal Tissue	0.0732	< 0.0120	Wet
n-triacontane					
	ENV2175A	Animal Tissue	0.0709	< 0.0116	Wet
n-tricosane					
	ENV2175A	Animal Tissue	0.0798	< 0.0131	Wet
n-tridecane					
	ENV2175A	Animal Tissue	0.0823	< 0.0135	Wet
n-tritriacontane					
	ENV2175A	Animal Tissue	0.0636	< 0.0104	Wet
n-undecane					
	ENV2175A	Animal Tissue	0.0718	< 0.0118	Wet
naphthalene					
	ENV2175A	Animal Tissue	0.00500	< 0.000820	Wet
o,p'-DDD					
	ENV2176A	Animal Tissue	0.00100	< 0.000206	Wet
o,p'-DDE					
	ENV2176A	Animal Tissue	0.000600	< 0.000119	Wet
o,p'-DDT					
	ENV2176A	Animal Tissue	0.000700	< 0.000155	Wet
oxychlordane					
	ENV2176A	Animal Tissue	0.000600	< 0.000121	Wet
p,p'-DDD					
	ENV2176A	Animal Tissue	0.000600	< 0.000134	Wet
p,p'-DDE					
	ENV2176A	Animal Tissue	0.000700	< 0.000144	Wet
p,p'-DDT					
	ENV2176A	Animal Tissue	0.000800	< 0.000166	Wet
pentachloro-anisole					
	ENV2176A	Animal Tissue	0.000700	< 0.000152	Wet
perylene					
	ENV2175A	Animal Tissue	0.00550	0.000900	Wet
phenanthrene					
	ENV2175A	Animal Tissue	0.00150	< 0.000240	Wet

Analyte	Lab Sample Number	Lab Sample Matrix	Result Total UG	** BEC (ppm/%)	Basis
phytane					
	ENV2175A	Animal Tissue	0.0746	< 0.0122	Wet
pristane					
	ENV2175A	Animal Tissue	0.0723	< 0.0118	Wet
pyrene					
	ENV2175A	Animal Tissue	0.00200	< 0.000330	Wet
toxaphene					
	ENV2176A	Animal Tissue	0.0500	< 0.0105	Wet
trans-nonachlor					
	ENV2176A	Animal Tissue	0.000700	< 0.000152	Wet

** Blank Equivalent Concentration

6. Duplicates

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
% Lipid								
	1-08-2		Partial Carcass	Percent	4.30	4.33	4.31	0.700
	10-07-5		Partial Carcass	Percent	5.77	5.84	5.80	1.21
	4-08-4		Partial Carcass	Percent	5.49	6.23	5.86	12.6
% Moisture								
	1-08-2		Partial Carcass	Percent	73.8	73.8	73.8	0.000
	10-07-5		Partial Carcass	Percent	73.7	73.7	73.7	0.000
	4-08-4		Partial Carcass	Percent	73.3	73.3	73.3	0.000
1,2,3,4,6,7,8-HpCDD								
	EQ0900410-02	SD	Water	Wet	0.000206	0.000214	0.000210	3.81
1,2,3,4,6,7,8-HpCDF								
	EQ0900410-02	SD	Water	Wet	0.000204	0.000205	0.000204	0.490
1,2,3,4,7,8,9-HpCDF								
	EQ0900410-02	SD	Water	Wet	0.000194	0.000197	0.000195	1.53
1,2,3,4,7,8-HxCDD								
	EQ0900410-02	SD	Water	Wet	0.000180	0.000186	0.000183	3.28
1,2,3,4,7,8-HxCDF								
	EQ0900410-02	SD	Water	Wet	0.000212	0.000212	0.000212	0.000
1,2,3,4-Tetrachlorobenzene								
	10-07-5		Partial Carcass	Wet	< 0.000151	< 0.000151	0.0000755	0.000

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
1,2,3,6,7,8-HxCDD								
	EQ0900410-02	SD	Water	Wet	0.000224	0.000236	0.000230	5.22
1,2,3,6,7,8-HxCDF								
	EQ0900410-02	SD	Water	Wet	0.000204	0.000213	0.000208	4.32
1,2,3,7,8,9-HxCDD								
	EQ0900410-02	SD	Water	Wet	0.000188	0.000189	0.000188	0.530
1,2,3,7,8,9-HxCDF								
	EQ0900410-02	SD	Water	Wet	0.000189	0.000195	0.000192	3.12
1,2,3,7,8-PeCDD								
	EQ0900410-02	SD	Water	Wet	0.000199	0.000200	0.000200	0.500
1,2,3,7,8-PeCDF								
	EQ0900410-02	SD	Water	Wet	0.000201	0.000205	0.000203	1.97
1,2,4,5-Tetrachlorobenzene								
	10-07-5		Partial Carcass	Wet	< 0.000269	< 0.000269	0.000134	0.000
1,6,7-Trimethyl-naphthalene								
	1-08-2		Partial Carcass	Wet	< 0.000446	< 0.000450	0.000224	0.890
1-methylnaphthalene								
	1-08-2		Partial Carcass	Wet	0.00370	0.00340	0.00355	8.45
1-methylphenanthrene								
	1-08-2		Partial Carcass	Wet	0.00780	0.00770	0.00775	1.29
2,3,4,6,7,8-HxCDF								
	EQ0900410-02	SD	Water	Wet	0.000194	0.000201	0.000198	3.54
2,3,4,7,8-PeCDF								

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	EQ0900410-02	SD	Water	Wet	0.000202	0.000210	0.000206	3.88
2,3,7,8-TCDD								
	EQ0900410-02	SD	Water	Wet	0.0000402	0.0000405	0.0000404	0.740
2,3,7,8-TCDF								
	EQ0900410-02	SD	Water	Wet	0.0000400	0.0000414	0.0000407	3.44
2,6-dimethylnaphthalene								
	1-08-2		Partial Carcass	Wet	0.00100	0.00120	0.00110	18.2
2-methylnaphthalene								
	1-08-2		Partial Carcass	Wet	0.00680	0.00650	0.00665	4.51
Aldrin								
	10-07-5		Partial Carcass	Wet	< 0.000122	< 0.000122	0.0000610	0.000
BDE# 1								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 10								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 100								
	4-08-4		Partial Carcass	Wet	0.0161	0.0154	0.0158	4.44
BDE# 11								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 116								
	4-08-4		Partial Carcass	Wet	< 0.000192	< 0.000187	0.0000948	2.64
BDE# 118								
	4-08-4		Partial	Wet	< 0.000192	< 0.000187	0.0000948	2.64

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
			Carcass					
BDE# 119								
	4-08-4		Partial Carcass	Wet	0.000800	0.000700	0.000750	13.3
BDE# 12								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 126								
	4-08-4		Partial Carcass	Wet	0.000300	0.000300	0.000300	0.000
BDE# 13								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 138								
	4-08-4		Partial Carcass	Wet	< 0.000143	< 0.000139	0.0000705	2.84
BDE# 15								
	4-08-4		Partial Carcass	Wet	0.000200	0.000200	0.000200	0.000
BDE# 153								
	4-08-4		Partial Carcass	Wet	0.00220	0.00220	0.00220	0.000
BDE# 154								
	4-08-4		Partial Carcass	Wet	0.00490	0.00500	0.00495	2.02
BDE# 155								
	4-08-4		Partial Carcass	Wet	< 0.000143	< 0.000139	0.0000705	2.84
BDE# 166								
	4-08-4		Partial Carcass	Wet	< 0.000143	< 0.000139	0.0000705	2.84
BDE# 17								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
BDE# 181								
	4-08-4		Partial Carcass	Wet	< 0.000133	< 0.000129	0.0000655	3.05
BDE# 183								
	4-08-4		Partial Carcass	Wet	0.000300	0.000300	0.000300	0.000
BDE# 190								
	4-08-4		Partial Carcass	Wet	0.000200	0.000200	0.000200	0.000
BDE# 194								
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 195								
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 196								
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 197								
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 198/199/203/200								
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 2								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 201								
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 202								
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 204								

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	4-08-4		Partial Carcass	Wet	< 0.000101	< 0.0000980	0.0000498	3.02
BDE# 205								
	4-08-4		Partial Carcass	Wet	0.000600	0.000700	0.000650	15.4
BDE# 206								
	4-08-4		Partial Carcass	Wet	0.000600	0.000500	0.000550	18.2
BDE# 207								
	4-08-4		Partial Carcass	Wet	0.000700	0.000600	0.000650	15.4
BDE# 208								
	4-08-4		Partial Carcass	Wet	0.000400	0.000500	0.000450	22.2
BDE# 209								
	4-08-4		Partial Carcass	Wet	< 0.00149	< 0.00145	0.000737	2.78
BDE# 25								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 28								
	4-08-4		Partial Carcass	Wet	0.00250	0.00250	0.00250	0.000
BDE# 3								
	4-08-4		Partial Carcass	Wet	0.000700	0.000800	0.000750	13.3
BDE# 30								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 32								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 33								
	4-08-4		Partial	Wet	< 0.000182	< 0.000177	0.0000898	2.79

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
			Carcass					
BDE# 35								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 37								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 47								
	4-08-4		Partial Carcass	Wet	0.129	0.126	0.128	2.35
BDE# 49/71								
	4-08-4		Partial Carcass	Wet	0.00320	0.00330	0.00325	3.08
BDE# 66								
	4-08-4		Partial Carcass	Wet	< 0.000116	< 0.000113	0.0000572	2.62
BDE# 7								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 75								
	4-08-4		Partial Carcass	Wet	< 0.000212	< 0.000206	0.000104	2.87
BDE# 77								
	4-08-4		Partial Carcass	Wet	0.000500	0.000500	0.000500	0.000
BDE# 8								
	4-08-4		Partial Carcass	Wet	< 0.000182	< 0.000177	0.0000898	2.79
BDE# 85								
	4-08-4		Partial Carcass	Wet	0.000400	< 0.000385	0.000296	70.0
BDE# 99								
	4-08-4		Partial Carcass	Wet	< 0.000192	< 0.000187	0.0000948	2.64

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
BDE-TOTAL								
	4-08-4		Partial Carcass	Wet	0.164	0.160	0.162	2.41
BHC (Total)								
	10-07-5		Partial Carcass	Wet	< 0.000359	< 0.000359	0.000180	0.000
Benzo(a)anthracene								
	1-08-2		Partial Carcass	Wet	< 0.000374	< 0.000377	0.000188	0.800
C1-Fluoranthenes & Pyrenes								
	1-08-2		Partial Carcass	Wet	< 0.00127	< 0.00128	0.000636	0.790
C1-Phenanthrenes & Anthracenes								
	1-08-2		Partial Carcass	Wet	0.00790	0.00780	0.00785	1.27
C1-chrysenes								
	1-08-2		Partial Carcass	Wet	< 0.000928	< 0.000936	0.000466	0.860
C1-dibenzothiophenes								
	1-08-2		Partial Carcass	Wet	< 0.000675	< 0.000681	0.000339	0.880
C1-fluorenes								
	1-08-2		Partial Carcass	Wet	0.00260	0.00240	0.00250	8.00
C1-naphthalenes								
	1-08-2		Partial Carcass	Wet	0.00680	0.00640	0.00660	6.06
C2-Phenanthrenes & Anthracenes								
	1-08-2		Partial Carcass	Wet	< 0.000591	< 0.000596	0.000297	0.840
C2-chrysenes								
	1-08-2		Partial Carcass	Wet	< 0.000928	< 0.000936	0.000466	0.860
C2-dibenzothiophenes								

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	1-08-2		Partial Carcass	Wet	< 0.000675	< 0.000681	0.000339	0.880
C2-fluorenes								
	1-08-2		Partial Carcass	Wet	< 0.000820	< 0.000826	0.000412	0.730
C2-naphthalenes								
	1-08-2		Partial Carcass	Wet	0.00370	0.00330	0.00350	11.4
C3-Phenanthrenes & Anthracenes								
	1-08-2		Partial Carcass	Wet	< 0.000591	< 0.000596	0.000297	0.840
C3-chrysenes								
	1-08-2		Partial Carcass	Wet	< 0.000928	< 0.000936	0.000466	0.860
C3-dibenzothiophenes								
	1-08-2		Partial Carcass	Wet	< 0.000675	< 0.000681	0.000339	0.880
C3-fluorenes								
	1-08-2		Partial Carcass	Wet	< 0.000820	< 0.000826	0.000412	0.730
C3-naphthalenes								
	1-08-2		Partial Carcass	Wet	< 0.00198	< 0.00199	0.000992	0.810
C4-Phenanthrenes & Anthracenes								
	1-08-2		Partial Carcass	Wet	< 0.000591	< 0.000596	0.000297	0.840
C4-chrysenes								
	1-08-2		Partial Carcass	Wet	< 0.000928	< 0.000936	0.000466	0.860
C4-naphthalenes								
	1-08-2		Partial Carcass	Wet	< 0.00198	< 0.00199	0.000992	0.810
DDMU								
	10-07-5		Partial	Wet	0.000340	0.000372	0.000356	8.99

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
			Carcass					
Dibenz(a,h)anthracene								
	1-08-2		Partial Carcass	Wet	< 0.000422	< 0.000425	0.000212	0.710
HCB								
	10-07-5		Partial Carcass	Wet	< 0.000226	< 0.000226	0.000113	0.000
Heptachlor								
	10-07-5		Partial Carcass	Wet	< 0.000133	< 0.000133	0.0000665	0.000
OCDD								
	EQ0900410-02	SD	Water	Wet	0.000464	0.000439	0.000451	5.54
OCDF								
	EQ0900410-02	SD	Water	Wet	0.000467	0.000468	0.000468	0.210
PCB# 1								
	EQ0900409-02	SD	Water	Wet	0.000185	0.000193	0.000189	4.23
	EQ0900415-02	SD	Water	Wet	0.000000906	0.000000937	0.000000922	3.36
PCB# 104								
	EQ0900409-02	SD	Water	Wet	0.000181	0.000196	0.000188	7.96
	EQ0900415-02	SD	Water	Wet	0.000000940	0.000000973	0.000000956	3.45
PCB# 105								
	EQ0900409-02	SD	Water	Wet	0.000227	0.000227	0.000227	0.000
	EQ0900415-02	SD	Water	Wet	0.00000102	0.00000113	0.00000107	10.2
PCB# 114								
	EQ0900409-02	SD	Water	Wet	0.000188	0.000206	0.000197	9.14

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	EQ0900415-02	SD	Water	Wet	0.000000880	0.000000964	0.000000922	9.11
PCB# 118								
	EQ0900409-02	SD	Water	Wet	0.000283	0.000263	0.000273	7.33
	EQ0900415-02	SD	Water	Wet	0.00000114	0.00000122	0.00000118	6.78
PCB# 123								
	EQ0900409-02	SD	Water	Wet	0.000184	0.000206	0.000195	11.3
	EQ0900415-02	SD	Water	Wet	0.000000870	0.000000950	0.000000910	8.79
PCB# 126								
	EQ0900409-02	SD	Water	Wet	0.000180	0.000198	0.000189	9.52
	EQ0900415-02	SD	Water	Wet	0.000000853	0.000000921	0.000000887	7.67
PCB# 15								
	EQ0900409-02	SD	Water	Wet	0.000204	0.000215	0.000210	5.25
	EQ0900415-02	SD	Water	Wet	0.00000100	0.00000107	0.00000103	6.76
PCB# 155								
	EQ0900409-02	SD	Water	Wet	< 0.000200	0.000212	0.000156	71.8
	EQ0900415-02	SD	Water	Wet	< 0.00000100	< 0.00000100	0.000000500	0.000
PCB# 156/157								
	EQ0900409-02	SD	Water	Wet	0.000411	0.000433	0.000422	5.21
	EQ0900415-02	SD	Water	Wet	0.00000185	0.00000202	0.00000194	8.79
PCB# 167								
	EQ0900409-02	SD	Water	Wet	0.000198	0.000212	0.000205	6.83

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	EQ0900415-02	SD	Water	Wet	0.000000932	0.00000102	0.000000976	9.02
PCB# 169								
	EQ0900409-02	SD	Water	Wet	0.000195	0.000220	0.000208	12.0
	EQ0900415-02	SD	Water	Wet	0.000000947	0.00000104	0.000000993	9.36
PCB# 188								
	EQ0900409-02	SD	Water	Wet	0.000185	0.000206	0.000195	10.7
	EQ0900415-02	SD	Water	Wet	0.000000876	0.000000927	0.000000902	5.66
PCB# 189								
	EQ0900409-02	SD	Water	Wet	0.000207	0.000225	0.000216	8.33
	EQ0900415-02	SD	Water	Wet	0.000000965	0.00000104	0.00000100	7.48
PCB# 19								
	EQ0900409-02	SD	Water	Wet	0.000172	0.000192	0.000182	11.0
	EQ0900415-02	SD	Water	Wet	0.000000956	0.000000958	0.000000957	0.210
PCB# 202								
	EQ0900409-02	SD	Water	Wet	< 0.000200	0.000221	0.000160	75.4
	EQ0900415-02	SD	Water	Wet	< 0.00000100	0.00000107	0.000000785	72.6
PCB# 205								
	EQ0900409-02	SD	Water	Wet	< 0.000200	0.000210	0.000155	71.0
	EQ0900415-02	SD	Water	Wet	< 0.00000100	< 0.00000100	0.000000500	0.000
PCB# 206								
	EQ0900409-02	SD	Water	Wet	0.000220	0.000217	0.000218	1.37

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	EQ0900415-02	SD	Water	Wet	< 0.00000100	0.00000105	0.000000775	71.0
PCB# 208								
	EQ0900409-02	SD	Water	Wet	0.000208	0.000220	0.000214	5.61
	EQ0900415-02	SD	Water	Wet	< 0.00000100	0.00000103	0.000000765	69.3
PCB# 209								
	EQ0900409-02	SD	Water	Wet	0.000180	0.000192	0.000186	6.45
	EQ0900415-02	SD	Water	Wet	0.000000930	0.000000993	0.000000962	6.55
PCB# 3								
	EQ0900409-02	SD	Water	Wet	0.000194	0.000204	0.000199	5.03
	EQ0900415-02	SD	Water	Wet	0.000000913	0.000000952	0.000000932	4.18
PCB# 37								
	EQ0900409-02	SD	Water	Wet	0.000199	0.000210	0.000204	5.38
	EQ0900415-02	SD	Water	Wet	0.000000950	0.00000105	0.00000100	10.0
PCB# 4								
	EQ0900409-02	SD	Water	Wet	0.000190	0.000195	0.000193	2.60
	EQ0900415-02	SD	Water	Wet	0.000000928	0.00000101	0.000000969	8.46
PCB# 54								
	EQ0900409-02	SD	Water	Wet	0.000181	0.000206	0.000194	12.9
	EQ0900415-02	SD	Water	Wet	0.000000881	0.000000970	0.000000926	9.62
PCB# 77								
	EQ0900409-02	SD	Water	Wet	0.000198	0.000213	0.000206	7.30

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	EQ0900415-02	SD	Water	Wet	0.000000936	0.00000101	0.000000973	7.61
PCB# 81								
	EQ0900409-02	SD	Water	Wet	0.000188	0.000205	0.000196	8.65
	EQ0900415-02	SD	Water	Wet	0.000000882	0.000000976	0.000000929	10.1
Pentachlorobenzene								
	10-07-5		Partial Carcass	Wet	0.00249	0.00257	0.00253	3.17
Total DDT's								
	10-07-5		Partial Carcass	Wet	0.0115	0.0121	0.0118	5.33
acenaphthalene								
	1-08-2		Partial Carcass	Wet	0.000400	< 0.000401	0.000300	66.4
acenaphthene								
	1-08-2		Partial Carcass	Wet	< 0.000410	< 0.000413	0.000206	0.730
alpha BHC								
	10-07-5		Partial Carcass	Wet	< 0.000209	< 0.000209	0.000104	0.000
alpha chlordane								
	10-07-5		Partial Carcass	Wet	0.00323	0.00325	0.00324	0.830
anthracene								
	1-08-2		Partial Carcass	Wet	0.000300	0.000300	0.000300	0.000
benzo(a)pyrene								
	1-08-2		Partial Carcass	Wet	< 0.000446	< 0.000450	0.000224	0.890
benzo(b)fluoranthene								
	1-08-2		Partial Carcass	Wet	0.000400	0.000400	0.000400	0.000

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
benzo(e)pyrene								
	1-08-2		Partial Carcass	Wet	< 0.000470	< 0.000474	0.000236	0.850
benzo(g,h,i)perylene								
	1-08-2		Partial Carcass	Wet	< 0.000530	< 0.000535	0.000266	0.940
benzo(k)fluoranthene								
	1-08-2		Partial Carcass	Wet	< 0.000410	< 0.000413	0.000206	0.730
beta BHC								
	10-07-5		Partial Carcass	Wet	< 0.000224	< 0.000224	0.000112	0.000
biphenyl								
	1-08-2		Partial Carcass	Wet	0.00180	0.00280	0.00230	43.5
chlorpyrifos								
	10-07-5		Partial Carcass	Wet	< 0.000277	< 0.000277	0.000138	0.000
chrysene								
	1-08-2		Partial Carcass	Wet	0.000500	0.000500	0.000500	0.000
cis-nonachlor								
	10-07-5		Partial Carcass	Wet	0.00174	0.00183	0.00178	4.93
delta BHC								
	10-07-5		Partial Carcass	Wet	< 0.000109	< 0.000109	0.0000545	0.000
dibenzothiophene								
	1-08-2		Partial Carcass	Wet	0.000400	0.000400	0.000400	0.000
dieldrin								
	10-07-5		Partial Carcass	Wet	0.00137	0.00144	0.00140	5.07
endosulfan I								

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	10-07-5		Partial Carcass	Wet	0.00214	0.00223	0.00219	3.98
endosulfan II								
	10-07-5		Partial Carcass	Wet	< 0.000148	< 0.000148	0.0000740	0.000
endosulfan sulfate								
	10-07-5		Partial Carcass	Wet	0.00198	0.00193	0.00196	2.71
endrin								
	10-07-5		Partial Carcass	Wet	< 0.000202	< 0.000202	0.000101	0.000
fluoranthene								
	1-08-2		Partial Carcass	Wet	0.00180	0.00160	0.00170	11.8
fluorene								
	1-08-2		Partial Carcass	Wet	0.00150	0.00160	0.00155	6.45
gamma BHC								
	10-07-5		Partial Carcass	Wet	< 0.0000940	< 0.0000940	0.0000470	0.000
gamma chlordane								
	10-07-5		Partial Carcass	Wet	0.00158	0.00159	0.00159	0.380
heptachlor epoxide								
	10-07-5		Partial Carcass	Wet	0.000248	0.000185	0.000216	29.1
indeno(1,2,3-cd)pyrene								
	1-08-2		Partial Carcass	Wet	< 0.000687	< 0.000693	0.000345	0.870
mirex								
	10-07-5		Partial Carcass	Wet	< 0.000123	< 0.000123	0.0000615	0.000
n-decane								
	1-08-2		Partial	Wet	0.0479	0.0489	0.0484	1.89

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
			Carcass					
n-docosane								
	1-08-2		Partial Carcass	Wet	< 0.0162	< 0.0164	0.00814	0.810
n-dodecane								
	1-08-2		Partial Carcass	Wet	0.0327	0.0303	0.0315	7.52
n-dotriacontane								
	1-08-2		Partial Carcass	Wet	< 0.0148	< 0.0149	0.00742	0.810
n-eicosane								
	1-08-2		Partial Carcass	Wet	< 0.0165	< 0.0167	0.00830	0.800
n-heneicosane								
	1-08-2		Partial Carcass	Wet	< 0.0174	< 0.0175	0.00872	0.810
n-hentriacontane								
	1-08-2		Partial Carcass	Wet	< 0.0151	< 0.0152	0.00758	0.810
n-heptacosane								
	1-08-2		Partial Carcass	Wet	0.102	0.0971	0.0994	4.60
n-heptadecane								
	1-08-2		Partial Carcass	Wet	6.49	6.07	6.28	6.68
n-hexacosane								
	1-08-2		Partial Carcass	Wet	0.0907	0.0954	0.0931	5.04
n-hexadecane								
	1-08-2		Partial Carcass	Wet	0.337	0.315	0.326	6.77
n-nonacosane								
	1-08-2		Partial Carcass	Wet	< 0.0132	< 0.0133	0.00663	0.810

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
n-nonadecane								
	1-08-2		Partial Carcass	Wet	0.419	0.412	0.416	1.81
n-octacosane								
	1-08-2		Partial Carcass	Wet	< 0.0143	< 0.0144	0.00717	0.800
n-octadecane								
	1-08-2		Partial Carcass	Wet	3.36	3.09	3.22	8.22
n-pentacosane								
	1-08-2		Partial Carcass	Wet	0.327	0.352	0.340	7.35
n-pentadecane								
	1-08-2		Partial Carcass	Wet	5.61	5.17	5.39	8.18
n-tetracosane								
	1-08-2		Partial Carcass	Wet	0.211	0.216	0.213	1.91
n-tetradecane								
	1-08-2		Partial Carcass	Wet	0.132	0.122	0.127	7.91
n-tetratriacontane								
	1-08-2		Partial Carcass	Wet	< 0.0145	< 0.0146	0.00726	0.810
n-triacontane								
	1-08-2		Partial Carcass	Wet	< 0.0140	< 0.0141	0.00703	0.800
n-tricosane								
	1-08-2		Partial Carcass	Wet	6.42	6.75	6.59	5.01
n-tridecane								
	1-08-2		Partial Carcass	Wet	0.333	0.329	0.331	1.00
n-tritriacontane								

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
	1-08-2		Partial Carcass	Wet	< 0.0126	< 0.0127	0.00631	0.810
n-undecane								
	1-08-2		Partial Carcass	Wet	0.0410	0.0419	0.0415	2.04
naphthalene								
	1-08-2		Partial Carcass	Wet	0.0139	0.0118	0.0128	16.3
o,p'-DDD								
	10-07-5		Partial Carcass	Wet	0.000399	0.000340	0.000370	16.0
o,p'-DDE								
	10-07-5		Partial Carcass	Wet	0.000375	0.000378	0.000376	0.800
o,p'-DDT								
	10-07-5		Partial Carcass	Wet	< 0.000155	< 0.000155	0.0000775	0.000
oxychlordane								
	10-07-5		Partial Carcass	Wet	0.000314	0.000340	0.000327	7.95
p,p'-DDD								
	10-07-5		Partial Carcass	Wet	0.00146	0.00155	0.00151	5.91
p,p'-DDE								
	10-07-5		Partial Carcass	Wet	0.00865	0.00925	0.00895	6.72
p,p'-DDT								
	10-07-5		Partial Carcass	Wet	0.000290	0.000252	0.000271	14.0
pentachloro-anisole								
	10-07-5		Partial Carcass	Wet	0.000314	0.000366	0.000340	15.3
perylene								
	1-08-2		Partial	Wet	< 0.000639	< 0.000644	0.000321	0.780

Analyte	Sample Number	**	Sample Matrix	Basis	Initial Result (ppm/%)	Duplicate Result (ppm/%)	Average	Relative Percent Diff.
			Carcass					
phenanthrene								
	1-08-2		Partial Carcass	Wet	0.00470	0.00480	0.00475	2.11
phytane								
	1-08-2		Partial Carcass	Wet	0.175	0.177	0.176	0.980
pristane								
	1-08-2		Partial Carcass	Wet	0.132	0.131	0.131	0.380
pyrene								
	1-08-2		Partial Carcass	Wet	0.000800	0.000700	0.000750	13.3
toxaphene								
	10-07-5		Partial Carcass	Wet	< 0.0105	< 0.0105	0.00527	0.000
trans-nonachlor								
	10-07-5		Partial Carcass	Wet	0.00410	0.00426	0.00418	3.92

* See "Laboratory Notes" section.** SD = Spiked Duplicate Result

7. Spike Recoveries

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
1,2,3,4,6,7,8-HpCDD								
	*EQ0900410-02		Water	Wet	0.000200	0.000206		103
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000214		107
1,2,3,4,6,7,8-HpCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000204		102
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000205		102
1,2,3,4,7,8,9-HpCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000194		97.0
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000197		98.5
1,2,3,4,7,8-HxCDD								
	*EQ0900410-02		Water	Wet	0.000200	0.000180		90.0
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000186		93.0
1,2,3,4,7,8-HxCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000212		106
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000212		106
1,2,3,4-Tetrachlorobenzene								
	10-06-3		Partial Carcass	Wet	0.00790	0.00745	105	94.3
1,2,3,6,7,8-HxCDD								
	*EQ0900410-02		Water	Wet	0.000200	0.000224		112
	*EQ0900410-	SD	Water	Wet	0.000200	0.000236		118

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	02							
1,2,3,6,7,8-HxCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000204		102
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000213		106
1,2,3,7,8,9-HxCDD								
	*EQ0900410-02		Water	Wet	0.000200	0.000188		94.0
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000189		94.5
1,2,3,7,8,9-HxCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000189		94.5
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000195		97.5
1,2,3,7,8-PeCDD								
	*EQ0900410-02		Water	Wet	0.000200	0.000199		99.5
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000200		100.
1,2,3,7,8-PeCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000201		100.
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000205		102
1,2,4,5-Tetrachlorobenzene								
	10-06-3		Partial Carcass	Wet	0.00790	0.00519	58.7	65.7
1,6,7-Trimethyl-naphthalene								
	1-08-6		Partial Carcass	Wet	0.0190	0.0219	86.8	115
1-methylnaphthalene								
	1-08-6		Partial	Wet	0.0189	0.0213	9.00	113

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
			Carcass					
1-methylphenanthrene								
	1-08-6		Partial Carcass	Wet	0.0190	0.0211	4.04	111
2,3,4,6,7,8-HxCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000194		97.0
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000201		100.
2,3,4,7,8-PeCDF								
	*EQ0900410-02		Water	Wet	0.000200	0.000202		101
	*EQ0900410-02	SD	Water	Wet	0.000200	0.000210		105
2,3,7,8-TCDD								
	*EQ0900410-02		Water	Wet	0.0000400	0.0000402		100.
	*EQ0900410-02	SD	Water	Wet	0.0000400	0.0000405		101
2,3,7,8-TCDF								
	*EQ0900410-02		Water	Wet	0.0000400	0.0000400		100.
	*EQ0900410-02	SD	Water	Wet	0.0000400	0.0000414		104
2,6-dimethylnaphthalene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0202	60.0	105
2-methylnaphthalene								
	1-08-6		Partial Carcass	Wet	0.0190	0.0211	5.00	111
Aldrin								
	10-06-3		Partial Carcass	Wet	0.00790	0.00734	130.	92.9
BDE# 100								

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	4-08-3		Partial Carcass	Wet	0.0196	0.0226	2.20	115
BDE# 138								
	4-08-3		Partial Carcass	Wet	0.0197	0.0229	283	116
BDE# 153								
	4-08-3		Partial Carcass	Wet	0.0196	0.0229	15.1	117
BDE# 154								
	4-08-3		Partial Carcass	Wet	0.0197	0.0227	7.30	115
BDE# 17								
	4-08-3		Partial Carcass	Wet	0.0194	0.0202	220.	104
BDE# 183								
	4-08-3		Partial Carcass	Wet	0.0196	0.0228	304	117
BDE# 190								
	4-08-3		Partial Carcass	Wet	0.0197	0.0233	406	118
BDE# 209								
	4-08-3		Partial Carcass	Wet	0.0965	0.103	133	107
BDE# 28								
	4-08-3		Partial Carcass	Wet	0.0196	0.0198	11.5	101
BDE# 47								
	4-08-3		Partial Carcass	Wet	0.0196	0.0166	0.260	84.7
BDE# 49/71								
	4-08-3		Partial Carcass	Wet	0.0195	0.0203	9.75	104
BDE# 66								
	4-08-3		Partial	Wet	0.0197	0.0206	352	105

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
			Carcass					
BDE# 85								
	4-08-3		Partial Carcass	Wet	0.0194	0.0216	101	111
BDE# 99								
	4-08-3		Partial Carcass	Wet	0.0196	0.0220	211	112
Benzo(a)anthracene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0204	105	106
DDMU								
	10-06-3		Partial Carcass	Wet	0.00790	0.00854	12.4	108
Dibenz(a,h)anthracene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0193	92.5	100.
HCB								
	10-06-3		Partial Carcass	Wet	0.00790	0.00827	11.1	105
Heptachlor								
	10-06-3		Partial Carcass	Wet	0.00790	0.00797	119	101
OCDD								
	*EQ0900410-02		Water	Wet	0.000400	0.000464		116
	*EQ0900410-02	SD	Water	Wet	0.000400	0.000439		110.
OCDF								
	*EQ0900410-02		Water	Wet	0.000400	0.000467		117
	*EQ0900410-02	SD	Water	Wet	0.000400	0.000468		117
PCB# 1								
	*EQ0900409-		Water	Wet	0.000200	0.000185		92.5

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	02							
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000193		96.5
	*EQ0900415-02		Water	Wet	0.00000100	0.000000906		90.6
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000937		93.7
PCB# 104								
	*EQ0900409-02		Water	Wet	0.000200	0.000181		90.5
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000196		98.0
	*EQ0900415-02		Water	Wet	0.00000100	0.000000940		94.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000973		97.3
PCB# 105								
	*EQ0900409-02		Water	Wet	0.000200	0.000227		114
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000227		114
	*EQ0900415-02		Water	Wet	0.00000100	0.00000102		102
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000113		113
PCB# 114								
	*EQ0900409-02		Water	Wet	0.000200	0.000188		94.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000206		103
	*EQ0900415-02		Water	Wet	0.00000100	0.000000880		88.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000964		96.4
PCB# 118								

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	*EQ0900409-02		Water	Wet	0.000200	0.000283		142
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000263		132
	*EQ0900415-02		Water	Wet	0.00000100	0.00000114		114
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000122		122
PCB# 123								
	*EQ0900409-02		Water	Wet	0.000200	0.000184		92.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000206		103
	*EQ0900415-02		Water	Wet	0.00000100	0.000000870		87.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000950		95.0
PCB# 126								
	*EQ0900409-02		Water	Wet	0.000200	0.000180		90.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000198		99.0
	*EQ0900415-02		Water	Wet	0.00000100	0.000000853		85.3
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000921		92.1
PCB# 15								
	*EQ0900409-02		Water	Wet	0.000200	0.000204		102
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000215		108
	*EQ0900415-02		Water	Wet	0.00000100	0.00000100		100.
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000107		107

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
PCB# 155								
	*EQ0900409-02		Water	Wet	0.000200	0.000100		50.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000212		106
	*EQ0900415-02		Water	Wet	0.00000100	0.000000500		50.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000500		50.0
PCB# 156/157								
	*EQ0900409-02		Water	Wet	0.000400	0.000411		103
	*EQ0900409-02	SD	Water	Wet	0.000400	0.000433		108
	*EQ0900415-02		Water	Wet	0.00000200	0.00000185		92.5
	*EQ0900415-02	SD	Water	Wet	0.00000200	0.00000202		101
PCB# 167								
	*EQ0900409-02		Water	Wet	0.000200	0.000198		99.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000212		106
	*EQ0900415-02		Water	Wet	0.00000100	0.000000932		93.2
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000102		102
PCB# 169								
	*EQ0900409-02		Water	Wet	0.000200	0.000195		97.5
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000220		110.
	*EQ0900415-02		Water	Wet	0.00000100	0.000000947		94.7
	*EQ0900415-	SD	Water	Wet	0.00000100	0.00000104		104

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	02							
PCB# 188								
	*EQ0900409-02		Water	Wet	0.000200	0.000185		92.5
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000206		103
	*EQ0900415-02		Water	Wet	0.00000100	0.000000876		87.6
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000927		92.7
PCB# 189								
	*EQ0900409-02		Water	Wet	0.000200	0.000207		104
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000225		112
	*EQ0900415-02		Water	Wet	0.00000100	0.000000965		96.5
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000104		104
PCB# 19								
	*EQ0900409-02		Water	Wet	0.000200	0.000172		86.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000192		96.0
	*EQ0900415-02		Water	Wet	0.00000100	0.000000956		95.6
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000958		95.8
PCB# 202								
	*EQ0900409-02		Water	Wet	0.000200	0.000100		50.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000221		110.
	*EQ0900415-02		Water	Wet	0.00000100	0.000000500		50.0

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000107		107
PCB# 205								
	*EQ0900409-02		Water	Wet	0.000200	0.000100		50.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000210		105
	*EQ0900415-02		Water	Wet	0.00000100	0.000000500		50.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000500		50.0
PCB# 206								
	*EQ0900409-02		Water	Wet	0.000200	0.000220		110.
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000217		108
	*EQ0900415-02		Water	Wet	0.00000100	0.000000500		50.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000105		105
PCB# 208								
	*EQ0900409-02		Water	Wet	0.000200	0.000208		104
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000220		110.
	*EQ0900415-02		Water	Wet	0.00000100	0.000000500		50.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000103		103
PCB# 209								
	*EQ0900409-02		Water	Wet	0.000200	0.000180		90.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000192		96.0
	*EQ0900415-		Water	Wet	0.00000100	0.000000930		93.0

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	02							
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000993		99.3
PCB# 3								
	*EQ0900409-02		Water	Wet	0.000200	0.000194		97.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000204		102
	*EQ0900415-02		Water	Wet	0.00000100	0.000000913		91.3
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000952		95.2
PCB# 37								
	*EQ0900409-02		Water	Wet	0.000200	0.000199		99.5
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000210		105
	*EQ0900415-02		Water	Wet	0.00000100	0.000000950		95.0
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000105		105
PCB# 4								
	*EQ0900409-02		Water	Wet	0.000200	0.000190		95.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000195		97.5
	*EQ0900415-02		Water	Wet	0.00000100	0.000000928		92.8
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000101		101
PCB# 54								
	*EQ0900409-02		Water	Wet	0.000200	0.000181		90.5
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000206		103

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	*EQ0900415-02		Water	Wet	0.00000100	0.000000881		88.1
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000970		97.0
PCB# 77								
	*EQ0900409-02		Water	Wet	0.000200	0.000198		99.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000213		106
	*EQ0900415-02		Water	Wet	0.00000100	0.000000936		93.6
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.00000101		101
PCB# 81								
	*EQ0900409-02		Water	Wet	0.000200	0.000188		94.0
	*EQ0900409-02	SD	Water	Wet	0.000200	0.000205		102
	*EQ0900415-02		Water	Wet	0.00000100	0.000000882		88.2
	*EQ0900415-02	SD	Water	Wet	0.00000100	0.000000976		97.6
Pentachlorobenzene								
	10-06-3		Partial Carcass	Wet	0.00790	0.00770	2.89	97.4
acenaphthalene								
	1-08-6		Partial Carcass	Wet	0.0189	0.0199	27.0	105
acenaphthene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0200	38.4	104
alpha BHC								
	10-06-3		Partial Carcass	Wet	0.00790	0.00622	75.6	78.7
alpha chlordane								

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	10-06-3		Partial Carcass	Wet	0.00790	0.00836	1.39	106
anthracene								
	1-08-6		Partial Carcass	Wet	0.0191	0.0218	63.7	114
benzo(a)pyrene								
	1-08-6		Partial Carcass	Wet	0.0191	0.0180	87.2	94.1
benzo(b)fluoranthene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0202	120.	105
benzo(e)pyrene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0223	83.1	116
benzo(g,h,i)perylene								
	1-08-6		Partial Carcass	Wet	0.0190	0.0160	72.9	84.4
benzo(k)fluoranthene								
	1-08-6		Partial Carcass	Wet	0.0190	0.0211	94.3	111
beta BHC								
	10-06-3		Partial Carcass	Wet	0.00790	0.00708	70.5	89.7
biphenyl								
	1-08-6		Partial Carcass	Wet	0.0190	0.0156	11.2	82.1
chlorpyrifos								
	10-06-3		Partial Carcass	Wet	0.00790	0.00644	57.0	81.5
chrysene								
	1-08-6		Partial Carcass	Wet	0.0189	0.0182	81.8	96.1
cis-nonachlor								
	10-06-3		Partial	Wet	0.00790	0.00794	3.58	101

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
			Carcass					
delta BHC								
	10-06-3		Partial Carcass	Wet	0.00790	0.00683	145	86.4
dibenzothiophene								
	1-08-6		Partial Carcass	Wet	0.0189	0.0199	114	105
dieldrin								
	10-06-3		Partial Carcass	Wet	0.00790	0.00821	1.90	104
endosulfan II								
	10-06-3		Partial Carcass	Wet	0.00790	0.00227	107	28.8
endosulfan sulfate								
	10-06-3		Partial Carcass	Wet	0.00790	0.00742	1.82	93.9
endrin								
	10-06-3		Partial Carcass	Wet	0.00790	0.00876	78.2	111
fluoranthene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0215	38.4	112
fluorene								
	1-08-6		Partial Carcass	Wet	0.0190	0.0221	13.6	116
gamma BHC								
	10-06-3		Partial Carcass	Wet	0.00790	0.00686	168	86.8
gamma chlordane								
	10-06-3		Partial Carcass	Wet	0.00790	0.00845	2.46	107
heptachlor epoxide								
	10-06-3		Partial Carcass	Wet	0.00790	0.00742	25.8	93.9

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
indeno(1,2,3-cd)pyrene								
	1-08-6		Partial Carcass	Wet	0.0189	0.0156	56.0	82.3
mirex								
	10-06-3		Partial Carcass	Wet	0.00790	0.00816	128	103
n-decane								
	1-08-6		Partial Carcass	Wet	1.92	2.24	21.3	117
n-docosane								
	1-08-6		Partial Carcass	Wet	1.88	1.76	236	93.4
n-dodecane								
	1-08-6		Partial Carcass	Wet	1.91	2.24	415	117
n-dotriacontane								
	1-08-6		Partial Carcass	Wet	1.89	1.59	260.	84.4
n-eicosane								
	1-08-6		Partial Carcass	Wet	1.92	2.01	236	105
n-heneicosane								
	1-08-6		Partial Carcass	Wet	1.92	2.08	225	108
n-hentriacontane								
	1-08-6		Partial Carcass	Wet	1.92	1.77	259	92.4
n-heptacosane								
	1-08-6		Partial Carcass	Wet	1.90	1.26	14.4	66.1
n-heptadecane								
	*1-08-6		Partial Carcass	Wet	1.92	1.56	0.110	81.3
n-hexacosane								

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
	1-08-6		Partial Carcass	Wet	1.90	1.19	3.36	62.5
n-hexadecane								
	1-08-6		Partial Carcass	Wet	1.91	1.92	3.69	100.
n-nonacosane								
	1-08-6		Partial Carcass	Wet	1.92	1.96	295	102
n-nonadecane								
	1-08-6		Partial Carcass	Wet	1.92	2.04	2.33	106
n-octacosane								
	1-08-6		Partial Carcass	Wet	1.91	1.55	273	80.9
n-octadecane								
	1-08-6		Partial Carcass	Wet	1.92	1.76	0.410	92.0
n-pentacosane								
	1-08-6		Partial Carcass	Wet	1.92	1.41	5.43	73.8
n-pentadecane								
	*1-08-6		Partial Carcass	Wet	1.90	2.02	0.660	106
n-tetracosane								
	1-08-6		Partial Carcass	Wet	1.92	2.20	14.3	115
n-tetradecane								
	1-08-6		Partial Carcass	Wet	1.91	2.12	14.0	111
n-tetratriacontane								
	1-08-6		Partial Carcass	Wet	1.91	1.75	270.	91.3
n-triacontane								
	1-08-6		Partial	Wet	1.92	1.65	279	85.8

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
			Carcass					
n-tricosane								
	*1-08-6		Partial Carcass	Wet	1.93	1.34	0.500	69.3
n-tridecane								
	1-08-6		Partial Carcass	Wet	1.91	1.80	1.75	94.2
n-tritriacontane								
	1-08-6		Partial Carcass	Wet	1.92	1.74	311	90.7
n-undecane								
	1-08-6		Partial Carcass	Wet	1.93	2.30	21.2	119
naphthalene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0211	1.11	110.
o,p'-DDD								
	10-06-3		Partial Carcass	Wet	0.00790	0.00790	13.8	100.
o,p'-DDE								
	10-06-3		Partial Carcass	Wet	0.00790	0.00757	18.8	95.8
o,p'-DDT								
	10-06-3		Partial Carcass	Wet	0.00790	0.00828	102	105
oxychlordane								
	10-06-3		Partial Carcass	Wet	0.00790	0.00724	23.8	91.7
p,p'-DDD								
	10-06-3		Partial Carcass	Wet	0.00790	0.00812	4.52	103
p,p'-DDE								
	10-06-3		Partial Carcass	Wet	0.00790	0.00797	0.420	101

Analyte	Sample Number	**	Sample Matrix	Basis	Spike Level (ppm/%)	Amount Recovered (ppm/%)	*** Spike Background	Percent Recovery
p,p'-DDT								
	10-06-3		Partial Carcass	Wet	0.00790	0.00824	11.9	104
pentachloro-anisole								
	10-06-3		Partial Carcass	Wet	0.00790	0.00853	8.04	108
perylene								
	1-08-6		Partial Carcass	Wet	0.0192	0.0194	61.2	101
phenanthrene								
	1-08-6		Partial Carcass	Wet	0.0189	0.0211	9.95	112
phytane								
	1-08-6		Partial Carcass	Wet	1.88	2.11	6.78	112
pristane								
	1-08-6		Partial Carcass	Wet	1.90	2.15	9.06	113
pyrene								
	1-08-6		Partial Carcass	Wet	0.0189	0.0210	96.7	111
trans-nonachlor								
	10-06-3		Partial Carcass	Wet	0.00790	0.00843	1.10	107

* See "Laboratory Notes" section.** SD = Spiked Duplicate Result; SB = Spike Blank Result*** For a spike to be a valid measure of method accuracy, this ratio must be higher than 1.0.

9. Laboratory Notes

Analyte	Sample Number	Result Modifier
1,2,3,4,6,7,8-HpCDD		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,4,6,7,8-HpCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,4,7,8,9-HpCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,4,7,8-HxCDD		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,4,7,8-HxCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,4-Tetrachlorobenzene		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation

	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
1,2,3,6,7,8-HxCDD		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,6,7,8-HxCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,7,8,9-HxCDD		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,7,8,9-HxCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,7,8-PeCDD		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,3,7,8-PeCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
1,2,4,5-Tetrachlorobenzene		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for

		performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
2,3,4,6,7,8-HxCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
2,3,4,7,8-PeCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
2,3,7,8-TCDD		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
2,3,7,8-TCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
Aldrin		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation

	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
BHC (Total)		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
DDMU		
	10-06-1	The analyte is < detection limit for performing the MS confirmation

	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
HCB		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation

	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
Heptachlor		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
OCDD		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
OCDF		
	EQ0900410-02	Lab reported Spike Blank Duplicate. ACF

		changed it to a Spike Duplicate.
PCB# 1		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 104		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 105		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 114		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 118		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 123		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 126		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 15		

	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 155		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 156/157		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 167		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 169		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 188		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 189		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 19		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF

		changed it to a Spike Duplicate.
PCB# 202		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 205		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 206		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 208		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 209		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 3		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 37		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 4		

	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 54		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 77		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
PCB# 81		
	EQ0900409-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
	EQ0900415-02	Lab reported Spike Blank Duplicate. ACF changed it to a Spike Duplicate.
Pentachlorobenzene		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation

	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
Total DDT's		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
alpha BHC		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation

	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
alpha chlordane		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation

	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
beta BHC		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-3	The analyte is < detection limit for performing the MS confirmation
	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
chlorpyrifos		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation

	10-06-5	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
cis-nonachlor		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation

	4-08-6	The analyte is < detection limit for performing the MS confirmation
delta BHC		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
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	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
dieldrin		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
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	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation

	10-07-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
endosulfan I		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation

endosulfan II		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
endosulfan sulfate		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
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	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
endrin		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
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	4-08-6	The analyte is < detection limit for performing the MS confirmation
gamma BHC		

	10-06-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
gamma chlordane		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
heptachlor epoxide		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
mirex		
	10-06-1	The analyte is < detection limit for performing the MS confirmation

	10-06-2	The analyte is < detection limit for performing the MS confirmation
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	10-06-4	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
n-eicosane		
	1-08-1	I
n-heptacosane		
	1-08-4	I
n-heptadecane		
	1-08-2	I
	1-08-3	I
	1-08-6	I
n-nonacosane		
	1-08-3	I
	1-08-4	I
n-nonadecane		
	1-08-3	I
n-octadecane		

	1-08-1	I
	1-08-2	I
	1-08-3	I
	1-08-4	I
	1-08-6	I
n-pentadecane		
	1-08-1	I
	1-08-2	I
	1-08-3	I
	1-08-4	I
	1-08-6	I
n-tricosane		
	1-08-1	I
	1-08-2	I
	1-08-3	I
	1-08-6	I
n-undecane		
	1-08-1	I
o,p'-DDD		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
	10-07-2	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
o,p'-DDE		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
o,p'-DDT		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation

	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
oxychlordane		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
p,p'-DDD		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
p,p'-DDE		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation

	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
p,p'-DDT		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
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	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation

	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
pentachloro-anisole		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
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	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
pristane		
	1-08-1	I
toxaphene		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
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	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
	10-07-1	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation
	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation
trans-nonachlor		
	10-06-1	The analyte is < detection limit for performing the MS confirmation
	10-06-2	The analyte is < detection limit for performing the MS confirmation
	10-06-3	The analyte is < detection limit for performing the MS confirmation
	10-06-4	The analyte is < detection limit for performing the MS confirmation
	10-06-5	The analyte is < detection limit for performing the MS confirmation
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	10-07-4	The analyte is < detection limit for performing the MS confirmation
	10-07-5	The analyte is < detection limit for performing the MS confirmation

	10-07-6	The analyte is < detection limit for performing the MS confirmation
	4-08-6	The analyte is < detection limit for performing the MS confirmation

Code List

If appropriate, labs are instructed to use the following codes when entering laboratory notes. The labs may use one or more of the codes in each note displayed above.

Code	Comment
A	Values reported based on Aldrin response factor.
C	Sample possibly compromised due to improper handling / packaging.
D	Sample was deleted from the catalog by the submitter.
H	Due to sample characteristics it was difficult to obtain adequate sample homogeneity - precision was impacted.
I	Interferences occurred during analysis.
L	Sample compromised or destroyed during shipment - sample not analyzed.
M	Compound identity was confirmed by GC/MS.
N	Sample was not analyzed.
P	Sample destroyed during preparation at lab - sample not analyzed.
Q	Insufficient sample quantity to perform requested analysis.
R	Sample is highly decomposed - results may be impacted.
S	Sample was substituted by the submitter.
T	Retention time relative to Aldrin.
U	GC/MS identifies the unknown compound to be _____ (fill in analyte).
W	Insufficient sample quantity to perform duplicate / spike analyses.
Y	Sample was analyzed but results may be impacted (see 'C')

10. QAQC Summary

1. Procedural Blank Summary

Procedural Blank Summary of Blank Equivalent Concentration (BEC) Data

Within a lab sample matrix, there must be three or more Blank results for a given analyte in order to generate a report.

10.2. Duplicate Summary

Duplicate Summary of Relative Percent Difference (RPD) Data

Within a lab sample matrix and concentration range, there must be three or more Duplicate results for a given analyte in order to generate a report.

10.3. Spike Summary

Spike Summary of Percent Recovery (PR) Data

Within a lab sample matrix, there must be three or more Spike results for a given analyte in order to generate a report.

10.4. SRM Summary

Standard Reference Material Summary of Percent Recovery (PR) Data

Within an SRM ID, there must be three or more Recoveries for a given analyte in order to generate a report.

11. QA/QC Anomalies

1. Blank Frequency Anomalies

The required number of blank sample analyses were performed with the following exceptions.				
Analyte	**Number of Samples**	**Number of Blanks**	**Frequency (%)**	**See QA/QC Note No.**
PCB-1242	13	0	0	1
PCB-1248	13	0	0	2
PCB-1254	13	0	0	3
PCB-1260	13	0	0	4

11.2. Duplicate Frequency Anomalies

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
1,2,3,4,6,7,8-HpCDD	Animal Tissue	8	0	0	5
1,2,3,4,6,7,8-HpCDF	Animal Tissue	8	0	0	6
1,2,3,4,7,8,9-HpCDF	Animal Tissue	8	0	0	7
1,2,3,4,7,8-HxCDD	Animal Tissue	8	0	0	8
1,2,3,4,7,8-HxCDF	Animal Tissue	8	0	0	9
1,2,3,6,7,8-HxCDD	Animal Tissue	8	0	0	10
1,2,3,6,7,8-HxCDF	Animal Tissue	8	0	0	11
1,2,3,7,8,9-HxCDD	Animal Tissue	8	0	0	12
1,2,3,7,8,9-HxCDF	Animal Tissue	8	0	0	13
1,2,3,7,8-PeCDD	Animal Tissue	8	0	0	14
1,2,3,7,8-PeCDF	Animal Tissue	8	0	0	15
2,3,4,6,7,8-HxCDF	Animal Tissue	8	0	0	16
2,3,4,7,8-PeCDF	Animal Tissue	8	0	0	17
2,3,7,8-TCDD	Animal Tissue	8	0	0	18
2,3,7,8-TCDF	Animal Tissue	8	0	0	19
Cl4-PCDD	Animal Tissue	8	0	0	20
Cl4-PCDF	Animal Tissue	8	0	0	21

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
Cl5-PCDD	Animal Tissue	8	0	0	22
Cl5-PCDF	Animal Tissue	8	0	0	23
Cl6-PCDD	Animal Tissue	8	0	0	24
Cl6-PCDF	Animal Tissue	8	0	0	25
Cl7-PCDD	Animal Tissue	8	0	0	26
Cl7-PCDF	Animal Tissue	8	0	0	27
OCDD	Animal Tissue	8	0	0	28
OCDF	Animal Tissue	8	0	0	29
PCB# 1	Animal Tissue	12	0	0	30
PCB# 1	Water	1	0	0	31
PCB# 10	Animal Tissue	12	0	0	32
PCB# 10	Water	1	0	0	33
PCB# 103	Animal Tissue	12	0	0	34
PCB# 103	Water	1	0	0	35
PCB# 104	Animal Tissue	12	0	0	36
PCB# 104	Water	1	0	0	37
PCB# 105	Animal Tissue	12	0	0	38
PCB# 105	Water	1	0	0	39
PCB# 106	Animal Tissue	12	0	0	40
PCB# 106	Water	1	0	0	41
PCB# 107	Animal Tissue	12	0	0	42
PCB# 107	Water	1	0	0	43
PCB# 108/124	Animal Tissue	12	0	0	44
PCB# 108/124	Water	1	0	0	45
PCB# 11	Animal Tissue	12	0	0	46
PCB# 11	Water	1	0	0	47
PCB# 110/115	Animal Tissue	12	0	0	48
PCB# 110/115	Water	1	0	0	49
PCB# 111	Animal Tissue	12	0	0	50
PCB# 111	Water	1	0	0	51
PCB# 112	Animal Tissue	12	0	0	52
PCB# 112	Water	1	0	0	53

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 114	Animal Tissue	12	0	0	54
PCB# 114	Water	1	0	0	55
PCB# 117	Animal Tissue	12	0	0	56
PCB# 117	Water	1	0	0	57
PCB# 118	Animal Tissue	12	0	0	58
PCB# 118	Water	1	0	0	59
PCB# 12/13	Animal Tissue	12	0	0	60
PCB# 12/13	Water	1	0	0	61
PCB# 120	Animal Tissue	12	0	0	62
PCB# 120	Water	1	0	0	63
PCB# 121	Animal Tissue	12	0	0	64
PCB# 121	Water	1	0	0	65
PCB# 122	Animal Tissue	12	0	0	66
PCB# 122	Water	1	0	0	67
PCB# 123	Animal Tissue	12	0	0	68
PCB# 123	Water	1	0	0	69
PCB# 126	Animal Tissue	12	0	0	70
PCB# 126	Water	1	0	0	71
PCB# 127	Animal Tissue	12	0	0	72
PCB# 127	Water	1	0	0	73
PCB# 128/166	Animal Tissue	12	0	0	74
PCB# 128/166	Water	1	0	0	75
PCB# 129/138/163	Animal Tissue	12	0	0	76
PCB# 129/138/163	Water	1	0	0	77
PCB# 130	Animal Tissue	12	0	0	78
PCB# 130	Water	1	0	0	79
PCB# 131	Animal Tissue	12	0	0	80
PCB# 131	Water	1	0	0	81
PCB# 132	Animal Tissue	12	0	0	82
PCB# 132	Water	1	0	0	83
PCB# 133	Animal Tissue	12	0	0	84
PCB# 133	Water	1	0	0	85

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 134	Animal Tissue	12	0	0	86
PCB# 134	Water	1	0	0	87
PCB# 135/151	Animal Tissue	12	0	0	88
PCB# 135/151	Water	1	0	0	89
PCB# 136	Animal Tissue	12	0	0	90
PCB# 136	Water	1	0	0	91
PCB# 137	Animal Tissue	12	0	0	92
PCB# 137	Water	1	0	0	93
PCB# 139/140	Animal Tissue	12	0	0	94
PCB# 139/140	Water	1	0	0	95
PCB# 14	Animal Tissue	12	0	0	96
PCB# 14	Water	1	0	0	97
PCB# 141	Animal Tissue	12	0	0	98
PCB# 141	Water	1	0	0	99
PCB# 142	Animal Tissue	12	0	0	100
PCB# 142	Water	1	0	0	101
PCB# 143	Animal Tissue	12	0	0	102
PCB# 143	Water	1	0	0	103
PCB# 144	Animal Tissue	12	0	0	104
PCB# 144	Water	1	0	0	105
PCB# 145	Animal Tissue	12	0	0	106
PCB# 145	Water	1	0	0	107
PCB# 146	Animal Tissue	12	0	0	108
PCB# 146	Water	1	0	0	109
PCB# 147/149	Animal Tissue	12	0	0	110
PCB# 147/149	Water	1	0	0	111
PCB# 148	Animal Tissue	12	0	0	112
PCB# 148	Water	1	0	0	113
PCB# 15	Animal Tissue	12	0	0	114
PCB# 15	Water	1	0	0	115
PCB# 150	Animal Tissue	12	0	0	116
PCB# 150	Water	1	0	0	117

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 152	Animal Tissue	12	0	0	118
PCB# 152	Water	1	0	0	119
PCB# 153/168	Animal Tissue	12	0	0	120
PCB# 153/168	Water	1	0	0	121
PCB# 154	Animal Tissue	12	0	0	122
PCB# 154	Water	1	0	0	123
PCB# 155	Animal Tissue	12	0	0	124
PCB# 155	Water	1	0	0	125
PCB# 156/157	Animal Tissue	12	0	0	126
PCB# 156/157	Water	1	0	0	127
PCB# 158	Animal Tissue	12	0	0	128
PCB# 158	Water	1	0	0	129
PCB# 159	Animal Tissue	12	0	0	130
PCB# 159	Water	1	0	0	131
PCB# 16	Animal Tissue	12	0	0	132
PCB# 16	Water	1	0	0	133
PCB# 160	Animal Tissue	12	0	0	134
PCB# 160	Water	1	0	0	135
PCB# 161	Animal Tissue	12	0	0	136
PCB# 161	Water	1	0	0	137
PCB# 162	Animal Tissue	12	0	0	138
PCB# 162	Water	1	0	0	139
PCB# 164	Animal Tissue	12	0	0	140
PCB# 164	Water	1	0	0	141
PCB# 165	Animal Tissue	12	0	0	142
PCB# 165	Water	1	0	0	143
PCB# 167	Animal Tissue	12	0	0	144
PCB# 167	Water	1	0	0	145
PCB# 169	Animal Tissue	12	0	0	146
PCB# 169	Water	1	0	0	147
PCB# 17	Animal Tissue	12	0	0	148
PCB# 17	Water	1	0	0	149

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 170	Animal Tissue	12	0	0	150
PCB# 170	Water	1	0	0	151
PCB# 171/173	Animal Tissue	12	0	0	152
PCB# 171/173	Water	1	0	0	153
PCB# 172	Animal Tissue	12	0	0	154
PCB# 172	Water	1	0	0	155
PCB# 174	Animal Tissue	12	0	0	156
PCB# 174	Water	1	0	0	157
PCB# 175	Animal Tissue	12	0	0	158
PCB# 175	Water	1	0	0	159
PCB# 176	Animal Tissue	12	0	0	160
PCB# 176	Water	1	0	0	161
PCB# 177	Animal Tissue	12	0	0	162
PCB# 177	Water	1	0	0	163
PCB# 178	Animal Tissue	12	0	0	164
PCB# 178	Water	1	0	0	165
PCB# 179	Animal Tissue	12	0	0	166
PCB# 179	Water	1	0	0	167
PCB# 18/30	Animal Tissue	12	0	0	168
PCB# 18/30	Water	1	0	0	169
PCB# 180/193	Animal Tissue	12	0	0	170
PCB# 180/193	Water	1	0	0	171
PCB# 181	Animal Tissue	12	0	0	172
PCB# 181	Water	1	0	0	173
PCB# 182	Animal Tissue	12	0	0	174
PCB# 182	Water	1	0	0	175
PCB# 183	Animal Tissue	12	0	0	176
PCB# 183	Water	1	0	0	177
PCB# 184	Animal Tissue	12	0	0	178
PCB# 184	Water	1	0	0	179
PCB# 185	Animal Tissue	12	0	0	180
PCB# 185	Water	1	0	0	181

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 186	Animal Tissue	12	0	0	182
PCB# 186	Water	1	0	0	183
PCB# 187	Animal Tissue	12	0	0	184
PCB# 187	Water	1	0	0	185
PCB# 188	Animal Tissue	12	0	0	186
PCB# 188	Water	1	0	0	187
PCB# 189	Animal Tissue	12	0	0	188
PCB# 189	Water	1	0	0	189
PCB# 19	Animal Tissue	12	0	0	190
PCB# 19	Water	1	0	0	191
PCB# 190	Animal Tissue	12	0	0	192
PCB# 190	Water	1	0	0	193
PCB# 191	Animal Tissue	12	0	0	194
PCB# 191	Water	1	0	0	195
PCB# 192	Animal Tissue	12	0	0	196
PCB# 192	Water	1	0	0	197
PCB# 194	Animal Tissue	12	0	0	198
PCB# 194	Water	1	0	0	199
PCB# 195	Animal Tissue	12	0	0	200
PCB# 195	Water	1	0	0	201
PCB# 196	Animal Tissue	12	0	0	202
PCB# 196	Water	1	0	0	203
PCB# 197	Animal Tissue	12	0	0	204
PCB# 197	Water	1	0	0	205
PCB# 198/199	Animal Tissue	12	0	0	206
PCB# 198/199	Water	1	0	0	207
PCB# 2	Animal Tissue	12	0	0	208
PCB# 2	Water	1	0	0	209
PCB# 20/28	Animal Tissue	12	0	0	210
PCB# 20/28	Water	1	0	0	211
PCB# 200	Animal Tissue	12	0	0	212
PCB# 200	Water	1	0	0	213

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 201	Animal Tissue	12	0	0	214
PCB# 201	Water	1	0	0	215
PCB# 202	Animal Tissue	12	0	0	216
PCB# 202	Water	1	0	0	217
PCB# 203	Animal Tissue	12	0	0	218
PCB# 203	Water	1	0	0	219
PCB# 204	Animal Tissue	12	0	0	220
PCB# 204	Water	1	0	0	221
PCB# 205	Animal Tissue	12	0	0	222
PCB# 205	Water	1	0	0	223
PCB# 206	Animal Tissue	12	0	0	224
PCB# 206	Water	1	0	0	225
PCB# 207	Animal Tissue	12	0	0	226
PCB# 207	Water	1	0	0	227
PCB# 208	Animal Tissue	12	0	0	228
PCB# 208	Water	1	0	0	229
PCB# 209	Animal Tissue	12	0	0	230
PCB# 209	Water	1	0	0	231
PCB# 21/33	Animal Tissue	12	0	0	232
PCB# 21/33	Water	1	0	0	233
PCB# 22	Animal Tissue	12	0	0	234
PCB# 22	Water	1	0	0	235
PCB# 23	Animal Tissue	12	0	0	236
PCB# 23	Water	1	0	0	237
PCB# 24	Animal Tissue	12	0	0	238
PCB# 24	Water	1	0	0	239
PCB# 25	Animal Tissue	12	0	0	240
PCB# 25	Water	1	0	0	241
PCB# 26/29	Animal Tissue	12	0	0	242
PCB# 26/29	Water	1	0	0	243
PCB# 27	Animal Tissue	12	0	0	244
PCB# 27	Water	1	0	0	245

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 3	Animal Tissue	12	0	0	246
PCB# 3	Water	1	0	0	247
PCB# 31	Animal Tissue	12	0	0	248
PCB# 31	Water	1	0	0	249
PCB# 32	Animal Tissue	12	0	0	250
PCB# 32	Water	1	0	0	251
PCB# 34	Animal Tissue	12	0	0	252
PCB# 34	Water	1	0	0	253
PCB# 35	Animal Tissue	12	0	0	254
PCB# 35	Water	1	0	0	255
PCB# 36	Animal Tissue	12	0	0	256
PCB# 36	Water	1	0	0	257
PCB# 37	Animal Tissue	12	0	0	258
PCB# 37	Water	1	0	0	259
PCB# 38	Animal Tissue	12	0	0	260
PCB# 38	Water	1	0	0	261
PCB# 39	Animal Tissue	12	0	0	262
PCB# 39	Water	1	0	0	263
PCB# 4	Animal Tissue	12	0	0	264
PCB# 4	Water	1	0	0	265
PCB# 40/41/70	Animal Tissue	12	0	0	266
PCB# 40/41/70	Water	1	0	0	267
PCB# 42	Animal Tissue	12	0	0	268
PCB# 42	Water	1	0	0	269
PCB# 43/73	Animal Tissue	12	0	0	270
PCB# 43/73	Water	1	0	0	271
PCB# 44/47/65	Animal Tissue	12	0	0	272
PCB# 44/47/65	Water	1	0	0	273
PCB# 45/51	Animal Tissue	12	0	0	274
PCB# 45/51	Water	1	0	0	275
PCB# 46	Animal Tissue	12	0	0	276
PCB# 46	Water	1	0	0	277

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 48	Animal Tissue	12	0	0	278
PCB# 48	Water	1	0	0	279
PCB# 49/69	Animal Tissue	12	0	0	280
PCB# 49/69	Water	1	0	0	281
PCB# 5	Animal Tissue	12	0	0	282
PCB# 5	Water	1	0	0	283
PCB# 50/53	Animal Tissue	12	0	0	284
PCB# 50/53	Water	1	0	0	285
PCB# 52	Animal Tissue	12	0	0	286
PCB# 52	Water	1	0	0	287
PCB# 54	Animal Tissue	12	0	0	288
PCB# 54	Water	1	0	0	289
PCB# 55	Animal Tissue	12	0	0	290
PCB# 55	Water	1	0	0	291
PCB# 56	Animal Tissue	12	0	0	292
PCB# 56	Water	1	0	0	293
PCB# 57	Animal Tissue	12	0	0	294
PCB# 57	Water	1	0	0	295
PCB# 58	Animal Tissue	12	0	0	296
PCB# 58	Water	1	0	0	297
PCB# 59/62/75	Animal Tissue	12	0	0	298
PCB# 59/62/75	Water	1	0	0	299
PCB# 6	Animal Tissue	12	0	0	300
PCB# 6	Water	1	0	0	301
PCB# 60	Animal Tissue	12	0	0	302
PCB# 60	Water	1	0	0	303
PCB# 63	Animal Tissue	12	0	0	304
PCB# 63	Water	1	0	0	305
PCB# 64	Animal Tissue	12	0	0	306
PCB# 64	Water	1	0	0	307
PCB# 66	Animal Tissue	12	0	0	308
PCB# 66	Water	1	0	0	309

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 67	Animal Tissue	12	0	0	310
PCB# 67	Water	1	0	0	311
PCB# 68	Animal Tissue	12	0	0	312
PCB# 68	Water	1	0	0	313
PCB# 7	Animal Tissue	12	0	0	314
PCB# 7	Water	1	0	0	315
PCB# 70/61/74/76	Animal Tissue	12	0	0	316
PCB# 70/61/74/76	Water	1	0	0	317
PCB# 72	Animal Tissue	12	0	0	318
PCB# 72	Water	1	0	0	319
PCB# 77	Animal Tissue	12	0	0	320
PCB# 77	Water	1	0	0	321
PCB# 78	Animal Tissue	12	0	0	322
PCB# 78	Water	1	0	0	323
PCB# 79	Animal Tissue	12	0	0	324
PCB# 79	Water	1	0	0	325
PCB# 8	Animal Tissue	12	0	0	326
PCB# 8	Water	1	0	0	327
PCB# 80	Animal Tissue	12	0	0	328
PCB# 80	Water	1	0	0	329
PCB# 81	Animal Tissue	12	0	0	330
PCB# 81	Water	1	0	0	331
PCB# 82	Animal Tissue	12	0	0	332
PCB# 82	Water	1	0	0	333
PCB# 83/99	Animal Tissue	12	0	0	334
PCB# 83/99	Water	1	0	0	335
PCB# 84	Animal Tissue	12	0	0	336
PCB# 84	Water	1	0	0	337
PCB# 85/116	Animal Tissue	12	0	0	338
PCB# 85/116	Water	1	0	0	339
PCB# 86/87/97/109/119/12	Animal Tissue	12	0	0	340

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
5					
PCB# 86/87/97/109/119/125	Water	1	0	0	341
PCB# 88/91	Animal Tissue	12	0	0	342
PCB# 88/91	Water	1	0	0	343
PCB# 89	Animal Tissue	12	0	0	344
PCB# 89	Water	1	0	0	345
PCB# 9	Animal Tissue	12	0	0	346
PCB# 9	Water	1	0	0	347
PCB# 90/101/113	Animal Tissue	12	0	0	348
PCB# 90/101/113	Water	1	0	0	349
PCB# 92	Animal Tissue	12	0	0	350
PCB# 92	Water	1	0	0	351
PCB# 94	Animal Tissue	12	0	0	352
PCB# 94	Water	1	0	0	353
PCB# 95	Animal Tissue	12	0	0	354
PCB# 95	Water	1	0	0	355
PCB# 96	Animal Tissue	12	0	0	356
PCB# 96	Water	1	0	0	357
PCB# 98/100	Animal Tissue	12	0	0	358
PCB# 98/100	Water	1	0	0	359
PCB# 98/102	Animal Tissue	12	0	0	360
PCB# 98/102	Water	1	0	0	361
PCB-1242	Animal Tissue	12	0	0	362
PCB-1242	Water	1	0	0	363
PCB-1248	Animal Tissue	12	0	0	364
PCB-1248	Water	1	0	0	365
PCB-1254	Animal Tissue	12	0	0	366
PCB-1254	Water	1	0	0	367
PCB-1260	Animal Tissue	12	0	0	368
PCB-1260	Water	1	0	0	369

The required number of duplicate sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Duplicates**	**Frequency (%)**	**See QA/QC Note No.**
PCB-TOTAL	Animal Tissue	12	0	0	370
PCB-TOTAL	Water	1	0	0	371

11.3. Spike Frequency Anomalies

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
1,2,3,4,6,7,8-HpCDD	Animal Tissue	8	0	0	372
1,2,3,4,6,7,8-HpCDF	Animal Tissue	8	0	0	373
1,2,3,4,7,8,9-HpCDF	Animal Tissue	8	0	0	374
1,2,3,4,7,8-HxCDD	Animal Tissue	8	0	0	375
1,2,3,4,7,8-HxCDF	Animal Tissue	8	0	0	376
1,2,3,6,7,8-HxCDD	Animal Tissue	8	0	0	377
1,2,3,6,7,8-HxCDF	Animal Tissue	8	0	0	378
1,2,3,7,8,9-HxCDD	Animal Tissue	8	0	0	379
1,2,3,7,8,9-HxCDF	Animal Tissue	8	0	0	380
1,2,3,7,8-PeCDD	Animal Tissue	8	0	0	381
1,2,3,7,8-PeCDF	Animal Tissue	8	0	0	382
2,3,4,6,7,8-HxCDF	Animal Tissue	8	0	0	383
2,3,4,7,8-PeCDF	Animal Tissue	8	0	0	384
2,3,7,8-TCDD	Animal Tissue	8	0	0	385
2,3,7,8-TCDF	Animal Tissue	8	0	0	386
BDE# 1	Animal Tissue	11	0	0	387
BDE# 10	Animal Tissue	11	0	0	388
BDE# 11	Animal Tissue	11	0	0	389
BDE# 116	Animal Tissue	11	0	0	390
BDE# 118	Animal Tissue	11	0	0	391
BDE# 119	Animal Tissue	11	0	0	392
BDE# 12	Animal Tissue	11	0	0	393

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
BDE# 126	Animal Tissue	11	0	0	394
BDE# 13	Animal Tissue	11	0	0	395
BDE# 15	Animal Tissue	11	0	0	396
BDE# 155	Animal Tissue	11	0	0	397
BDE# 166	Animal Tissue	11	0	0	398
BDE# 181	Animal Tissue	11	0	0	399
BDE# 194	Animal Tissue	11	0	0	400
BDE# 195	Animal Tissue	11	0	0	401
BDE# 196	Animal Tissue	11	0	0	402
BDE# 197	Animal Tissue	11	0	0	403
BDE# 198/199/203/200	Animal Tissue	11	0	0	404
BDE# 2	Animal Tissue	11	0	0	405
BDE# 201	Animal Tissue	11	0	0	406
BDE# 202	Animal Tissue	11	0	0	407
BDE# 204	Animal Tissue	11	0	0	408
BDE# 205	Animal Tissue	11	0	0	409
BDE# 206	Animal Tissue	11	0	0	410
BDE# 207	Animal Tissue	11	0	0	411
BDE# 208	Animal Tissue	11	0	0	412
BDE# 25	Animal Tissue	11	0	0	413
BDE# 3	Animal Tissue	11	0	0	414
BDE# 30	Animal Tissue	11	0	0	415
BDE# 32	Animal Tissue	11	0	0	416
BDE# 33	Animal Tissue	11	0	0	417
BDE# 35	Animal Tissue	11	0	0	418
BDE# 37	Animal Tissue	11	0	0	419
BDE# 7	Animal Tissue	11	0	0	420
BDE# 75	Animal Tissue	11	0	0	421
BDE# 77	Animal Tissue	11	0	0	422
BDE# 8	Animal Tissue	11	0	0	423
BDE-TOTAL	Animal Tissue	11	0	0	424

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
BHC (Total)	Animal Tissue	12	0	0	425
C1-chrysenes	Animal Tissue	5	0	0	426
C1-dibenzothiophenes	Animal Tissue	5	0	0	427
C1-Fluoranthenes & Pyrenes	Animal Tissue	5	0	0	428
C1-fluorenes	Animal Tissue	5	0	0	429
C1-naphthalenes	Animal Tissue	5	0	0	430
C1-Phenanthrenes & Anthracenes	Animal Tissue	5	0	0	431
C2-chrysenes	Animal Tissue	5	0	0	432
C2-dibenzothiophenes	Animal Tissue	5	0	0	433
C2-fluorenes	Animal Tissue	5	0	0	434
C2-naphthalenes	Animal Tissue	5	0	0	435
C2-Phenanthrenes & Anthracenes	Animal Tissue	5	0	0	436
C3-chrysenes	Animal Tissue	5	0	0	437
C3-dibenzothiophenes	Animal Tissue	5	0	0	438
C3-fluorenes	Animal Tissue	5	0	0	439
C3-naphthalenes	Animal Tissue	5	0	0	440
C3-Phenanthrenes & Anthracenes	Animal Tissue	5	0	0	441
C4-chrysenes	Animal Tissue	5	0	0	442
C4-naphthalenes	Animal Tissue	5	0	0	443
C4-Phenanthrenes & Anthracenes	Animal Tissue	5	0	0	444
Cl4-PCDD	Animal Tissue	8	0	0	445
Cl4-PCDF	Animal Tissue	8	0	0	446
Cl5-PCDD	Animal Tissue	8	0	0	447
Cl5-PCDF	Animal Tissue	8	0	0	448
Cl6-PCDD	Animal Tissue	8	0	0	449

The required number of spike sample analyses were performed with the following exceptions.

Analyte	Lab Matrix	Number of Samples	Number of Spikes	Frequency (%)	See QA/QC Note No.
Cl6-PCDF	Animal Tissue	8	0	0	450
Cl7-PCDD	Animal Tissue	8	0	0	451
Cl7-PCDF	Animal Tissue	8	0	0	452
endosulfan I	Animal Tissue	12	0	0	453
OCDD	Animal Tissue	8	0	0	454
OCDF	Animal Tissue	8	0	0	455
PCB# 1	Animal Tissue	12	0	0	456
PCB# 10	Animal Tissue	12	0	0	457
PCB# 10	Water	1	0	0	458
PCB# 103	Animal Tissue	12	0	0	459
PCB# 103	Water	1	0	0	460
PCB# 104	Animal Tissue	12	0	0	461
PCB# 105	Animal Tissue	12	0	0	462
PCB# 106	Animal Tissue	12	0	0	463
PCB# 106	Water	1	0	0	464
PCB# 107	Animal Tissue	12	0	0	465
PCB# 107	Water	1	0	0	466
PCB# 108/124	Animal Tissue	12	0	0	467
PCB# 108/124	Water	1	0	0	468
PCB# 11	Animal Tissue	12	0	0	469
PCB# 11	Water	1	0	0	470
PCB# 110/115	Animal Tissue	12	0	0	471
PCB# 110/115	Water	1	0	0	472
PCB# 111	Animal Tissue	12	0	0	473
PCB# 111	Water	1	0	0	474
PCB# 112	Animal Tissue	12	0	0	475
PCB# 112	Water	1	0	0	476
PCB# 114	Animal Tissue	12	0	0	477
PCB# 117	Animal Tissue	12	0	0	478
PCB# 117	Water	1	0	0	479
PCB# 118	Animal Tissue	12	0	0	480
PCB# 12/13	Animal Tissue	12	0	0	481

The required number of spike sample analyses were performed with the following exceptions.

Analyte	Lab Matrix	Number of Samples	Number of Spikes	Frequency (%)	See QA/QC Note No.
PCB# 12/13	Water	1	0	0	482
PCB# 120	Animal Tissue	12	0	0	483
PCB# 120	Water	1	0	0	484
PCB# 121	Animal Tissue	12	0	0	485
PCB# 121	Water	1	0	0	486
PCB# 122	Animal Tissue	12	0	0	487
PCB# 122	Water	1	0	0	488
PCB# 123	Animal Tissue	12	0	0	489
PCB# 126	Animal Tissue	12	0	0	490
PCB# 127	Animal Tissue	12	0	0	491
PCB# 127	Water	1	0	0	492
PCB# 128/166	Animal Tissue	12	0	0	493
PCB# 128/166	Water	1	0	0	494
PCB# 129/138/163	Animal Tissue	12	0	0	495
PCB# 129/138/163	Water	1	0	0	496
PCB# 130	Animal Tissue	12	0	0	497
PCB# 130	Water	1	0	0	498
PCB# 131	Animal Tissue	12	0	0	499
PCB# 131	Water	1	0	0	500
PCB# 132	Animal Tissue	12	0	0	501
PCB# 132	Water	1	0	0	502
PCB# 133	Animal Tissue	12	0	0	503
PCB# 133	Water	1	0	0	504
PCB# 134	Animal Tissue	12	0	0	505
PCB# 134	Water	1	0	0	506
PCB# 135/151	Animal Tissue	12	0	0	507
PCB# 135/151	Water	1	0	0	508
PCB# 136	Animal Tissue	12	0	0	509
PCB# 136	Water	1	0	0	510
PCB# 137	Animal Tissue	12	0	0	511
PCB# 137	Water	1	0	0	512
PCB# 139/140	Animal Tissue	12	0	0	513

The required number of spike sample analyses were performed with the following exceptions.

Analyte	Lab Matrix	Number of Samples	Number of Spikes	Frequency (%)	See QA/QC Note No.
PCB# 139/140	Water	1	0	0	514
PCB# 14	Animal Tissue	12	0	0	515
PCB# 14	Water	1	0	0	516
PCB# 141	Animal Tissue	12	0	0	517
PCB# 141	Water	1	0	0	518
PCB# 142	Animal Tissue	12	0	0	519
PCB# 142	Water	1	0	0	520
PCB# 143	Animal Tissue	12	0	0	521
PCB# 143	Water	1	0	0	522
PCB# 144	Animal Tissue	12	0	0	523
PCB# 144	Water	1	0	0	524
PCB# 145	Animal Tissue	12	0	0	525
PCB# 145	Water	1	0	0	526
PCB# 146	Animal Tissue	12	0	0	527
PCB# 146	Water	1	0	0	528
PCB# 147/149	Animal Tissue	12	0	0	529
PCB# 147/149	Water	1	0	0	530
PCB# 148	Animal Tissue	12	0	0	531
PCB# 148	Water	1	0	0	532
PCB# 15	Animal Tissue	12	0	0	533
PCB# 150	Animal Tissue	12	0	0	534
PCB# 150	Water	1	0	0	535
PCB# 152	Animal Tissue	12	0	0	536
PCB# 152	Water	1	0	0	537
PCB# 153/168	Animal Tissue	12	0	0	538
PCB# 153/168	Water	1	0	0	539
PCB# 154	Animal Tissue	12	0	0	540
PCB# 154	Water	1	0	0	541
PCB# 155	Animal Tissue	12	0	0	542
PCB# 156/157	Animal Tissue	12	0	0	543
PCB# 158	Animal Tissue	12	0	0	544
PCB# 158	Water	1	0	0	545

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 159	Animal Tissue	12	0	0	546
PCB# 159	Water	1	0	0	547
PCB# 16	Animal Tissue	12	0	0	548
PCB# 16	Water	1	0	0	549
PCB# 160	Animal Tissue	12	0	0	550
PCB# 160	Water	1	0	0	551
PCB# 161	Animal Tissue	12	0	0	552
PCB# 161	Water	1	0	0	553
PCB# 162	Animal Tissue	12	0	0	554
PCB# 162	Water	1	0	0	555
PCB# 164	Animal Tissue	12	0	0	556
PCB# 164	Water	1	0	0	557
PCB# 165	Animal Tissue	12	0	0	558
PCB# 165	Water	1	0	0	559
PCB# 167	Animal Tissue	12	0	0	560
PCB# 169	Animal Tissue	12	0	0	561
PCB# 17	Animal Tissue	12	0	0	562
PCB# 17	Water	1	0	0	563
PCB# 170	Animal Tissue	12	0	0	564
PCB# 170	Water	1	0	0	565
PCB# 171/173	Animal Tissue	12	0	0	566
PCB# 171/173	Water	1	0	0	567
PCB# 172	Animal Tissue	12	0	0	568
PCB# 172	Water	1	0	0	569
PCB# 174	Animal Tissue	12	0	0	570
PCB# 174	Water	1	0	0	571
PCB# 175	Animal Tissue	12	0	0	572
PCB# 175	Water	1	0	0	573
PCB# 176	Animal Tissue	12	0	0	574
PCB# 176	Water	1	0	0	575
PCB# 177	Animal Tissue	12	0	0	576
PCB# 177	Water	1	0	0	577

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 178	Animal Tissue	12	0	0	578
PCB# 178	Water	1	0	0	579
PCB# 179	Animal Tissue	12	0	0	580
PCB# 179	Water	1	0	0	581
PCB# 18/30	Animal Tissue	12	0	0	582
PCB# 18/30	Water	1	0	0	583
PCB# 180/193	Animal Tissue	12	0	0	584
PCB# 180/193	Water	1	0	0	585
PCB# 181	Animal Tissue	12	0	0	586
PCB# 181	Water	1	0	0	587
PCB# 182	Animal Tissue	12	0	0	588
PCB# 182	Water	1	0	0	589
PCB# 183	Animal Tissue	12	0	0	590
PCB# 183	Water	1	0	0	591
PCB# 184	Animal Tissue	12	0	0	592
PCB# 184	Water	1	0	0	593
PCB# 185	Animal Tissue	12	0	0	594
PCB# 185	Water	1	0	0	595
PCB# 186	Animal Tissue	12	0	0	596
PCB# 186	Water	1	0	0	597
PCB# 187	Animal Tissue	12	0	0	598
PCB# 187	Water	1	0	0	599
PCB# 188	Animal Tissue	12	0	0	600
PCB# 189	Animal Tissue	12	0	0	601
PCB# 19	Animal Tissue	12	0	0	602
PCB# 190	Animal Tissue	12	0	0	603
PCB# 190	Water	1	0	0	604
PCB# 191	Animal Tissue	12	0	0	605
PCB# 191	Water	1	0	0	606
PCB# 192	Animal Tissue	12	0	0	607
PCB# 192	Water	1	0	0	608
PCB# 194	Animal Tissue	12	0	0	609

The required number of spike sample analyses were performed with the following exceptions.

Analyte	Lab Matrix	Number of Samples	Number of Spikes	Frequency (%)	See QA/QC Note No.
PCB# 194	Water	1	0	0	610
PCB# 195	Animal Tissue	12	0	0	611
PCB# 195	Water	1	0	0	612
PCB# 196	Animal Tissue	12	0	0	613
PCB# 196	Water	1	0	0	614
PCB# 197	Animal Tissue	12	0	0	615
PCB# 197	Water	1	0	0	616
PCB# 198/199	Animal Tissue	12	0	0	617
PCB# 198/199	Water	1	0	0	618
PCB# 2	Animal Tissue	12	0	0	619
PCB# 2	Water	1	0	0	620
PCB# 20/28	Animal Tissue	12	0	0	621
PCB# 20/28	Water	1	0	0	622
PCB# 200	Animal Tissue	12	0	0	623
PCB# 200	Water	1	0	0	624
PCB# 201	Animal Tissue	12	0	0	625
PCB# 201	Water	1	0	0	626
PCB# 202	Animal Tissue	12	0	0	627
PCB# 203	Animal Tissue	12	0	0	628
PCB# 203	Water	1	0	0	629
PCB# 204	Animal Tissue	12	0	0	630
PCB# 204	Water	1	0	0	631
PCB# 205	Animal Tissue	12	0	0	632
PCB# 206	Animal Tissue	12	0	0	633
PCB# 207	Animal Tissue	12	0	0	634
PCB# 207	Water	1	0	0	635
PCB# 208	Animal Tissue	12	0	0	636
PCB# 209	Animal Tissue	12	0	0	637
PCB# 21/33	Animal Tissue	12	0	0	638
PCB# 21/33	Water	1	0	0	639
PCB# 22	Animal Tissue	12	0	0	640
PCB# 22	Water	1	0	0	641

The required number of spike sample analyses were performed with the following exceptions.

Analyte	Lab Matrix	Number of Samples	Number of Spikes	Frequency (%)	See QA/QC Note No.
PCB# 23	Animal Tissue	12	0	0	642
PCB# 23	Water	1	0	0	643
PCB# 24	Animal Tissue	12	0	0	644
PCB# 24	Water	1	0	0	645
PCB# 25	Animal Tissue	12	0	0	646
PCB# 25	Water	1	0	0	647
PCB# 26/29	Animal Tissue	12	0	0	648
PCB# 26/29	Water	1	0	0	649
PCB# 27	Animal Tissue	12	0	0	650
PCB# 27	Water	1	0	0	651
PCB# 3	Animal Tissue	12	0	0	652
PCB# 31	Animal Tissue	12	0	0	653
PCB# 31	Water	1	0	0	654
PCB# 32	Animal Tissue	12	0	0	655
PCB# 32	Water	1	0	0	656
PCB# 34	Animal Tissue	12	0	0	657
PCB# 34	Water	1	0	0	658
PCB# 35	Animal Tissue	12	0	0	659
PCB# 35	Water	1	0	0	660
PCB# 36	Animal Tissue	12	0	0	661
PCB# 36	Water	1	0	0	662
PCB# 37	Animal Tissue	12	0	0	663
PCB# 38	Animal Tissue	12	0	0	664
PCB# 38	Water	1	0	0	665
PCB# 39	Animal Tissue	12	0	0	666
PCB# 39	Water	1	0	0	667
PCB# 4	Animal Tissue	12	0	0	668
PCB# 40/41/70	Animal Tissue	12	0	0	669
PCB# 40/41/70	Water	1	0	0	670
PCB# 42	Animal Tissue	12	0	0	671
PCB# 42	Water	1	0	0	672
PCB# 43/73	Animal Tissue	12	0	0	673

The required number of spike sample analyses were performed with the following exceptions.

Analyte	Lab Matrix	Number of Samples	Number of Spikes	Frequency (%)	See QA/QC Note No.
PCB# 43/73	Water	1	0	0	674
PCB# 44/47/65	Animal Tissue	12	0	0	675
PCB# 44/47/65	Water	1	0	0	676
PCB# 45/51	Animal Tissue	12	0	0	677
PCB# 45/51	Water	1	0	0	678
PCB# 46	Animal Tissue	12	0	0	679
PCB# 46	Water	1	0	0	680
PCB# 48	Animal Tissue	12	0	0	681
PCB# 48	Water	1	0	0	682
PCB# 49/69	Animal Tissue	12	0	0	683
PCB# 49/69	Water	1	0	0	684
PCB# 5	Animal Tissue	12	0	0	685
PCB# 5	Water	1	0	0	686
PCB# 50/53	Animal Tissue	12	0	0	687
PCB# 50/53	Water	1	0	0	688
PCB# 52	Animal Tissue	12	0	0	689
PCB# 52	Water	1	0	0	690
PCB# 54	Animal Tissue	12	0	0	691
PCB# 55	Animal Tissue	12	0	0	692
PCB# 55	Water	1	0	0	693
PCB# 56	Animal Tissue	12	0	0	694
PCB# 56	Water	1	0	0	695
PCB# 57	Animal Tissue	12	0	0	696
PCB# 57	Water	1	0	0	697
PCB# 58	Animal Tissue	12	0	0	698
PCB# 58	Water	1	0	0	699
PCB# 59/62/75	Animal Tissue	12	0	0	700
PCB# 59/62/75	Water	1	0	0	701
PCB# 6	Animal Tissue	12	0	0	702
PCB# 6	Water	1	0	0	703
PCB# 60	Animal Tissue	12	0	0	704
PCB# 60	Water	1	0	0	705

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 63	Animal Tissue	12	0	0	706
PCB# 63	Water	1	0	0	707
PCB# 64	Animal Tissue	12	0	0	708
PCB# 64	Water	1	0	0	709
PCB# 66	Animal Tissue	12	0	0	710
PCB# 66	Water	1	0	0	711
PCB# 67	Animal Tissue	12	0	0	712
PCB# 67	Water	1	0	0	713
PCB# 68	Animal Tissue	12	0	0	714
PCB# 68	Water	1	0	0	715
PCB# 7	Animal Tissue	12	0	0	716
PCB# 7	Water	1	0	0	717
PCB# 70/61/74/76	Animal Tissue	12	0	0	718
PCB# 70/61/74/76	Water	1	0	0	719
PCB# 72	Animal Tissue	12	0	0	720
PCB# 72	Water	1	0	0	721
PCB# 77	Animal Tissue	12	0	0	722
PCB# 78	Animal Tissue	12	0	0	723
PCB# 78	Water	1	0	0	724
PCB# 79	Animal Tissue	12	0	0	725
PCB# 79	Water	1	0	0	726
PCB# 8	Animal Tissue	12	0	0	727
PCB# 8	Water	1	0	0	728
PCB# 80	Animal Tissue	12	0	0	729
PCB# 80	Water	1	0	0	730
PCB# 81	Animal Tissue	12	0	0	731
PCB# 82	Animal Tissue	12	0	0	732
PCB# 82	Water	1	0	0	733
PCB# 83/99	Animal Tissue	12	0	0	734
PCB# 83/99	Water	1	0	0	735
PCB# 84	Animal Tissue	12	0	0	736
PCB# 84	Water	1	0	0	737

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
PCB# 85/116	Animal Tissue	12	0	0	738
PCB# 85/116	Water	1	0	0	739
PCB# 86/87/97/109/119/125	Animal Tissue	12	0	0	740
PCB# 86/87/97/109/119/125	Water	1	0	0	741
PCB# 88/91	Animal Tissue	12	0	0	742
PCB# 88/91	Water	1	0	0	743
PCB# 89	Animal Tissue	12	0	0	744
PCB# 89	Water	1	0	0	745
PCB# 9	Animal Tissue	12	0	0	746
PCB# 9	Water	1	0	0	747
PCB# 90/101/113	Animal Tissue	12	0	0	748
PCB# 90/101/113	Water	1	0	0	749
PCB# 92	Animal Tissue	12	0	0	750
PCB# 92	Water	1	0	0	751
PCB# 94	Animal Tissue	12	0	0	752
PCB# 94	Water	1	0	0	753
PCB# 95	Animal Tissue	12	0	0	754
PCB# 95	Water	1	0	0	755
PCB# 96	Animal Tissue	12	0	0	756
PCB# 96	Water	1	0	0	757
PCB# 98/100	Animal Tissue	12	0	0	758
PCB# 98/100	Water	1	0	0	759
PCB# 98/102	Animal Tissue	12	0	0	760
PCB# 98/102	Water	1	0	0	761
PCB-1242	Animal Tissue	12	0	0	762
PCB-1242	Water	1	0	0	763
PCB-1248	Animal Tissue	12	0	0	764
PCB-1248	Water	1	0	0	765

The required number of spike sample analyses were performed with the following exceptions.					
Analyte	**Lab Matrix**	**Number of Samples**	**Number of Spikes**	**Frequency (%)**	**See QA/QC Note No.**
PCB-1254	Animal Tissue	12	0	0	766
PCB-1254	Water	1	0	0	767
PCB-1260	Animal Tissue	12	0	0	768
PCB-1260	Water	1	0	0	769
PCB-TOTAL	Animal Tissue	12	0	0	770
PCB-TOTAL	Water	1	0	0	771
Total DDT's	Animal Tissue	12	0	0	772
toxaphene	Animal Tissue	12	0	0	773
1,2,3,4,6,7,8-HpCDD	Water	0	1	0	774
1,2,3,4,6,7,8-HpCDF	Water	0	1	0	775
1,2,3,4,7,8,9-HpCDF	Water	0	1	0	776
1,2,3,4,7,8-HxCDD	Water	0	1	0	777
1,2,3,4,7,8-HxCDF	Water	0	1	0	778
1,2,3,6,7,8-HxCDD	Water	0	1	0	779
1,2,3,6,7,8-HxCDF	Water	0	1	0	780
1,2,3,7,8,9-HxCDD	Water	0	1	0	781
1,2,3,7,8,9-HxCDF	Water	0	1	0	782
1,2,3,7,8-PeCDD	Water	0	1	0	783
1,2,3,7,8-PeCDF	Water	0	1	0	784
2,3,4,6,7,8-HxCDF	Water	0	1	0	785
2,3,4,7,8-PeCDF	Water	0	1	0	786
2,3,7,8-TCDD	Water	0	1	0	787
2,3,7,8-TCDF	Water	0	1	0	788
OCDD	Water	0	1	0	789
OCDF	Water	0	1	0	790

11.4. Reference Material Frequency Anomalies

No Standard Reference Material data exists in this set of results; therefore, the anomaly test was not performed.

11.5. Mass Spec Frequency Anomalies

The required number of mass spec confirmations were performed with the following exceptions.				
Lab Matrix	**Number of Analytes**	**Number of Confirmations**	**Frequency (%)**	**See QA/QC Note No.**
Animal Tissue	94	0	0	791

11.6. Limit of Detection Anomalies

Limits of Detection were within the contract requirements with the following exceptions.							
Analyte	**Sample Number**	**Lab Matrix**	*** CRDL (ppm/%)**	**Basis**	**Acceptable To (ppm/%)**	**LOD (ppm/%)**	**See QA/QC Note No.**
BDE# 209	4-08-1	Animal Tissue	0.0001	Wet	0.000300	0.00147	792
BDE# 209	7-06-6	Animal Tissue	0.0001	Wet	0.000300	0.00137	793
BDE# 209	4-08-2	Animal Tissue	0.0001	Wet	0.000300	0.00144	794
BDE# 209	7-06-1	Animal Tissue	0.0001	Wet	0.000300	0.00145	795
BDE# 209	7-06-2	Animal Tissue	0.0001	Wet	0.000300	0.00146	796
BDE# 209	7-06-3	Animal Tissue	0.0001	Wet	0.000300	0.00147	797
BDE# 209	4-08-6	Animal Tissue	0.0001	Wet	0.000300	0.00143	798
BDE# 209	4-08-4	Animal Tissue	0.0001	Wet	0.000300	0.00149	799
BDE# 209	7-06-4	Animal Tissue	0.0001	Wet	0.000300	0.00141	800
BDE# 209	7-06-5	Animal Tissue	0.0001	Wet	0.000300	0.00142	801
BDE# 209	4-08-3	Animal Tissue	0.0001	Wet	0.000300	0.00145	802
BDE# 85	7-06-6	Animal Tissue	0.0001	Wet	0.000300	0.000364	803
BDE# 85	7-06-1	Animal Tissue	0.0001	Wet	0.000300	0.000384	804
BDE# 85	4-08-6	Animal Tissue	0.0001	Wet	0.000300	0.000380	805
BDE# 85	7-06-5	Animal Tissue	0.0001	Wet	0.000300	0.000377	806
BDE# 85	4-08-2	Animal Tissue	0.0001	Wet	0.000300	0.000383	807
BDE# 85	4-08-3	Animal Tissue	0.0001	Wet	0.000300	0.000384	808
BDE# 85	4-08-1	Animal Tissue	0.0001	Wet	0.000300	0.000391	809
BDE# 85	7-06-2	Animal Tissue	0.0001	Wet	0.000300	0.000387	810

* CRDL = Contract Required Detection Limit.

11.7. Blank Anomalies

Procedural Blank analyses were acceptable with the following exceptions.					
Analyte	**Lab Matrix**	*** BEC (ppm/%)**	**LOD (ppm/%)**	**Basis**	**See QA/QC Note No.**
PCB# 11	Water	0.000581	0.00004	Wet	811
PCB# 11	Water	8.32E-7	2E-7	Wet	812
PCB-TOTAL	Water	0.0024	0.0002	Wet	813
PCB-TOTAL	Water	0.00000386	0.000001	Wet	814

* BEC = Blank Equivalent Concentration.

11.8. Duplicate Anomalies

All duplicate results were within normal limits.

11.9. Spike Anomalies

All spike results were within normal limits with the following exceptions.									
Analyte	**Sample Number**	**Lab Matrix**	**Sample Result ppm/%**	**LOD ppm/%**	**Spike Result ppm/%**	**Spike Level ppm/%**	**% Recovery**	**Spike / Background**	**See QA/QC Note No.**
endosulfan II	10-06-3	Animal Tissue	< 0.000148	0.000148	0.00235	0.00790	29.7	53.4	815
n-heptacosane	1-08-6	Animal Tissue	0.132	0.0140	1.39	1.90	66.1	14.4	816
n-pentacosane	1-08-6	Animal Tissue	0.353	0.0162	1.77	1.92	73.8	5.43	817

11.10. S.R.M. Anomalies

No SRM data exists in this set of results; therefore, the anomaly test was not performed.

11.11. QA/QC Notes

QA/QC Note Number and Comments
Additional TDI Comments: 06385810 (Water) Insufficient sample for analysis. per Juan. ACF Comments: 1-4 Other PCB blank analyses were performed. This is acceptable. 5-371 A blank spike duplicate analyses was performed. This is acceptable. 372-790 A Blank spike analyses was performed. This is acceptable 791 Analyte detectable levels were to low for mass spec confirmation. 792-810 These LODs were higher than the ECDMS default. They are acceptable. 811-814 These analytes were reported present in the blank. Results that are less than ten times the amount found in the blank are considered non-detects. 815-817 The spike recoveries for these analytes were low. Results may be Biased low. QA/QC reviewed 04/06/2010 Walter Riley

12. Analytical Methods

Below are the analytical methods used by TDI to produce the results included in this report.

Method Codes:	001

Lab Matrix	Analyte
Animal Tissue	% Lipid
	% Moisture

Method Code: 001

LABORATORY: TDI Brooks International, Inc.

Tissue Extraction Method (PAH and OCs)

Tissue samples are either immediately processed or stored frozen (-20°C) until processing. Tissues are processed as appropriate for the tissue type (e.g. dissection, shucking). Processed tissues are homogenized using a variety of mechanical methods (Waring blender, Hobart meat grinder or Tissumisser), depending upon the tissue amount and type. After homogenization, an approximate 1 g aliquot is removed and dried in an oven at 105°C to a constant weight to determine % moisture. The remaining samples are stored in certified pre-cleaned jars frozen (- 20°C) until analysis. Prior to extraction, tissue samples may be lypholized or a wet aliquot is chemically dried using a dessicant such as Hydromatrix or sodium sulfate. Samples are then extracted using a Dionex ASE200 Accelerated Solvent Extractor (ASE). The dried sample or the sample and dessicant material is loaded into 22 or 33 mL stainless steel ASE extraction tubes. The extraction are performed using 100% dicholormethane at 100°C and 2000 psi. The extracted organics dissolved in the solvent are collected in 60 mL glass vials. The extract is concentrated to approximately 10 mL in the collection vials and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60°C. If lipid weight is required, a 100 mL aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic material (primarily lipids) must be removed prior to instrument analyses.

The extract is processed through silica gel/alumina chromatography columns and High Performance Liquid Chromatography (HPLC). The remaining 2.9 mL of sample extracted are loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50°C. The samples is transferred into 4 mL amber vials. Extracts subsequently processed by HPLC to further remove lipid interferences. Lipid removal is accomplished by flushing samples with dichlormethane through size exclusion Phenogel 10 m GPC 100 A columns. Approximately 40 mL is collected using a fraction collector, which is concentrated to 0.5 mL using a water bath at 55-60C. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography is required to separate PCBs from toxaphene/pesticides when toxphene analysis is required and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract prior to HPLC clean up is processed through a 3% deactivated silica gel column. The column is packed in dicloromethane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60°C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42°C and 20 psi. The sample is ready for instrument analysis.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Environmental Protection Agency, "Method 3545: Pressurized Fluid Extraction (PFE)," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997)

Zuloaga, O.; Etxebarria, N.; Fernandez L. A.;Madariaga, J.M.; Optimization and comparison of MAE, ASE and Soxhlet extraction for the determination of HCH isomers in soil samples. Fresenius J Anal Chem, 2000, 367, 733-737.

Schantz, M.; Nichols, J. J.; Wise, S. A.; Evaluation of Pressurized Fluid Extraction for the Extraction of Environmental Matrix Reference Material, Anal. Chem., 1997, 69, 4210-4219.

Method Codes:	001 002

Lab Matrix	Analyte
Animal Tissue	1,6,7-Trimethyl-naphthalene
	1-methylnaphthalene

1-methylphenanthrene
2,6-dimethylnaphthalene
2-methylnaphthalene
Benzo(a)anthracene
C1-Fluoranthenes & Pyrenes
C1-Phenanthrenes & Anthracenes
C1-chrysenes
C1-dibenzothiophenes
C1-fluorenes
C1-naphthalenes
C2-Phenanthrenes & Anthracenes
C2-chrysenes
C2-dibenzothiophenes
C2-fluorenes
C2-naphthalenes
C3-Phenanthrenes & Anthracenes
C3-chrysenes
C3-dibenzothiophenes
C3-fluorenes
C3-naphthalenes
C4-Phenanthrenes & Anthracenes
C4-chrysenes
C4-naphthalenes
Dibenz(a,h)anthracene
acenaphthalene
acenaphthene
anthracene
benzo(a)pyrene
benzo(b)fluoranthene
benzo(e)pyrene
benzo(g,h,i)perylene
benzo(k)fluoranthene
biphenyl
chrysene

dibenzothiophene
fluoranthene
fluorene
indeno(1,2,3-cd)pyrene
naphthalene
perylene
phenanthrene
pyrene

Method Code: 001

LABORATORY: TDI Brooks International, Inc.

Tissue Extraction Method (PAH and OCs)

Tissue samples are either immediately processed or stored frozen (-20°C) until processing. Tissues are processed as appropriate for the tissue type (e.g. dissection, shucking). Processed tissues are homogenized using a variety of mechanical methods (Waring blender, Hobart meat grinder or Tissumisser), depending upon the tissue amount and type. After homogenization, an approximate 1 g aliquot is removed and dried in an oven at 105°C to a constant weight to determine % moisture. The remaining samples are stored in certified pre-cleaned jars frozen (- 20°C) until analysis. Prior to extraction, tissue samples may be lypholized or a wet aliquot is chemically dried using a dessicant such as Hydromatrix or sodium sulfate. Samples are then extracted using a Dionex ASE200 Accelerated Solvent Extractor (ASE). The dried sample or the sample and dessicant material is loaded into 22 or 33 mL stainless steel ASE extraction tubes. The extraction are performed using 100% dicholormethane at 100°C and 2000 psi. The extracted organics dissolved in the solvent are collected in 60 mL glass vials. The extract is concentrated to approximately 10 mL in the collection vials and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60°C. If lipid weight is required, a 100 mL aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic material (primarily lipids) must be removed prior to instrument analyses.

The extract is processed through silica gel/alumina chromatography columns and High Performance Liquid Chromatography (HPLC). The remaining 2.9 mL of sample extracted are loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50°C. The samples is transferred into 4 mL amber vials. Extracts subsequently processed by HPLC to further remove lipid interferences. Lipid removal is accomplished by flushing samples with dichlormethane through size exclusion Phenogel 10 m GPC 100 A columns. Approximately 40 mL is collected using a fraction collector, which is concentrated to 0.5 mL using a water bath at 55-60C. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography is required to separate PCBs from toxaphene/pesticides when toxphene analysis is required

and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract prior to HPLC clean up is processed through a 3% deactivated silica gel column. The column is packed in dicloromethane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60°C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42°C and 20 psi. The sample is ready for instrument analysis.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Environmental Protection Agency, "Method 3545: Pressurized Fluid Extraction (PFE)," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997)

Zuloaga, O.; Etxebarria, N.; Fernandez L. A.;Madariaga, J.M.; Optimization and comparison of MAE, ASE and Soxhlet extraction for the determination of HCH isomers in soil samples. Fresenius J Anal Chem, 2000, 367, 733-737.

Schantz, M.; Nichols, J. J.; Wise, S. A.; Evaluation of Pressurized Fluid Extraction for the Extraction of Environmental Matrix Reference Material, Anal. Chem., 1997, 69, 4210-4219.

Method Code: 002

LABORATORY: TDI Brooks International, Inc.

Aromatic Hydrocarbon Determination by Selected Ion Monitoring (SIM) Gas Chromatography/ Mass Spectrometry (GC/MS)

Polycyclic aromatic hydrocarbons (PAH) and their alkylated homologues are analyzed in sample extracts by a HewlettPackard, model 5890 GS and model 5972 MS operated in SIM using a capillary column. The GC is operated in splitless mode and the capillary column is an Agilent Technologies HP-5MS (60 m x 0.25 mm ID and 0.25 mm film thickness). The carrier gas is helium at

a flow rate of 1 mL/minute. The temperature of the injection port is 300°C and transfer line is 290°C. The initial oven temperature is 60°C, the ramp rate is 7°C/minutes to a final oven temperature of 310°C and held for 20 minutes. For analyte identification, the extracted ion current profiles of the primary m/z and the confirmatory ion for each analyte must be at a maximum in the same scan or within one scan of each other and the retention time must fall with 5 seconds of the retention time of the authentic standard or alkyl homologue grouping. The pattern of alkylated PAH homologue groupings is established by analysis of reference oil standards. The relative peak heights of the primary mass ion compared to the confirmation or secondary mass ion must fall within 30 % of the relative intensities of these masses in a reference mass spectrum.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Method Codes:	001 003

Lab Matrix	Analyte
Animal Tissue	1,2,3,4-Tetrachlorobenzene
	1,2,4,5-Tetrachlorobenzene
	Aldrin
	BHC (Total)
	DDMU
	HCB
	Heptachlor
	Pentachlorobenzene
	Total DDT's
	alpha BHC
	alpha chlordane
	beta BHC
	chlorpyrifos
	cis-nonachlor
	delta BHC

dieldrin
endosulfan I
endosulfan II
endosulfan sulfate
endrin
gamma BHC
gamma chlordane
heptachlor epoxide
mirex
o,p'-DDD
o,p'-DDE
o,p'-DDT
oxychlordane
p,p'-DDD
p,p'-DDE
p,p'-DDT
pentachloro-anisole
toxaphene
trans-nonachlor

Method Code: 001

LABORATORY: TDI Brooks International, Inc.

Tissue Extraction Method (PAH and OCs)

Tissue samples are either immediately processed or stored frozen (-20°C) until processing. Tissues are processed as appropriate for the tissue type (e.g. dissection, shucking). Processed tissues are homogenized using a variety of mechanical methods (Waring blender, Hobart meat grinder or Tissumisser), depending upon the tissue amount and type. After homogenization, an approximate 1 g aliquot is removed and dried in an oven at 105°C to a constant weight to determine % moisture. The remaining samples are stored in certified pre-cleaned jars frozen (- 20°C) until analysis. Prior to extraction, tissue samples may be lypholized or a wet aliquot is chemically dried using a dessicant such as Hydromatrix or sodium sulfate. Samples are then extracted using a Dionex ASE200 Accelerated Solvent Extractor (ASE). The dried sample or the sample and dessicant material is loaded into 22 or 33 mL stainless steel ASE extraction tubes. The extraction are performed using 100% dicholormethane at 100°C and 2000 psi. The extracted organics dissolved in the solvent are collected in 60 mL glass vials. The extract is concentrated to approximately 10 mL in the collection vials and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60°C. If lipid weight is required, a 100 mL aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic material (primarily lipids) must be removed prior to instrument analyses.

The extract is processed through silica gel/alumina chromatography columns and High Performance Liquid Chromatography (HPLC). The remaining 2.9 mL of sample extracted are loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50°C. The samples is transferred into 4 mL amber vials. Extracts subsequently processed by HPLC to further remove lipid interferences. Lipid removal is accomplished by flushing samples with dichlormethane through size exclusion Phenogel 10 m GPC 100 A columns. Approximately 40 mL is collected using a fraction collector, which is concentrated to 0.5 mL using a water bath at 55-60C. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography is required to separate PCBs from toxaphene/pesticides when toxphene analysis is required and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract prior to HPLC clean up is processed through a 3% deactivated silica gel column. The column is packed in dicloromethane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60°C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42°C and 20 psi. The sample is ready for instrument analysis.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Environmental Protection Agency, "Method 3545: Pressurized Fluid Extraction (PFE)," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997)

Zuloaga, O.; Etxebarria, N.; Fernandez L. A.;Madariaga, J.M.; Optimization and comparison of MAE, ASE and Soxhlet extraction for the determination of HCH isomers in soil samples. Fresenius J Anal Chem, 2000, 367, 733-737.

Schantz, M.; Nichols, J. J.; Wise, S. A.; Evaluation of Pressurized Fluid Extraction for the Extraction of Environmental Matrix Reference Material, Anal. Chem., 1997, 69, 4210-4219.

Method Code: 003

LABORATORY: TDI Brooks International, Inc.

Chlorinated Hydrocarbon Determination by Gas Chromatography/Electron Capture Detection (GC/ECD)

Chlorinated hydrocarbons are determined in samples by GC/ECD. Samples are extracted as previously described and analyzed on a HewletPackard (HP), model 5890 GC equipped with an ECD. Between 1 to 5 mL of sample is injected using an HP, model 7673A autosampler. The instrument is set up with dual columns. The primary capillary column is a J&W DB-5 (30 m x 24 mm ID and 0.25 mm film thickness). The second column, a confirmation column, is a J&W DB-17HT (30 m x 0.25 mm ID and 0.15 mm film thickness). The inlet system is splitless and the carrier gas is helium at a flow rate of 1 mL/min. For the analysis of standard halogenated hydrocarbons, the temperature of the injection port is 275°C and the detector is 325°C. The initial oven temperature is 100°C with a hold time of 1 minute. The ramp rate is 5°C/minute to 140°C with a hold time of 1 minute, followed by a ramp rate of 1.5°C /minute to 250C with a hold time of 1 minute and finally a ramp rate of 10°C/minutes to 300°C with a final hold time of 5 minutes. For planar PCBs the instrument is operated in the splitless mode with helium as the carrier gas with a flow rate of 1 mL/minute. The temperature of the injection port is 275°C and the detector is 325°C. The initial oven temperature is 100°C, which is held for 1 minute. The ramp rate is 10°C/minute to 150°C, followed by a ramp rate of 6.0°C/minute to 270°C with a hold time of 3 minutes. The retention time of sample analytes must fall within 15 seconds of the retention time of analytes in a calibration standard or a retention index solutions. The levels of aroclors and toxophene are determined using retention index solutions of both complex mixtures. Arochlors are determined in a similar method to that described in EPA SW-846 Test Methods for Evaluating Solid Waste Physical/Chemical Methods, Method 8082 (1997).

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

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Environmental Protection Agency, "Method 8082: Polychlorinated Biphenls (PCBs) by Gas Chromatography," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997)

Method Codes:	001 009

Lab Matrix	Analyte
Animal Tissue	n-decane
	n-docosane
	n-dodecane
	n-dotriacontane
	n-eicosane
	n-heneicosane
	n-hentriacontane
	n-heptacosane
	n-heptadecane
	n-hexacosane
	n-hexadecane
	n-nonacosane
	n-nonadecane
	n-octacosane
	n-octadecane
	n-pentacosane
	n-pentadecane
	n-tetracosane
	n-tetradecane
	n-tetratriacontane
	n-triacontane
	n-tricosane
	n-tridecane
	n-tritriacontane
	n-undecane
	phytane
	pristane

Method Code: 001
LABORATORY: TDI Brooks International, Inc. Tissue Extraction Method (PAH and OCs)

Tissue samples are either immediately processed or stored frozen (-20°C) until processing. Tissues are processed as appropriate for the tissue type (e.g. dissection, shucking). Processed tissues are homogenized using a variety of mechanical methods (Waring blender, Hobart meat grinder or Tissumisser), depending upon the tissue amount and type. After homogenization, an approximate 1 g aliquot is removed and dried in an oven at 105°C to a constant weight to determine % moisture. The remaining samples are stored in certified pre-cleaned jars frozen (- 20°C) until analysis. Prior to extraction, tissue samples may be lypholized or a wet aliquot is chemically dried using a dessicant such as Hydromatrix or sodium sulfate. Samples are then extracted using a Dionex ASE200 Accelerated Solvent Extractor (ASE). The dried sample or the sample and dessicant material is loaded into 22 or 33 mL stainless steel ASE extraction tubes. The extraction are performed using 100% dicholormethane at 100°C and 2000 psi. The extracted organics dissolved in the solvent are collected in 60 mL glass vials. The extract is concentrated to approximately 10 mL in the collection vials and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60°C. If lipid weight is required, a 100 mL aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic material (primarily lipids) must be removed prior to instrument analyses.

The extract is processed through silica gel/alumina chromatography columns and High Performance Liquid Chromatography (HPLC). The remaining 2.9 mL of sample extracted are loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50°C. The samples is transferred into 4 mL amber vials. Extracts subsequently processed by HPLC to further remove lipid interferences. Lipid removal is accomplished by flushing samples with dichlormethane through size exclusion Phenogel 10 m GPC 100 A columns. Approximately 40 mL is collected using a fraction collector, which is concentrated to 0.5 mL using a water bath at 55-60C. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography is required to separate PCBs from toxaphene/pesticides when toxphene analysis is required and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract prior to HPLC clean up is processed through a 3% deactivated silica gel column. The column is packed in diclorometh ane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60°C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42°C and 20 psi. The sample is ready for instrument analysis.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National

Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

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Schantz, M.; Nichols, J. J.; Wise, S. A.; Evaluation of Pressurized Fluid Extraction for the Extraction of Environmental Matrix Reference Material, Anal. Chem., 1997, 69, 4210-4219.

Method Code: 009

LABORATORY: TDI Brooks International, Inc.

Determination of Aliphatic Hydrocarbons in Soil/Sediment

Sediment samples are extracted as described in method 005. Aliphatic hydrocarbons are determined by quantifying target analytes with a gas chromatography/flame ionization detection (GC/FID)

Aliphatic hydrocarbons are analyzed using a HewelettPackard, model 5890 Gas Chromatograph (GC) with a Flame Ionization Detector (FID) operated in a splitless mode. A HP-1MS capillary column (30m x 0.25 mm ID and 0.25 mm film thickness) is used to resolve peaks. The carrier gas is helium at a flow rate of 1.5 mL/min. The temperature of the injection port is 300°C and transfer line is 300C. The initial oven temperature is 60°C, the ramp rate is 12°C/min to a final oven temperature of 180°C. Normal alkanes with 10 to 34 carbons and the isoprenoids pristine and phytane are determined using this procedure. For analytes of interest, a response factor relative to the internal standard is determined at each calibration level. All 5 response factors are averaged for a mean relative response factor. Data are surrogate corrected.

Environmental Protection Agency, "Method 8100/8015. Polynuclear Aromatic Hydrocarbons/Nonhalogenated Organics using GC/FID"," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997).

Method Codes:	001 012

Lab Matrix	Analyte
Animal Tissue	BDE# 1

BDE# 10
BDE# 100
BDE# 11
BDE# 116
BDE# 118
BDE# 119
BDE# 12
BDE# 126
BDE# 13
BDE# 138
BDE# 15
BDE# 153
BDE# 154
BDE# 155
BDE# 166
BDE# 17
BDE# 181
BDE# 183
BDE# 190
BDE# 194
BDE# 195
BDE# 196
BDE# 197
BDE# 198/199/203/200
BDE# 2
BDE# 201
BDE# 202
BDE# 204
BDE# 205
BDE# 206
BDE# 207
BDE# 208
BDE# 209
BDE# 25

BDE# 28
BDE# 3
BDE# 30
BDE# 32
BDE# 33
BDE# 35
BDE# 37
BDE# 47
BDE# 49/71
BDE# 66
BDE# 7
BDE# 75
BDE# 77
BDE# 8
BDE# 85
BDE# 99
BDE-TOTAL

Method Code: 001

LABORATORY: TDI Brooks International, Inc.

Tissue Extraction Method (PAH and OCs)

Tissue samples are either immediately processed or stored frozen (-20°C) until processing. Tissues are processed as appropriate for the tissue type (e.g. dissection, shucking). Processed tissues are homogenized using a variety of mechanical methods (Waring blender, Hobart meat grinder or Tissumisser), depending upon the tissue amount and type. After homogenization, an approximate 1 g aliquot is removed and dried in an oven at 105°C to a constant weight to determine % moisture. The remaining samples are stored in certified pre-cleaned jars frozen (- 20°C) until analysis. Prior to extraction, tissue samples may be lypholized or a wet aliquot is chemically dried using a dessicant such as Hydromatrix or sodium sulfate. Samples are then extracted using a Dionex ASE200 Accelerated Solvent Extractor (ASE). The dried sample or the sample and dessicant material is loaded into 22 or 33 mL stainless steel ASE extraction tubes. The extraction are performed using 100% dicholormethane at 100°C and 2000 psi. The extracted organics dissolved in the solvent are collected in 60 mL glass vials. The extract is concentrated to approximately 10 mL in the collection vials and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60°C. If lipid weight is required, a 100 mL aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic material (primarily lipids) must be removed prior to instrument analyses.

The extract is processed through silica gel/alumina chromatography columns and High Performance Liquid Chromatography

(HPLC). The remaining 2.9 mL of sample extracted are loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50°C. The samples is transferred into 4 mL amber vials. Extracts subsequently processed by HPLC to further remove lipid interferences. Lipid removal is accomplished by flushing samples with dichlormethane through size exclusion Phenogel 10 m GPC 100 A columns. Approximately 40 mL is collected using a fraction collector, which is concentrated to 0.5 mL using a water bath at 55-60C. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography is required to separate PCBs from toxaphene/pesticides when toxphene analysis is required and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract prior to HPLC clean up is processed through a 3% deactivated silica gel column. The column is packed in dicloromethane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60°C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42°C and 20 psi. The sample is ready for instrument analysis.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Environmental Protection Agency, "Method 3545: Pressurized Fluid Extraction (PFE)," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997)

Zuloaga, O.; Etxebarria, N.; Fernandez L. A.;Madariaga, J.M.; Optimization and comparison of MAE, ASE and Soxhlet extraction for the determination of HCH isomers in soil samples. Fresenius J Anal Chem, 2000, 367, 733-737.

Schantz, M.; Nichols, J. J.; Wise, S. A.; Evaluation of Pressurized Fluid Extraction for the Extraction of Environmental Matrix

Reference Material, Anal. Chem., 1997, 69, 4210-4219.

Method Code: 012

Laboratory: TDI Brooks International, Inc.

Polybrominated Diphenyl Ether (PBDE) Determination by Negative Chemical Ionization (NCI)-Selected Ion Monitoring (SIM) Gas Chromatography/Mass Spectrometry (GC/MS)

Polybrominated diphenyl ethers are analyzed in sample extracts by a Thermo Trace GC and DSQ-II MS operated in SIM using a capillary column. The GC is operated in splitless mode using a PTV injection port and the capillary column is an Agilent Technologies DB-XLB (15 m x 0.25 mm ID and 0.10 mm film thickness). The carrier gas is helium at a flow rate of 1 mL/minute and methane is used as the reactant gas with a flow rate of 2.0 mL/min. The temperature of the injection port is 40 degrees C (ramp to 300 degrees C) and transfer line is 290 degrees C. The initial oven temperature is 110 degrees C, the ramp rate is 7 degrees C/minutes to a final oven temperature of 280 degrees C and held for 20 minutes. For analyte identification, the extracted ion current profiles of the primary m/z and the confirmatory ion for each analyte must be at a maximum in the same scan or within one scan of each other and the retention time must fall with 5 seconds of the retention time of the authentic standard.

Method Codes:	001 014

Lab Matrix	Analyte
Animal Tissue	1,2,3,4,6,7,8-HpCDD
	1,2,3,4,6,7,8-HpCDF
	1,2,3,4,7,8,9-HpCDF
	1,2,3,4,7,8-HxCDD
	1,2,3,4,7,8-HxCDF
	1,2,3,6,7,8-HxCDD
	1,2,3,6,7,8-HxCDF
	1,2,3,7,8,9-HxCDD
	1,2,3,7,8,9-HxCDF
	1,2,3,7,8-PeCDD
	1,2,3,7,8-PeCDF
	2,3,4,6,7,8-HxCDF
	2,3,4,7,8-PeCDF
	2,3,7,8-TCDD
	2,3,7,8-TCDF
	Cl4-PCDD

	Cl4-PCDF
	Cl5-PCDD
	Cl5-PCDF
	Cl6-PCDD
	Cl6-PCDF
	Cl7-PCDD
	Cl7-PCDF
	OCDD
	OCDF
Water	1,2,3,4,6,7,8-HpCDD
	1,2,3,4,6,7,8-HpCDF
	1,2,3,4,7,8,9-HpCDF
	1,2,3,4,7,8-HxCDD
	1,2,3,4,7,8-HxCDF
	1,2,3,6,7,8-HxCDD
	1,2,3,6,7,8-HxCDF
	1,2,3,7,8,9-HxCDD
	1,2,3,7,8,9-HxCDF
	1,2,3,7,8-PeCDD
	1,2,3,7,8-PeCDF
	2,3,4,6,7,8-HxCDF
	2,3,4,7,8-PeCDF
	2,3,7,8-TCDD
	2,3,7,8-TCDF
	Cl4-PCDD
	Cl4-PCDF
	Cl5-PCDD
	Cl5-PCDF
	Cl6-PCDD
	Cl6-PCDF
	Cl7-PCDD
	Cl7-PCDF
	OCDD
	OCDF

Method Code: 001

LABORATORY: TDI Brooks International, Inc.

Tissue Extraction Method (PAH and OCs)

Tissue samples are either immediately processed or stored frozen (-20°C) until processing. Tissues are processed as appropriate for the tissue type (e.g. dissection, shucking). Processed tissues are homogenized using a variety of mechanical methods (Waring blender, Hobart meat grinder or Tissumisser), depending upon the tissue amount and type. After homogenization, an approximate 1 g aliquot is removed and dried in an oven at 105°C to a constant weight to determine % moisture. The remaining samples are stored in certified pre-cleaned jars frozen (- 20°C) until analysis. Prior to extraction, tissue samples may be lypholized or a wet aliquot is chemically dried using a dessicant such as Hydromatrix or sodium sulfate. Samples are then extracted using a Dionex ASE200 Accelerated Solvent Extractor (ASE). The dried sample or the sample and dessicant material is loaded into 22 or 33 mL stainless steel ASE extraction tubes. The extraction are performed using 100% dicholormethane at 100°C and 2000 psi. The extracted organics dissolved in the solvent are collected in 60 mL glass vials. The extract is concentrated to approximately 10 mL in the collection vials and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60°C. If lipid weight is required, a 100 mL aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic material (primarily lipids) must be removed prior to instrument analyses.

The extract is processed through silica gel/alumina chromatography columns and High Performance Liquid Chromatography (HPLC). The remaining 2.9 mL of sample extracted are loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50°C. The samples is transferred into 4 mL amber vials. Extracts subsequently processed by HPLC to further remove lipid interferences. Lipid removal is accomplished by flushing samples with dichlormethane through size exclusion Phenogel 10 m GPC 100 A columns. Approximately 40 mL is collected using a fraction collector, which is concentrated to 0.5 mL using a water bath at 55-60C. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography is required to separate PCBs from toxaphene/pesticides when toxphene analysis is required and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract prior to HPLC clean up is processed through a 3% deactivated silica gel column. The column is packed in dicloromethane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60°C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL

of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42°C and 20 psi. The sample is ready for instrument analysis.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Environmental Protection Agency, "Method 3545: Pressurized Fluid Extraction (PFE)," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997)

Zuloaga, O.; Etxebarria, N.; Fernandez L. A.;Madariaga, J.M.; Optimization and comparison of MAE, ASE and Soxhlet extraction for the determination of HCH isomers in soil samples. Fresenius J Anal Chem, 2000, 367, 733-737.

Schantz, M.; Nichols, J. J.; Wise, S. A.; Evaluation of Pressurized Fluid Extraction for the Extraction of Environmental Matrix Reference Material, Anal. Chem., 1997, 69, 4210-4219.

Method Code: 014

LABORATORY: TDI Brooks International, Inc

Summary of Method 1613B

This procedure uses matrix specific extraction, analyte specific cleanup, and HRGC/HRMS analysis techniques for dioxins and furans.

If interferences are encountered, the method provides selected cleanup procedures to aid the analyst in their elimination.

A specified amount of sample is spiked with a solution containing specified amounts of each of the fifteen isotopically labeled PCDDs/PCDFs. The sample is then extracted according to a matrix specific extraction procedure. Aqueous samples that are judged to contain 1 percent or more solids, and solid samples that show an aqueous phase, are filtered, the solid phase (including the filter) and the aqueous phase extracted separately, and the extracts combined before extract cleanup.

The extraction procedures are:

 Toluene: Soxhlet extraction for soil, sediment, fly ash, and paper pulp samples;

 Methylene chloride, liquid-liquid extraction for water samples

 Toluene: Dean-Stark extraction for fuel oil, and aqueous sludge samples;

 Toluene extraction for still bottom samples;

 Hexane/methylene chloride, Soxhlet extraction or methylene chloride, Soxhlet extraction for fish tissue samples

 Methylene chloride extraction for human adipose tissue samples

As an option, all solid samples (wet or dry) may be extracted with toluene using a Soxhlet/Dean Stark extraction system. The decision for the selection of an extraction procedure for chemical reactor residue samples is based on the appearance (consistency, viscosity) of the samples. Generally, they can be handled according to the procedure used for still bottom (or chemical sludge) samples.

The extracts are submitted to an acidic washing treatment and dried. Following a solvent exchange step, the extracts are cleaned up by column chromatography on silica gel and activated carbon. The extracts from adipose tissue samples are treated with silica gel impregnated with sulfuric acid before chromatography on acidic silica gel, and activated carbon.

Fish tissue and paper pulp extracts are subjected to an acid wash treatment only, prior to chromatography on alumina and activated carbon.

The preparation of the final extract for HRGC/HRMS analysis is accomplished by adding 20µL of a nonane solution containing 100pg/µL of the internal standard compounds, 13C12 -1,2,3,4-TCDD and 13C12 -1,2,3,7,8,9-HxCDD. The former is used to determine the percent recoveries of tetra- and pentachlorinated PCDD/PCDF congeners, while the latter is used to determine the percent recoveries of the hexa-, hepta- and octachlorinated PCDD/PCDF congeners.

Two µL of the concentrated extract are injected into an HRGC/HRMS system capable of performing selected ion monitoring at resolving powers of at least 10,000 (10 percent valley definition).

The identification of sixteen 2,3,7,8-substituted congeners, for which a 13C12-labeled standard is available in the sample and the labeled standard spiking solutions, is based on their elution at their exact retention time (within 0.005 retention time units measured in the routine calibration) and the simultaneous detection of the two most abundant ions in the molecular ion region. The identification of OCDF is based on its retention time relative to
13C12 -OCDD and the simultaneous detection of the two most abundant ions in the molecular ion region. Identification is also based on a comparison of the ratios of the integrated ion abundance of the molecular ion species to their theoretical abundance ratios.

Quantitation of the individual congeners, total PCDDs and total PCDFs is achieved in conjunction with the establishment of a multipoint (five points) calibration curve for each homologue, during which each calibration solution is analyzed once.

Method Codes:	001 015

Lab Matrix	Analyte
Animal Tissue	PCB# 1
	PCB# 10
	PCB# 103
	PCB# 104
	PCB# 105
	PCB# 106
	PCB# 107
	PCB# 108/124
	PCB# 11
	PCB# 110/115
	PCB# 111
	PCB# 112
	PCB# 114
	PCB# 117
	PCB# 118
	PCB# 12/13
	PCB# 120
	PCB# 121
	PCB# 122
	PCB# 123
	PCB# 126
	PCB# 127
	PCB# 128/166
	PCB# 129/138/163
	PCB# 130
	PCB# 131
	PCB# 132
	PCB# 133
	PCB# 134
	PCB# 135/151
	PCB# 136
	PCB# 137

PCB# 139/140
PCB# 14
PCB# 141
PCB# 142
PCB# 143
PCB# 144
PCB# 145
PCB# 146
PCB# 147/149
PCB# 148
PCB# 15
PCB# 150
PCB# 152
PCB# 153/168
PCB# 154
PCB# 155
PCB# 156/157
PCB# 158
PCB# 159
PCB# 16
PCB# 160
PCB# 161
PCB# 162
PCB# 164
PCB# 165
PCB# 167
PCB# 169
PCB# 17
PCB# 170
PCB# 171/173
PCB# 172
PCB# 174
PCB# 175
PCB# 176

PCB# 177
PCB# 178
PCB# 179
PCB# 18/30
PCB# 180/193
PCB# 181
PCB# 182
PCB# 183
PCB# 184
PCB# 185
PCB# 186
PCB# 187
PCB# 188
PCB# 189
PCB# 19
PCB# 190
PCB# 191
PCB# 192
PCB# 194
PCB# 195
PCB# 196
PCB# 197
PCB# 198/199
PCB# 2
PCB# 20/28
PCB# 200
PCB# 201
PCB# 202
PCB# 203
PCB# 204
PCB# 205
PCB# 206
PCB# 207
PCB# 208

PCB# 209
PCB# 21/33
PCB# 22
PCB# 23
PCB# 24
PCB# 25
PCB# 26/29
PCB# 27
PCB# 3
PCB# 31
PCB# 32
PCB# 34
PCB# 35
PCB# 36
PCB# 37
PCB# 38
PCB# 39
PCB# 4
PCB# 40/41/70
PCB# 42
PCB# 43/73
PCB# 44/47/65
PCB# 45/51
PCB# 46
PCB# 48
PCB# 49/69
PCB# 5
PCB# 50/53
PCB# 52
PCB# 54
PCB# 55
PCB# 56
PCB# 57
PCB# 58

PCB# 59/62/75
PCB# 6
PCB# 60
PCB# 63
PCB# 64
PCB# 66
PCB# 67
PCB# 68
PCB# 7
PCB# 70/61/74/76
PCB# 72
PCB# 77
PCB# 78
PCB# 79
PCB# 8
PCB# 80
PCB# 81
PCB# 82
PCB# 83/99
PCB# 84
PCB# 85/116
PCB# 86/87/97/109/119/125
PCB# 88/91
PCB# 89
PCB# 9
PCB# 90/101/113
PCB# 92
PCB# 94
PCB# 95
PCB# 96
PCB# 98/100
PCB# 98/102
PCB-1242
PCB-1248

	PCB-1254
	PCB-1260
	PCB-TOTAL
Water	PCB# 1
	PCB# 10
	PCB# 103
	PCB# 104
	PCB# 105
	PCB# 106
	PCB# 107
	PCB# 108/124
	PCB# 11
	PCB# 110/115
	PCB# 111
	PCB# 112
	PCB# 114
	PCB# 117
	PCB# 118
	PCB# 12/13
	PCB# 120
	PCB# 121
	PCB# 122
	PCB# 123
	PCB# 126
	PCB# 127
	PCB# 128/166
	PCB# 129/138/163
	PCB# 130
	PCB# 131
	PCB# 132
	PCB# 133
	PCB# 134
	PCB# 135/151
	PCB# 136

PCB# 137
PCB# 139/140
PCB# 14
PCB# 141
PCB# 142
PCB# 143
PCB# 144
PCB# 145
PCB# 146
PCB# 147/149
PCB# 148
PCB# 15
PCB# 150
PCB# 152
PCB# 153/168
PCB# 154
PCB# 155
PCB# 156/157
PCB# 158
PCB# 159
PCB# 16
PCB# 160
PCB# 161
PCB# 162
PCB# 164
PCB# 165
PCB# 167
PCB# 169
PCB# 17
PCB# 170
PCB# 171/173
PCB# 172
PCB# 174
PCB# 175

PCB# 176
PCB# 177
PCB# 178
PCB# 179
PCB# 18/30
PCB# 180/193
PCB# 181
PCB# 182
PCB# 183
PCB# 184
PCB# 185
PCB# 186
PCB# 187
PCB# 188
PCB# 189
PCB# 19
PCB# 190
PCB# 191
PCB# 192
PCB# 194
PCB# 195
PCB# 196
PCB# 197
PCB# 198/199
PCB# 2
PCB# 20/28
PCB# 200
PCB# 201
PCB# 202
PCB# 203
PCB# 204
PCB# 205
PCB# 206
PCB# 207

PCB# 208
PCB# 209
PCB# 21/33
PCB# 22
PCB# 23
PCB# 24
PCB# 25
PCB# 26/29
PCB# 27
PCB# 3
PCB# 31
PCB# 32
PCB# 34
PCB# 35
PCB# 36
PCB# 37
PCB# 38
PCB# 39
PCB# 4
PCB# 40/41/70
PCB# 42
PCB# 43/73
PCB# 44/47/65
PCB# 45/51
PCB# 46
PCB# 48
PCB# 49/69
PCB# 5
PCB# 50/53
PCB# 52
PCB# 54
PCB# 55
PCB# 56
PCB# 57

PCB# 58
PCB# 59/62/75
PCB# 6
PCB# 60
PCB# 63
PCB# 64
PCB# 66
PCB# 67
PCB# 68
PCB# 7
PCB# 70/61/74/76
PCB# 72
PCB# 77
PCB# 78
PCB# 79
PCB# 8
PCB# 80
PCB# 81
PCB# 82
PCB# 83/99
PCB# 84
PCB# 85/116
PCB# 86/87/97/109/119/125
PCB# 88/91
PCB# 89
PCB# 9
PCB# 90/101/113
PCB# 92
PCB# 94
PCB# 95
PCB# 96
PCB# 98/100
PCB# 98/102
PCB-TOTAL

Method Code: 001

LABORATORY: TDI Brooks International, Inc.

Tissue Extraction Method (PAH and OCs)

Tissue samples are either immediately processed or stored frozen (-20°C) until processing. Tissues are processed as appropriate for the tissue type (e.g. dissection, shucking). Processed tissues are homogenized using a variety of mechanical methods (Waring blender, Hobart meat grinder or Tissumisser), depending upon the tissue amount and type. After homogenization, an approximate 1 g aliquot is removed and dried in an oven at 105°C to a constant weight to determine % moisture. The remaining samples are stored in certified pre-cleaned jars frozen (- 20°C) until analysis. Prior to extraction, tissue samples may be lypholized or a wet aliquot is chemically dried using a dessicant such as Hydromatrix or sodium sulfate. Samples are then extracted using a Dionex ASE200 Accelerated Solvent Extractor (ASE). The dried sample or the sample and dessicant material is loaded into 22 or 33 mL stainless steel ASE extraction tubes. The extraction are performed using 100% dicholormethane at 100°C and 2000 psi. The extracted organics dissolved in the solvent are collected in 60 mL glass vials. The extract is concentrated to approximately 10 mL in the collection vials and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60°C. If lipid weight is required, a 100 mL aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic material (primarily lipids) must be removed prior to instrument analyses.

The extract is processed through silica gel/alumina chromatography columns and High Performance Liquid Chromatography (HPLC). The remaining 2.9 mL of sample extracted are loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50°C. The samples is transferred into 4 mL amber vials. Extracts subsequently processed by HPLC to further remove lipid interferences. Lipid removal is accomplished by flushing samples with dichlormethane through size exclusion Phenogel 10 m GPC 100 A columns. Approximately 40 mL is collected using a fraction collector, which is concentrated to 0.5 mL using a water bath at 55-60C. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography is required to separate PCBs from toxaphene/pesticides when toxphene analysis is required and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract prior to HPLC clean up is processed through a 3% deactivated silica gel column. The column is packed in dicloromethane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60°C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL

of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42°C and 20 psi. The sample is ready for instrument analysis.

REFERENCES:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Environmental Protection Agency, "Method 3545: Pressurized Fluid Extraction (PFE)," in Test Methods for Evaluating Solid Waste, Physical/Chemical Methods EPA SW-846 [Version 2 (December 1997), Integrated Manual through Update III] Washington DC, U.S. Environmental Protection Agency (1997)

Zuloaga, O.; Etxebarria, N.; Fernandez L. A.;Madariaga, J.M.; Optimization and comparison of MAE, ASE and Soxhlet extraction for the determination of HCH isomers in soil samples. Fresenius J Anal Chem, 2000, 367, 733-737.

Schantz, M.; Nichols, J. J.; Wise, S. A.; Evaluation of Pressurized Fluid Extraction for the Extraction of Environmental Matrix Reference Material, Anal. Chem., 1997, 69, 4210-4219.

Method Code: 015

LABORATORY: TDI Brooks International, Inc

Summary of Method 1668A

This procedure uses matrix specific extraction, analyte specific cleanup, and HRGC/HRMS analysis techniques for PCB congeners.

Aqueous samples (samples containing less than one percent solids) ¨C Stable isotopically labeled analogs of the toxics and labeled earliest and latest level of chlorination (LOC) PCBs are spiked into the sample aliquot. The sample is extracted using a separatory funnel extraction and concentrated for clean up.

Solid, semi-solid, and multi-phase samples (excluding tissue) ¨C The labeled compounds are spiked into sample material that corresponds to approximately 5g on a dry weight basis. For samples with high moisture content, the amount of wet weight material to add the labeled compounds to would necessarily be greater than 5g. All samples are homogenized properly and extracted in a Soxhlet extraction apparatus or Accelerated Solvent Extractor (ASE). The extract is concentrated for clean up.

Fish and other tissue ¨C An aliquot of tissue sufficient enough to provide sample for PCB and lipid analysis is homogenized and spiked with labeled compounds. The sample is mixed with anhydrous sodium sulfate, allowed to dry for 12-24 hours, and extracted for 16-24 hours using a 1:1 dichloromethane/hexane solution in a Soxhlet extraction apparatus. The extract is split; with one aliquot concentrated for clean up and the other concentrated for lipid determination.

After extraction, a labeled clean up standard is spiked into the extract which is then cleaned up using back-extraction with sulfuric acid and/or base, and silica gel or Florisil chromatography.

After clean up, the extract is concentrated to 20¦ÌL. Immediately prior to injection, labeled injection internal standards are added to each extract and an aliquot of the extract is injected into the gas chromatograph (GC). The analytes are separated by the GC and detected by a high resolution (¡Ý10,000) mass spectrometer. Two exact m/z¡¯s are monitored at each level of chlorination throughout a pre-determined retention time window.

An individual PCB congener is identified by comparing the GC retention time and ion-abundance ratio of two exact m/z¡¯s with the corresponding retention time of an authentic standard and the theoretical or acquired ion-abundance ratio of the two exact m/z¡¯s. Isomer specificity for certain of the CB congeners is achieved using GC columns that resolve these congeners.

Quantitative analysis is performed in one of two ways using selected ion current profile (SICP) areas:

For the toxics and the earliest and latest level of chlorination CBs (Toxics/LOC), the GC/MS is multi-point calibrated and the concentration is determined using the isotope dilution technique.

For all other congeners, the GC/MS is calibrated at a single concentration and the concentrations are determined using the internal standard technique.

For the labeled congeners, the GC/MS is calibrated using replicates at a single concentration and the concentrations of these labeled compounds are determined using the internal standard technique.

The quality of analysis is assured through reproducible calibration and testing of the extraction, clean up and GC/MS systems.

Method Codes:	011 015

Lab Matrix	Analyte
Water	PCB# 1
	PCB# 10
	PCB# 103
	PCB# 104
	PCB# 105
	PCB# 106
	PCB# 107
	PCB# 108/124
	PCB# 11
	PCB# 110/115

PCB# 111
PCB# 112
PCB# 114
PCB# 117
PCB# 118
PCB# 12/13
PCB# 120
PCB# 121
PCB# 122
PCB# 123
PCB# 126
PCB# 127
PCB# 128/166
PCB# 129/138/163
PCB# 130
PCB# 131
PCB# 132
PCB# 133
PCB# 134
PCB# 135/151
PCB# 136
PCB# 137
PCB# 139/140
PCB# 14
PCB# 141
PCB# 142
PCB# 143
PCB# 144
PCB# 145
PCB# 146
PCB# 147/149
PCB# 148
PCB# 15
PCB# 150

PCB# 152
PCB# 153/168
PCB# 154
PCB# 155
PCB# 156/157
PCB# 158
PCB# 159
PCB# 16
PCB# 160
PCB# 161
PCB# 162
PCB# 164
PCB# 165
PCB# 167
PCB# 169
PCB# 17
PCB# 170
PCB# 171/173
PCB# 172
PCB# 174
PCB# 175
PCB# 176
PCB# 177
PCB# 178
PCB# 179
PCB# 18/30
PCB# 180/193
PCB# 181
PCB# 182
PCB# 183
PCB# 184
PCB# 185
PCB# 186
PCB# 187

PCB# 188
PCB# 189
PCB# 19
PCB# 190
PCB# 191
PCB# 192
PCB# 194
PCB# 195
PCB# 196
PCB# 197
PCB# 198/199
PCB# 2
PCB# 20/28
PCB# 200
PCB# 201
PCB# 202
PCB# 203
PCB# 204
PCB# 205
PCB# 206
PCB# 207
PCB# 208
PCB# 209
PCB# 21/33
PCB# 22
PCB# 23
PCB# 24
PCB# 25
PCB# 26/29
PCB# 27
PCB# 3
PCB# 31
PCB# 32
PCB# 34

PCB# 35
PCB# 36
PCB# 37
PCB# 38
PCB# 39
PCB# 4
PCB# 40/41/70
PCB# 42
PCB# 43/73
PCB# 44/47/65
PCB# 45/51
PCB# 46
PCB# 48
PCB# 49/69
PCB# 5
PCB# 50/53
PCB# 52
PCB# 54
PCB# 55
PCB# 56
PCB# 57
PCB# 58
PCB# 59/62/75
PCB# 6
PCB# 60
PCB# 63
PCB# 64
PCB# 66
PCB# 67
PCB# 68
PCB# 7
PCB# 70/61/74/76
PCB# 72
PCB# 77

PCB# 78
PCB# 79
PCB# 8
PCB# 80
PCB# 81
PCB# 82
PCB# 83/99
PCB# 84
PCB# 85/116
PCB# 86/87/97/109/119/125
PCB# 88/91
PCB# 89
PCB# 9
PCB# 90/101/113
PCB# 92
PCB# 94
PCB# 95
PCB# 96
PCB# 98/100
PCB# 98/102
PCB-1242
PCB-1248
PCB-1254
PCB-1260
PCB-TOTAL

Method Code: 011

LABORATORY: TDI-Brooks International, Inc.

Water Extraction Method (PAH and OC)

Water samples are either immediately processed or stored refrigerated (4 degrees C) until processing. The pH of samples is adjusted to <2 by the addition of 1 mL of hydrochloric acid. Samples are serially extracted using separatory funnels and dichloromethane. For a 1 L sample, 100 mL of dichloromethane is added. The sample is then shaken for three minutes. The organic layer is allowed to separate from the aqueous phase. The organic layer is drained into a 500 mL round bottom flask

through a glass funnel containing a glass fiber filter and combusted sodium sulfate. This process is repeated twice more. The extract is concentrated to approximately 10 mL in the 500 mL round bottom flasks in a water bath at 55-60 degrees C and then transferred to 25 mL Kurdena-Danish (KD) concentrator tubes. The sample extract is concentrated to 3 mL in a water bath at 55-60 degrees C. If extractable organic material weight is required, a 100 micro litter aliquot is removed and weighed using a microbalance. Interfering non-contaminant organic materials may be removed prior to instrument analyses.

If the extract is colored then sample clean up may be required. The extract is processed through silica gel/alumina chromatography columns. The sample extract is loaded on top of 300 mm x 19 mm glass liquid chromatography columns packed with 10 g of deactivated alumina and 20 g of deactivated silica gel. The columns are loaded in 100 % dichloromethane. The dichloromethane is replaced by adding 40 mL of pentane. The extract is carefully added to the top of the chromatography columns. The column is flushed at a rate of 1-2 mL per minute using 200 mL of 50:50 pentane/dichloromethane and collected into 250 mL flasks. The eluent collected in the 250 mL flask is evaporated to 2 mL using a waterbath at 55-50 degrees C. The samples is transferred into 2 mL amber vials. The concentrated extract is then analyzed by GC/MS for polynuclear aromatic hydrocarbons (PAHs) or GC/ECD for selected organochlorines (OCs).

Additional column chromatography may be required to separate PCBs from toxaphene/pesticides when toxphene analysis is required and to separate planar PCBs. If toxaphene analyses is required, an aliquot of the extract after silica/alumina clean-up is processed through a 3% deactivated silica gel column. The column is packed in diclorometh ane which is then flushed with 50 mL of pentane. The sample extract is transferred to the top of the column and flushed with 100 mL of pentane. The fraction contains PCBs and DDTs. The column is then flushed with 120 mL of 50:50 pentane/dichloromethane. This fraction contains toxaphene and chlorinated pesticides. Both fractions are reduced to 1 mL using a water bath at 55-60 degrees C. The extracts are then ready for instrument analysis.

If planar PCB analyses are required, the PCB/DDT fraction prepared by 3% silica gel column is further processed by column chromatography packed with 2 g of 1:19 (5% by weight) mixture of activated carbon/Celite. The column and flushed with 25 mL of 1:4 dichloromethane/cyclohexane mixture. The sample is added to the top of the column and flushed with 50 mL of 1:4 dichloromethane/cyclohexane mixture, followed by 30 mL of 9:1 dichloromethane/toluene. This is followed by the addition of 40 mL of toluene. The toluene fraction contains the planar PCBs and is concentrated to 1 mL in a Zymark TurboVap II concentrator at 42 degrees C and 20 psi. The sample is ready for instrument analysis.

References:

Lauenstein, G.G. and A.Y. Cantillo, ed. (1993). Sampling Analytical Methods of the National Status and Trends Program National Benthic Surveillance and Mussel Watch Projects 1984-1992; Volume IV: Comprehensive Descriptions of Trace Organic Analytical Methods. NOAA Technical memorandum NOS ORCA 71, Silver Spring, MD.

U.S. Environmental Protection Agency. 2001. National Coastal Assessment Quality Assurance Project Plan 2001-2004. United States Environmental Protection Agency, Office of Research and Development, National Health and Environmental Effects Research Laboratory, Gulf Ecology Division, Gulf Breeze, FL. EPA/620/R-01/002.

Method Code: 015

LABORATORY: TDI Brooks International, Inc

Summary of Method 1668A

This procedure uses matrix specific extraction, analyte specific cleanup, and HRGC/HRMS analysis techniques for PCB congeners.

Aqueous samples (samples containing less than one percent solids) ¨C Stable isotopically labeled analogs of the toxics and labeled earliest and latest level of chlorination (LOC) PCBs are spiked into the sample aliquot. The sample is extracted using a separatory funnel extraction and concentrated for clean up.

Solid, semi-solid, and multi-phase samples (excluding tissue) ¨C The labeled compounds are spiked into sample material that corresponds to approximately 5g on a dry weight basis. For samples with high moisture content, the amount of wet weight material to add the labeled compounds to would necessarily be greater than 5g. All samples are homogenized properly and extracted in a Soxhlet extraction apparatus or Accelerated Solvent Extractor (ASE). The extract is concentrated for clean up.

Fish and other tissue ¨C An aliquot of tissue sufficient enough to provide sample for PCB and lipid analysis is homogenized and spiked with labeled compounds. The sample is mixed with anhydrous sodium sulfate, allowed to dry for 12-24 hours, and extracted for 16-24 hours using a 1:1 dichloromethane/hexane solution in a Soxhlet extraction apparatus. The extract is split; with one aliquot concentrated for clean up and the other concentrated for lipid determination.

After extraction, a labeled clean up standard is spiked into the extract which is then cleaned up using back-extraction with sulfuric acid and/or base, and silica gel or Florisil chromatography.

After clean up, the extract is concentrated to 20¦ÌL. Immediately prior to injection, labeled injection internal standards are added to each extract and an aliquot of the extract is injected into the gas chromatograph (GC). The analytes are separated by the GC and detected by a high resolution (¡Ý10,000) mass spectrometer. Two exact m/z¡¯s are monitored at each level of chlorination throughout a pre-determined retention time window.

An individual PCB congener is identified by comparing the GC retention time and ion-abundance ratio of two exact m/z¡¯s with the corresponding retention time of an authentic standard and the theoretical or acquired ion-abundance ratio of the two exact m/z¡¯s. Isomer specificity for certain of the CB congeners is achieved using GC columns that resolve these congeners.

Quantitative analysis is performed in one of two ways using selected ion current profile (SICP) areas:

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For all other congeners, the GC/MS is calibrated at a single concentration and the concentrations are determined using the internal standard technique.

For the labeled congeners, the GC/MS is calibrated using replicates at a single concentration and the concentrations of these labeled compounds are determined using the internal standard technique.

The quality of analysis is assured through reproducible calibration and testing of the extraction, clean up and GC/MS systems.

Appendix 3C. Complete Catalog 2020128 of Rio Grande silvery minnow carcasses and collection information.

ECDMS Complete Catalog - 2020128

08/26/09

Catalog Submitter

Catalog Title	Rio Grande Silvery Minnow Health Study		
Catalog Submitter	Lusk, Joel		
Submitter Title	Senior Environmental Contaminant Specialist		
Location	New Mexico Ecological Services Field Office, NM		
Submitters Email	joel_lusk@fws.gov		
Phone Number	(505) 761-4709 ext. 109	**Fax Number**	505-346-2542

Catalog Information

Number of Samples	45	**Date Submitted**	08/10/09
Disposition of Samples	Return	**Catalog Validation**	Validated
Chain of Custody	No	**Fields Missing**	0
Submitter has Split Samples	No	**Non-Routine Request**	No
Request Permanent Lock	No	**Rapid Turnaround**	No
Catalog Data Lock	Catalog and sample data are unlocked.		
Results Data Lock	There are no results for this catalog.		

Project Information

DEQ Project ID	N/A	**Regional Study ID**	N/A

FWS Refuge Names

The results of this catalog are directly applicable to the following FWS refuges:
Bosque Del Apache National Wildlife Refuge , New Mexico
Sevilleta National Wildlife Refuge , New Mexico

Catalog Description
Catalog Instructions
The priority where samples have multiple chemical analyses requested is 209 PCB by HRGC/HRMS, lipids, organochlorine pesticides, heavy metals (and As, Hg, Se), PBDEs, dioxins, aromatics and then other analytes. Note that the fish was dissected and stored in a freezer for up to 2 years, and therefore, moisture content has likely largely been lost and is NOT a priority for analysis although this "analyte" has been requested.

Cost Code Information

Priority	Cost Code	Dollars Available

1	09-22420-1946-2013	$40,600
	Total	$40,600

Regional Approval

Region	Current Regional Coordinator	Approval Date	Phone Number	E-Mail Address
2	Laila Lienesch	08/14/09	505-248-6494 ext.	laila_lienesch@fws.gov

Catalog Status Information

Purchase Order Number	Lab Name	Analysis Type	PO Generated	Lab Approval	Current *PCD	QA/QC Approval	Results Location

Catalog Cost

Purchase Order Number	Lab Name	Analysis Type	Cost Code	Cost

Sample Number - 7-06-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	79.6 grams	**Collection Date**	07/27/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG near Bernalillo, NM above and below 550 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32121025	35 N 19 ' 16 "	3909779 N
Longitude	-106.55842789	-106 E 33 ' 30 "	358343 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	83.6 grams	**Collection Date**	07/26/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site2 - MRG at Alameda NM, below AMAFCA NDC		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20897414	35 N 12 ' 32 "	3897417 N
Longitude	-106.61797315	-106 E 37 ' 4 "	352727 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

PBDE - Polybrominated Diphenly Ethers

Inorganic Analysis

Metals Scan

Sample Number - 7-06-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.9 grams	**Collection Date**	07/25/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	SIte3-MRG at Los Padillas, NM above I-25 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.97261343	34 N 58 ' 21 "	3871311 N
Longitude	-106.69041188	-106 E 41 ' 25 "	345688 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all fish from the site were removed from foil and composited into a jar for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	59.0 grams	**Collection Date**	07/20/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 - MRG at Los Lunas - above Main Street Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81456722	34 N 48 ' 52 "	3853814 N
Longitude	-106.71109672	-106 E 42 ' 39 "	343500 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	64.5 grams	**Collection Date**	07/19/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM. above Sevilleta NWR portion of Rio		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**

Latitude	34.31230631	34 N 18 ' 44 "	3798341 N
Longitude	-106.854025	-106 E 51 ' 14 "	329403 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	40.3 grams	**Collection Date**	07/18/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 - MRG near San Antonio, NM north of Bosque del Apache NWR border		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	33.87452474	33 N 52 ' 28 "	3749783 N
Longitude	-106.84959324	-106 E 50 ' 58 "	328932 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	63.0 grams	**Collection Date**	07/16/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG at Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32125645	35 N 19 ' 16 "	3909784 N
Longitude	-106.55834077	-106 E 33 ' 30 "	358351 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*

N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	53.6 grams	**Collection Date**	07/15/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM above Alameda Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20892995	35 N 12 ' 32 "	3897412 N
Longitude	-106.61790636	-106 E 37 ' 4 "	352733 E

Coordinate Method	UTM Coordinates

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 4-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	189.9 grams	**Collection Date**	04/13/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 - MRG at Los Padillas (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.01396395	35 N 0 ' 50 "	3875872 N
Longitude	-106.67395835	-106 E 40 ' 26 "	347267 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	28.3 grams	**Collection Date**	07/10/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4- MRG near Los Lunas (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81916416	34 N 49 ' 8 "	3854327 N
Longitude	-106.713116	-106 E 42 ' 47 "	343324 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*

N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-08-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	76.3 grams	**Collection Date**	01/23/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31200231	34 N 18 ' 43 "	3798307 N
Longitude	-106.85385532	-106 E 51 ' 13 "	329418 E

Coordinate Method	UTM Coordinates

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 10-07-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	25.7 grams	**Collection Date**	10/31/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.3125136	35 N 18 ' 45 "	3908815 N
Longitude	-106.55884398	-106 E 33 ' 31 "	358290 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	26.7 grams	**Collection Date**	10/30/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG at Alameda, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20886524	35 N 12 ' 31 "	3897405 N
Longitude	-106.61802592	-106 E 37 ' 4 "	352722 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	43.3 grams	**Collection Date**	10/29/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas, NM (above I-25 Bridge)		

FWS Refuge	N/A

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.9735791	34 N 58 ' 24 "	3871418 N
Longitude	-106.6903441	-106 E 41 ' 25 "	345696 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	39.8 grams	**Collection Date**	10/25/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM above main st bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81863921	34 N 49 ' 7 "	3854269 N
Longitude	-106.7132582	-106 E 42 ' 47 "	343310 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	46.9 grams	**Collection Date**	10/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31200231	34 N 18 ' 43 "	3798307 N
Longitude	-106.85385532	-106 E 51 ' 13 "	329418 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	29.0 grams	**Collection Date**	10/23/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio NM (above BdANWR border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87600472	33 N 52 ' 33 "	3749947 N
Longitude	-106.84951707	-106 E 50 ' 58 "	328942 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 1-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	31.9 grams	**Collection Date**	01/30/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.31245022	35 N 18 ' 44 "	3908808 N
Longitude	-106.55886476	-106 E 33 ' 31 "	358288 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	52.6 grams	**Collection Date**	01/28/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM above Alameda Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's

Latitude	35.20892167	35 N 12 ' 32 "	3897411 N
Longitude	-106.61785127	-106 E 37 ' 4 "	352738 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	50.3 grams	**Collection Date**	01/28/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20786166	35 N 12 ' 28 "	3897411 N
Longitude	-106.69530751	-106 E 41 ' 43 "	345685 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	47.1 grams	**Collection Date**	01/24/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81820555	34 N 49 ' 5 "	3854220 N
Longitude	-106.71268071	-106 E 42 ' 45 "	343362 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	55.9 grams	**Collection Date**	01/22/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (just above BdANWR north border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87474141	33 N 52 ' 29 "	3749807 N
Longitude	-106.84957629	-106 E 50 ' 58 "	328934 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis

Metals Scan

Sample Number - 4-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	33.5 grams	**Collection Date**	04/14/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32108775	35 N 19 ' 15 "	3909765 N
Longitude	-106.55813952	-106 E 33 ' 29 "	358369 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 10-06-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	77.8 grams	**Collection Date**	10/12/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM (above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21089567	35 N 12 ' 39 "	3897432 N
Longitude	-106.61757152	-106 E 37 ' 3 "	352764 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	79.0 grams	**Collection Date**	10/07/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas (above I-25 Bridge)		

FWS Refuge	N/A

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.95743122	34 N 57 ' 26 "	3869618 N
Longitude	-106.68450383	-106 E 41 ' 4 "	346199 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
Necropsy fish composite - Individual health observations available. Gonads were removed. Some fluids were likely lost also.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize all minnow in sample before analyses. Note that priority for analysis is 209 PCB congeners, then OC + PCB aroclors, then dioxin/furan scan.

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)

PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	122.0 grams	**Collection Date**	10/16/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81717205	34 N 49 ' 1 "	3854105 N
Longitude	-106.71244066	-106 E 42 ' 44 "	343382 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil

and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	110.2 grams	**Collection Date**	10/05/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.31230631	34 N 18 ' 44 "	3798341 N
Longitude	-106.854025	-106 E 51 ' 14 "	329403 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 4-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A

Sample Weight	47.2 grams	**Collection Date**	04/15/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.2107287	35 N 12 ' 38 "	3897615 N
Longitude	-106.62025999	-106 E 37 ' 12 "	352522 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for

results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 4-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	134.7 grams	**Collection Date**	04/08/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 - MRG at Los Lunas, NM (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81907187	34 N 49 ' 8 "	3854317 N
Longitude	-106.71326716	-106 E 42 ' 47 "	343310 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 4-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.4 grams	**Collection Date**	04/09/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (above BdANWR border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87436858	33 N 52 ' 27 "	3749766 N
Longitude	-106.84978446	-106 E 50 ' 59 "	328914 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PBDE - Polybrominated Diphenly Ethers
PCB Quantification of Individual Aroclors
Inorganic Analysis
Metals Scan

Sample Number - 7-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	16.1 grams	**Collection Date**	07/14/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3- MRG at Los Padillas, NM above I-25 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.96696085	34 N 58 ' 1 "	3870683 N
Longitude	-106.68972619	-106 E 41 ' 23 "	345740 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	10.5 grams	**Collection Date**	07/09/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG at La Joya, NM above Sevilleta NWR		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.29909525	34 N 17 ' 56 "	3796864 N
Longitude	-106.84696582	-106 E 50 ' 49 "	330026 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities

(These observations were not necessarily determined by a pathologist)
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	16.6 grams	**Collection Date**	07/08/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 - MRG near San Antonio, NM above Bosque del Apache NWR		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.88534008	33 N 53 ' 7 "	3750975 N

Longitude	-106.8451451	-106 E 50 ' 42 "	329365 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

N/A

Inorganic Analysis

Metals Scan

Methyl Mercury

Sample Number - 10-06-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	38.5 grams	**Collection Date**	10/11/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21213359	35 N 12 ' 43 "	3897763 N
Longitude	-106.61498184	-106 E 36 ' 53 "	353005 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 4-07-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	49.2 grams	**Collection Date**	04/26/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG near Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.31455303	35 N 18 ' 52 "	3909042 N
Longitude	-106.5594001	-106 E 33 ' 33 "	358243 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		

Family	Cyprinidae
Genus / Kingdom	Hybognathus / Animalia
Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	46.7 grams	**Collection Date**	04/25/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG near Alameda, NM (in side channel above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21174211	35 N 12 ' 42 "	3897723 N
Longitude	-106.61729205	-106 E 37 ' 2 "	352794 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis

Metals Scan

Sample Number - 4-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.9 grams	**Collection Date**	04/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas, NM (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.97036675	34 N 58 ' 13 "	3871061 N
Longitude	-106.68988377	-106 E 41 ' 23 "	345732 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	53.1 grams	**Collection Date**	04/19/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM (above Main Street Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81798048	34 N 49 ' 4 "	3854193 N
Longitude	-106.71137504	-106 E 42 ' 40 "	343481 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	29.9 grams	**Collection Date**	04/18/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM (above Sevilleta NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31455461	34 N 18 ' 52 "	3798592 N
Longitude	-106.85500898	-106 E 51 ' 18 "	329317 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 1-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	36.5 grams	**Collection Date**	01/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG near Alameda, NM (above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20905205	35 N 12 ' 32 "	3897433 N
Longitude	-106.62292904	-106 E 37 ' 22 "	352276 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	95.3 grams	**Collection Date**	01/23/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG near Los Padillas, NM (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.21174211	35 N 12 ' 42 "	3897723 N
Longitude	-106.61729205	-106 E 37 ' 2 "	352794 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 2-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	117.5 grams	**Collection Date**	02/01/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG near Los Lunas, NM (above Main Street Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81427857	34 N 48 ' 51 "	3853783 N
Longitude	-106.7117467	-106 E 42 ' 42 "	343440 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Methyl Mercury

Sample Number - 1-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	42.7 grams	**Collection Date**	01/31/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM (above Sevilleta NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.30541029	34 N 18 ' 19 "	3797569 N
Longitude	-106.84973359	-106 E 50 ' 59 "	329784 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-07-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	118.0 grams	**Collection Date**	01/30/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (above Bosque del Apache NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87900195	33 N 52 ' 44 "	3750278 N
Longitude	-106.84868442	-106 E 50 ' 55 "	329025 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		

Genus / Kingdom	Hybognathus / Animalia
Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 06385810

Sample Matrix	Water	**NWHC Case Number**	N/A
Sample Weight	N/A grams	**Collection Date**	06/25/09
Sample Volume	1000.0 ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Survey
Location	San Jose Drain		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.04416667	35 N 2 ' 39 "	3879184 N
Longitude	-106.64944444	-106 E 38 ' 58 "	349559 E
Coordinate Method		Latitude/Longitude Degrees	
Latitude/Longitude Method		GPS Unit - NAD 83 Reference System	

Specimen Information

Sex	N/A	**Age**	N/A
Egg Condition	N/A		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
Water sample collected with teflon nozzle intregated sampler using rod. Added to teflon churn, homogenized then split, unfiltered into container. Sample contains suspended sediment. Kept chilled and shipped chilled for chemical analysis.

Field Preparation Comments
Water sample collected with teflon nozzle intregated sampler using rod. Added to teflon churn, homogenized then split, unfiltered into 1 liter HDPE container. Sample contains suspended sediment. Kept chilled and shipped chilled for chemical analysis.
Analysis Requested Comments
Water sample contains suspended sediment, please homogenize before analysis.

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
Metals Scan

ECDMS Complete Catalog - 2020128

08/26/09

Catalog Submitter

Catalog Title	Rio Grande Silvery Minnow Health Study		
Catalog Submitter	Lusk, Joel		
Submitter Title	Senior Environmental Contaminant Specialist		
Location	New Mexico Ecological Services Field Office, NM		
Submitters Email	joel_lusk@fws.gov		
Phone Number	(505) 761-4709 ext. 109	**Fax Number**	505-346-2542

Catalog Information

Number of Samples	45	**Date Submitted**	08/10/09
Disposition of Samples	Return	**Catalog Validation**	Validated
Chain of Custody	No	**Fields Missing**	0
Submitter has Split Samples	No	**Non-Routine Request**	No
Request Permanent Lock	No	**Rapid Turnaround**	No
Catalog Data Lock	Catalog and sample data are unlocked.		
Results Data Lock	There are no results for this catalog.		

Project Information

DEQ Project ID	N/A	**Regional Study ID**	N/A

FWS Refuge Names

The results of this catalog are directly applicable to the following FWS refuges:
Bosque Del Apache National Wildlife Refuge , New Mexico
Sevilleta National Wildlife Refuge , New Mexico

Catalog Description
Catalog Instructions
The priority where samples have multiple chemical analyses requested is 209 PCB by HRGC/HRMS, lipids, organochlorine pesticides, heavy metals (and As, Hg, Se), PBDEs, dioxins, aromatics and then other analytes. Note that the fish was dissected and stored in a freezer for up to 2 years, and therefore, moisture content has likely largely been lost and is NOT a priority for analysis although this "analyte" has been requested.

Cost Code Information

Priority	Cost Code	Dollars Available

1	09-22420-1946-2013	$40,600
	Total	$40,600

Regional Approval

Region	Current Regional Coordinator	Approval Date	Phone Number	E-Mail Address
2	Laila Lienesch	08/14/09	505-248-6494 ext.	laila_lienesch@fws.gov

Catalog Status Information

Purchase Order Number	Lab Name	Analysis Type	PO Generated	Lab Approval	Current *PCD	QA/QC Approval	Results Location

Catalog Cost

Purchase Order Number	Lab Name	Analysis Type	Cost Code	Cost

Sample Number - 7-06-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	79.6 grams	**Collection Date**	07/27/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG near Bernalillo, NM above and below 550 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32121025	35 N 19 ' 16 "	3909779 N
Longitude	-106.55842789	-106 E 33 ' 30 "	358343 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	83.6 grams	**Collection Date**	07/26/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site2 - MRG at Alameda NM, below AMAFCA NDC		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20897414	35 N 12 ' 32 "	3897417 N
Longitude	-106.61797315	-106 E 37 ' 4 "	352727 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

PBDE - Polybrominated Diphenly Ethers

Inorganic Analysis

Metals Scan

Sample Number - 7-06-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.9 grams	**Collection Date**	07/25/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	SIte3-MRG at Los Padillas, NM above I-25 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.97261343	34 N 58 ' 21 "	3871311 N
Longitude	-106.69041188	-106 E 41 ' 25 "	345688 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all fish from the site were removed from foil and composited into a jar for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	59.0 grams	**Collection Date**	07/20/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 - MRG at Los Lunas - above Main Street Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81456722	34 N 48 ' 52 "	3853814 N
Longitude	-106.71109672	-106 E 42 ' 39 "	343500 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	64.5 grams	**Collection Date**	07/19/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM. above Sevilleta NWR portion of Rio		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**

Latitude	34.31230631	34 N 18 ' 44 "	3798341 N
Longitude	-106.854025	-106 E 51 ' 14 "	329403 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	40.3 grams	**Collection Date**	07/18/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 - MRG near San Antonio, NM north of Bosque del Apache NWR border		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	33.87452474	33 N 52 ' 28 "	3749783 N
Longitude	-106.84959324	-106 E 50 ' 58 "	328932 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	63.0 grams	**Collection Date**	07/16/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG at Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32125645	35 N 19 ' 16 "	3909784 N
Longitude	-106.55834077	-106 E 33 ' 30 "	358351 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*

N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	53.6 grams	**Collection Date**	07/15/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM above Alameda Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20892995	35 N 12 ' 32 "	3897412 N
Longitude	-106.61790636	-106 E 37 ' 4 "	352733 E

Coordinate Method	UTM Coordinates

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 4-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	189.9 grams	**Collection Date**	04/13/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 - MRG at Los Padillas (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.01396395	35 N 0 ' 50 "	3875872 N
Longitude	-106.67395835	-106 E 40 ' 26 "	347267 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	28.3 grams	**Collection Date**	07/10/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4- MRG near Los Lunas (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81916416	34 N 49 ' 8 "	3854327 N
Longitude	-106.713116	-106 E 42 ' 47 "	343324 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*

N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-08-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	76.3 grams	**Collection Date**	01/23/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31200231	34 N 18 ' 43 "	3798307 N
Longitude	-106.85385532	-106 E 51 ' 13 "	329418 E

Coordinate Method	UTM Coordinates

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 10-07-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	25.7 grams	**Collection Date**	10/31/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.3125136	35 N 18 ' 45 "	3908815 N
Longitude	-106.55884398	-106 E 33 ' 31 "	358290 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	26.7 grams	**Collection Date**	10/30/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG at Alameda, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20886524	35 N 12 ' 31 "	3897405 N
Longitude	-106.61802592	-106 E 37 ' 4 "	352722 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	43.3 grams	**Collection Date**	10/29/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas, NM (above I-25 Bridge)		

FWS Refuge	N/A

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.9735791	34 N 58 ' 24 "	3871418 N
Longitude	-106.6903441	-106 E 41 ' 25 "	345696 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	39.8 grams	**Collection Date**	10/25/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM above main st bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81863921	34 N 49 ' 7 "	3854269 N
Longitude	-106.7132582	-106 E 42 ' 47 "	343310 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	46.9 grams	**Collection Date**	10/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31200231	34 N 18 ' 43 "	3798307 N
Longitude	-106.85385532	-106 E 51 ' 13 "	329418 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	29.0 grams	**Collection Date**	10/23/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio NM (above BdANWR border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87600472	33 N 52 ' 33 "	3749947 N
Longitude	-106.84951707	-106 E 50 ' 58 "	328942 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 1-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	31.9 grams	**Collection Date**	01/30/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.31245022	35 N 18 ' 44 "	3908808 N
Longitude	-106.55886476	-106 E 33 ' 31 "	358288 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	52.6 grams	**Collection Date**	01/28/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM above Alameda Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's

Latitude	35.20892167	35 N 12 ' 32 "	3897411 N
Longitude	-106.61785127	-106 E 37 ' 4 "	352738 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	50.3 grams	**Collection Date**	01/28/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20786166	35 N 12 ' 28 "	3897411 N
Longitude	-106.69530751	-106 E 41 ' 43 "	345685 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	47.1 grams	**Collection Date**	01/24/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81820555	34 N 49 ' 5 "	3854220 N
Longitude	-106.71268071	-106 E 42 ' 45 "	343362 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	55.9 grams	**Collection Date**	01/22/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (just above BdANWR north border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87474141	33 N 52 ' 29 "	3749807 N
Longitude	-106.84957629	-106 E 50 ' 58 "	328934 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis

Metals Scan

Sample Number - 4-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	33.5 grams	**Collection Date**	04/14/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32108775	35 N 19 ' 15 "	3909765 N
Longitude	-106.55813952	-106 E 33 ' 29 "	358369 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 10-06-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	77.8 grams	**Collection Date**	10/12/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM (above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21089567	35 N 12 ' 39 "	3897432 N
Longitude	-106.61757152	-106 E 37 ' 3 "	352764 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	79.0 grams	**Collection Date**	10/07/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas (above I-25 Bridge)		

FWS Refuge	N/A

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.95743122	34 N 57 ' 26 "	3869618 N
Longitude	-106.68450383	-106 E 41 ' 4 "	346199 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
Necropsy fish composite - Individual health observations available. Gonads were removed. Some fluids were likely lost also.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize all minnow in sample before analyses. Note that priority for analysis is 209 PCB congeners, then OC + PCB aroclors, then dioxin/furan scan.

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)

PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	122.0 grams	**Collection Date**	10/16/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81717205	34 N 49 ' 1 "	3854105 N
Longitude	-106.71244066	-106 E 42 ' 44 "	343382 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil

and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	110.2 grams	**Collection Date**	10/05/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.31230631	34 N 18 ' 44 "	3798341 N
Longitude	-106.854025	-106 E 51 ' 14 "	329403 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 4-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A

Sample Weight	47.2 grams	**Collection Date**	04/15/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.2107287	35 N 12 ' 38 "	3897615 N
Longitude	-106.62025999	-106 E 37 ' 12 "	352522 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for

results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 4-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	134.7 grams	**Collection Date**	04/08/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 - MRG at Los Lunas, NM (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81907187	34 N 49 ' 8 "	3854317 N
Longitude	-106.71326716	-106 E 42 ' 47 "	343310 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 4-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.4 grams	**Collection Date**	04/09/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (above BdANWR border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87436858	33 N 52 ' 27 "	3749766 N
Longitude	-106.84978446	-106 E 50 ' 59 "	328914 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PBDE - Polybrominated Diphenly Ethers
PCB Quantification of Individual Aroclors
Inorganic Analysis
Metals Scan

Sample Number - 7-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	16.1 grams	**Collection Date**	07/14/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3- MRG at Los Padillas, NM above I-25 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.96696085	34 N 58 ' 1 "	3870683 N
Longitude	-106.68972619	-106 E 41 ' 23 "	345740 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	10.5 grams	**Collection Date**	07/09/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG at La Joya, NM above Sevilleta NWR		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.29909525	34 N 17 ' 56 "	3796864 N
Longitude	-106.84696582	-106 E 50 ' 49 "	330026 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities

(These observations were not necessarily determined by a pathologist)
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	16.6 grams	**Collection Date**	07/08/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 - MRG near San Antonio, NM above Bosque del Apache NWR		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.88534008	33 N 53 ' 7 "	3750975 N

Longitude	-106.8451451	-106 E 50 ' 42 "	329365 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

N/A

Inorganic Analysis

Metals Scan

Methyl Mercury

Sample Number - 10-06-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	38.5 grams	**Collection Date**	10/11/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21213359	35 N 12 ' 43 "	3897763 N
Longitude	-106.61498184	-106 E 36 ' 53 "	353005 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 4-07-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	49.2 grams	**Collection Date**	04/26/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG near Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.31455303	35 N 18 ' 52 "	3909042 N
Longitude	-106.5594001	-106 E 33 ' 33 "	358243 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		

Family	Cyprinidae
Genus / Kingdom	Hybognathus / Animalia
Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	46.7 grams	**Collection Date**	04/25/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG near Alameda, NM (in side channel above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21174211	35 N 12 ' 42 "	3897723 N
Longitude	-106.61729205	-106 E 37 ' 2 "	352794 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis

Metals Scan

Sample Number - 4-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.9 grams	**Collection Date**	04/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas, NM (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.97036675	34 N 58 ' 13 "	3871061 N
Longitude	-106.68988377	-106 E 41 ' 23 "	345732 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	53.1 grams	**Collection Date**	04/19/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM (above Main Street Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81798048	34 N 49 ' 4 "	3854193 N
Longitude	-106.71137504	-106 E 42 ' 40 "	343481 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	29.9 grams	**Collection Date**	04/18/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM (above Sevilleta NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31455461	34 N 18 ' 52 "	3798592 N
Longitude	-106.85500898	-106 E 51 ' 18 "	329317 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 1-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	36.5 grams	**Collection Date**	01/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG near Alameda, NM (above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20905205	35 N 12 ' 32 "	3897433 N
Longitude	-106.62292904	-106 E 37 ' 22 "	352276 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	95.3 grams	**Collection Date**	01/23/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG near Los Padillas, NM (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.21174211	35 N 12 ' 42 "	3897723 N
Longitude	-106.61729205	-106 E 37 ' 2 "	352794 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 2-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	117.5 grams	**Collection Date**	02/01/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG near Los Lunas, NM (above Main Street Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81427857	34 N 48 ' 51 "	3853783 N
Longitude	-106.7117467	-106 E 42 ' 42 "	343440 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Methyl Mercury

Sample Number - 1-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	42.7 grams	**Collection Date**	01/31/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM (above Sevilleta NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.30541029	34 N 18 ' 19 "	3797569 N
Longitude	-106.84973359	-106 E 50 ' 59 "	329784 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-07-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	118.0 grams	**Collection Date**	01/30/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (above Bosque del Apache NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87900195	33 N 52 ' 44 "	3750278 N
Longitude	-106.84868442	-106 E 50 ' 55 "	329025 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		

Genus / Kingdom	Hybognathus / Animalia
Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 06385810

Sample Matrix	Water	**NWHC Case Number**	N/A
Sample Weight	N/A grams	**Collection Date**	06/25/09
Sample Volume	1000.0 ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Survey
Location	San Jose Drain		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.04416667	35 N 2 ' 39 "	3879184 N
Longitude	-106.64944444	-106 E 38 ' 58 "	349559 E
Coordinate Method		Latitude/Longitude Degrees	
Latitude/Longitude Method		GPS Unit - NAD 83 Reference System	

Specimen Information

Sex	N/A	**Age**	N/A
Egg Condition	N/A		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
Water sample collected with teflon nozzle intregated sampler using rod. Added to teflon churn, homogenized then split, unfiltered into container. Sample contains suspended sediment. Kept chilled and shipped chilled for chemical analysis.

Field Preparation Comments
Water sample collected with teflon nozzle intregated sampler using rod. Added to teflon churn, homogenized then split, unfiltered into 1 liter HDPE container. Sample contains suspended sediment. Kept chilled and shipped chilled for chemical analysis.
Analysis Requested Comments
Water sample contains suspended sediment, please homogenize before analysis.

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
Metals Scan

ECDMS Complete Catalog - 2020128

08/26/09

Catalog Submitter

Catalog Title	Rio Grande Silvery Minnow Health Study		
Catalog Submitter	Lusk, Joel		
Submitter Title	Senior Environmental Contaminant Specialist		
Location	New Mexico Ecological Services Field Office, NM		
Submitters Email	joel_lusk@fws.gov		
Phone Number	(505) 761-4709 ext. 109	**Fax Number**	505-346-2542

Catalog Information

Number of Samples	45	**Date Submitted**	08/10/09
Disposition of Samples	Return	**Catalog Validation**	Validated
Chain of Custody	No	**Fields Missing**	0
Submitter has Split Samples	No	**Non-Routine Request**	No
Request Permanent Lock	No	**Rapid Turnaround**	No
Catalog Data Lock	Catalog and sample data are unlocked.		
Results Data Lock	There are no results for this catalog.		

Project Information

DEQ Project ID	N/A	**Regional Study ID**	N/A

FWS Refuge Names

The results of this catalog are directly applicable to the following FWS refuges:
Bosque Del Apache National Wildlife Refuge , New Mexico
Sevilleta National Wildlife Refuge , New Mexico

Catalog Description
Catalog Instructions
The priority where samples have multiple chemical analyses requested is 209 PCB by HRGC/HRMS, lipids, organochlorine pesticides, heavy metals (and As, Hg, Se), PBDEs, dioxins, aromatics and then other analytes. Note that the fish was dissected and stored in a freezer for up to 2 years, and therefore, moisture content has likely largely been lost and is NOT a priority for analysis although this "analyte" has been requested.

Cost Code Information

Priority	Cost Code	Dollars Available

1	09-22420-1946-2013	$40,600
	Total	$40,600

Regional Approval

Region	Current Regional Coordinator	Approval Date	Phone Number	E-Mail Address
2	Laila Lienesch	08/14/09	505-248-6494 ext.	laila_lienesch@fws.gov

Catalog Status Information

Purchase Order Number	Lab Name	Analysis Type	PO Generated	Lab Approval	Current *PCD	QA/QC Approval	Results Location

Catalog Cost

Purchase Order Number	Lab Name	Analysis Type	Cost Code	Cost

Sample Number - 7-06-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	79.6 grams	**Collection Date**	07/27/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG near Bernalillo, NM above and below 550 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32121025	35 N 19 ' 16 "	3909779 N
Longitude	-106.55842789	-106 E 33 ' 30 "	358343 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	83.6 grams	**Collection Date**	07/26/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site2 - MRG at Alameda NM, below AMAFCA NDC		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20897414	35 N 12 ' 32 "	3897417 N
Longitude	-106.61797315	-106 E 37 ' 4 "	352727 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

PBDE - Polybrominated Diphenly Ethers

Inorganic Analysis

Metals Scan

Sample Number - 7-06-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.9 grams	**Collection Date**	07/25/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	SIte3-MRG at Los Padillas, NM above I-25 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.97261343	34 N 58 ' 21 "	3871311 N
Longitude	-106.69041188	-106 E 41 ' 25 "	345688 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all fish from the site were removed from foil and composited into a jar for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	59.0 grams	**Collection Date**	07/20/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 - MRG at Los Lunas - above Main Street Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81456722	34 N 48 ' 52 "	3853814 N
Longitude	-106.71109672	-106 E 42 ' 39 "	343500 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	64.5 grams	**Collection Date**	07/19/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM. above Sevilleta NWR portion of Rio		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**

Latitude	34.31230631	34 N 18 ' 44 "	3798341 N
Longitude	-106.854025	-106 E 51 ' 14 "	329403 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-06-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	40.3 grams	**Collection Date**	07/18/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 - MRG near San Antonio, NM north of Bosque del Apache NWR border		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	33.87452474	33 N 52 ' 28 "	3749783 N
Longitude	-106.84959324	-106 E 50 ' 58 "	328932 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	63.0 grams	**Collection Date**	07/16/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG at Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32125645	35 N 19 ' 16 "	3909784 N
Longitude	-106.55834077	-106 E 33 ' 30 "	358351 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*

N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	53.6 grams	**Collection Date**	07/15/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM above Alameda Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20892995	35 N 12 ' 32 "	3897412 N
Longitude	-106.61790636	-106 E 37 ' 4 "	352733 E

Coordinate Method	UTM Coordinates

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 4-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	189.9 grams	**Collection Date**	04/13/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 - MRG at Los Padillas (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.01396395	35 N 0 ' 50 "	3875872 N
Longitude	-106.67395835	-106 E 40 ' 26 "	347267 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 7-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	28.3 grams	**Collection Date**	07/10/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4- MRG near Los Lunas (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81916416	34 N 49 ' 8 "	3854327 N
Longitude	-106.713116	-106 E 42 ' 47 "	343324 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*

N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-08-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	76.3 grams	**Collection Date**	01/23/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31200231	34 N 18 ' 43 "	3798307 N
Longitude	-106.85385532	-106 E 51 ' 13 "	329418 E

Coordinate Method	UTM Coordinates

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 10-07-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	25.7 grams	**Collection Date**	10/31/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 - MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.3125136	35 N 18 ' 45 "	3908815 N
Longitude	-106.55884398	-106 E 33 ' 31 "	358290 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	26.7 grams	**Collection Date**	10/30/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG at Alameda, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20886524	35 N 12 ' 31 "	3897405 N
Longitude	-106.61802592	-106 E 37 ' 4 "	352722 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	43.3 grams	**Collection Date**	10/29/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas, NM (above I-25 Bridge)		

FWS Refuge	N/A

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.9735791	34 N 58 ' 24 "	3871418 N
Longitude	-106.6903441	-106 E 41 ' 25 "	345696 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	39.8 grams	**Collection Date**	10/25/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM above main st bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81863921	34 N 49 ' 7 "	3854269 N
Longitude	-106.7132582	-106 E 42 ' 47 "	343310 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	46.9 grams	**Collection Date**	10/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31200231	34 N 18 ' 43 "	3798307 N
Longitude	-106.85385532	-106 E 51 ' 13 "	329418 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-07-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	29.0 grams	**Collection Date**	10/23/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio NM (above BdANWR border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87600472	33 N 52 ' 33 "	3749947 N
Longitude	-106.84951707	-106 E 50 ' 58 "	328942 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 1-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	31.9 grams	**Collection Date**	01/30/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.31245022	35 N 18 ' 44 "	3908808 N
Longitude	-106.55886476	-106 E 33 ' 31 "	358288 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	52.6 grams	**Collection Date**	01/28/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM above Alameda Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's

Latitude	35.20892167	35 N 12 ' 32 "	3897411 N
Longitude	-106.61785127	-106 E 37 ' 4 "	352738 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	50.3 grams	**Collection Date**	01/28/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20786166	35 N 12 ' 28 "	3897411 N
Longitude	-106.69530751	-106 E 41 ' 43 "	345685 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	47.1 grams	**Collection Date**	01/24/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81820555	34 N 49 ' 5 "	3854220 N
Longitude	-106.71268071	-106 E 42 ' 45 "	343362 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis
Metals Scan

Sample Number - 1-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	55.9 grams	**Collection Date**	01/22/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (just above BdANWR north border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87474141	33 N 52 ' 29 "	3749807 N
Longitude	-106.84957629	-106 E 50 ' 58 "	328934 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
Aliphatic Scan
Aromatic Scan
Inorganic Analysis

Metals Scan

Sample Number - 4-08-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	33.5 grams	**Collection Date**	04/14/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.32108775	35 N 19 ' 15 "	3909765 N
Longitude	-106.55813952	-106 E 33 ' 29 "	358369 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 10-06-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	77.8 grams	**Collection Date**	10/12/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda, NM (above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21089567	35 N 12 ' 39 "	3897432 N
Longitude	-106.61757152	-106 E 37 ' 3 "	352764 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	79.0 grams	**Collection Date**	10/07/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas (above I-25 Bridge)		

FWS Refuge	N/A

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.95743122	34 N 57 ' 26 "	3869618 N
Longitude	-106.68450383	-106 E 41 ' 4 "	346199 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
Necropsy fish composite - Individual health observations available. Gonads were removed. Some fluids were likely lost also.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize all minnow in sample before analyses. Note that priority for analysis is 209 PCB congeners, then OC + PCB aroclors, then dioxin/furan scan.

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)

PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	122.0 grams	**Collection Date**	10/16/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.81717205	34 N 49 ' 1 "	3854105 N
Longitude	-106.71244066	-106 E 42 ' 44 "	343382 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil

and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 10-06-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	110.2 grams	**Collection Date**	10/05/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG near La Joya, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.31230631	34 N 18 ' 44 "	3798341 N
Longitude	-106.854025	-106 E 51 ' 14 "	329403 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 4-08-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A

Sample Weight	47.2 grams	**Collection Date**	04/15/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 - MRG at Alameda		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.2107287	35 N 12 ' 38 "	3897615 N
Longitude	-106.62025999	-106 E 37 ' 12 "	352522 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for

results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 4-08-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	134.7 grams	**Collection Date**	04/08/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 - MRG at Los Lunas, NM (above Main St Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81907187	34 N 49 ' 8 "	3854317 N
Longitude	-106.71326716	-106 E 42 ' 47 "	343310 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
PBDE - Polybrominated Diphenly Ethers
Inorganic Analysis
Metals Scan

Sample Number - 4-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.4 grams	**Collection Date**	04/09/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (above BdANWR border)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87436858	33 N 52 ' 27 "	3749766 N
Longitude	-106.84978446	-106 E 50 ' 59 "	328914 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PBDE - Polybrominated Diphenly Ethers
PCB Quantification of Individual Aroclors
Inorganic Analysis
Metals Scan

Sample Number - 7-08-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	16.1 grams	**Collection Date**	07/14/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3- MRG at Los Padillas, NM above I-25 Bridge		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.96696085	34 N 58 ' 1 "	3870683 N
Longitude	-106.68972619	-106 E 41 ' 23 "	345740 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	10.5 grams	**Collection Date**	07/09/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 - MRG at La Joya, NM above Sevilleta NWR		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.29909525	34 N 17 ' 56 "	3796864 N
Longitude	-106.84696582	-106 E 50 ' 49 "	330026 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities

(These observations were not necessarily determined by a pathologist)
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 7-08-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	16.6 grams	**Collection Date**	07/08/08
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 - MRG near San Antonio, NM above Bosque del Apache NWR		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.88534008	33 N 53 ' 7 "	3750975 N

Longitude	-106.8451451	-106 E 50 ' 42 "	329365 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities
(These observations were not necessarily determined by a pathologist)

N/A

Sample Comments

These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments

Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.

Analysis Requested Comments

Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis

N/A

Inorganic Analysis

Metals Scan

Methyl Mercury

Sample Number - 10-06-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	38.5 grams	**Collection Date**	10/11/06
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 1 MRG at Bernalillo, NM		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21213359	35 N 12 ' 43 "	3897763 N
Longitude	-106.61498184	-106 E 36 ' 53 "	353005 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Dioxin & Furan Full Scan
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
N/A

Sample Number - 4-07-1

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	49.2 grams	**Collection Date**	04/26/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Sandoval	**Site Classification**	Monitoring
Location	Site 1 MRG near Bernalillo, NM (above and below 550 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.31455303	35 N 18 ' 52 "	3909042 N
Longitude	-106.5594001	-106 E 33 ' 33 "	358243 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		

Family	Cyprinidae
Genus / Kingdom	Hybognathus / Animalia
Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	46.7 grams	**Collection Date**	04/25/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG near Alameda, NM (in side channel above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.21174211	35 N 12 ' 42 "	3897723 N
Longitude	-106.61729205	-106 E 37 ' 2 "	352794 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis

Metals Scan

Sample Number - 4-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	32.9 grams	**Collection Date**	04/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG at Los Padillas, NM (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.97036675	34 N 58 ' 13 "	3871061 N
Longitude	-106.68988377	-106 E 41 ' 23 "	345732 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	53.1 grams	**Collection Date**	04/19/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG at Los Lunas, NM (above Main Street Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81798048	34 N 49 ' 4 "	3854193 N
Longitude	-106.71137504	-106 E 42 ' 40 "	343481 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 4-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	29.9 grams	**Collection Date**	04/18/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM (above Sevilleta NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.31455461	34 N 18 ' 52 "	3798592 N
Longitude	-106.85500898	-106 E 51 ' 18 "	329317 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Sample Number - 1-07-2

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	36.5 grams	**Collection Date**	01/24/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 2 MRG near Alameda, NM (above Alameda Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.20905205	35 N 12 ' 32 "	3897433 N
Longitude	-106.62292904	-106 E 37 ' 22 "	352276 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in

foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-07-3

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	95.3 grams	**Collection Date**	01/23/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Monitoring
Location	Site 3 MRG near Los Padillas, NM (above I-25 Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	35.21174211	35 N 12 ' 42 "	3897723 N
Longitude	-106.61729205	-106 E 37 ' 2 "	352794 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		

Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 2-07-4

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	117.5 grams	**Collection Date**	02/01/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Valencia	**Site Classification**	Monitoring
Location	Site 4 MRG near Los Lunas, NM (above Main Street Bridge)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	34.81427857	34 N 48 ' 51 "	3853783 N
Longitude	-106.7117467	-106 E 42 ' 42 "	343440 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan

Methyl Mercury

Sample Number - 1-07-5

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	42.7 grams	**Collection Date**	01/31/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 5 MRG near La Joya, NM (above Sevilleta NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	**Deg./Min./Sec.**	**UTM's**
Latitude	34.30541029	34 N 18 ' 19 "	3797569 N
Longitude	-106.84973359	-106 E 50 ' 59 "	329784 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		
Genus / Kingdom	Hybognathus / Animalia		
Species	Hybognathus amarus		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 1-07-6

Sample Matrix	Partial Carcass	**NWHC Case Number**	N/A
Sample Weight	118.0 grams	**Collection Date**	01/30/07
Sample Volume	N/A ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Socorro	**Site Classification**	Monitoring
Location	Site 6 MRG near San Antonio, NM (above Bosque del Apache NWR)		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	33.87900195	33 N 52 ' 44 "	3750278 N
Longitude	-106.84868442	-106 E 50 ' 55 "	329025 E
Coordinate Method		UTM Coordinates	

Specimen Information

Sex	Mixed (composite)	**Age**	Adult
Egg Condition	N/A		
Taxonomical Group	Fish		
Common Name	Rio Grande Silvery Minnow		
Family	Cyprinidae		

Genus / Kingdom	Hybognathus / Animalia
Species	Hybognathus amarus

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
These Rio Grande silvery minnow were seined from the Rio Grande at various sites and dates. They were held in a live well until necropsy. After being subdued by a blow to the head, individual health observations were made and recorded elsewhere. The gonads were removed for histopathology. During the necropsy, these organs as well as various fluids were lost from the sample. The remaining innards were tucked back in and each individual fish was wrapped in aluminum foil and stored in freezer for up to 2 years. Up to 30 fish (or more) were then composited into a labeled jar and shipped frozen for chemical analyses.

Field Preparation Comments
Seine collection, kept alive, subdued by blow to head, dissected for observations, gonads removed, individual frozen in foil, then all from site removed and composited for chemical analyses.
Analysis Requested Comments
Please homogenize and composite all minnow in a sample. For those samples with multiple analyses, the priority for results is the 209 PCB by HRGC/HRMS, then the Organochlorine scan (with determination of percent lipids is Priority 2), then determination of PBDE and/or dioxin/furan, then the Metal Scan and/or methyl mercury is Priority 4 (moisture content is also requested but is not a priority).

Analysis Requested

Organic Analysis
N/A
Inorganic Analysis
Metals Scan
Methyl Mercury

Sample Number - 06385810

Sample Matrix	Water	**NWHC Case Number**	N/A
Sample Weight	N/A grams	**Collection Date**	06/25/09
Sample Volume	1000.0 ml.	**Composite Sample**	Yes
State	New Mexico	**USGS Watershed**	13020203
County	Bernalillo	**Site Classification**	Survey
Location	San Jose Drain		
FWS Refuge	N/A		

Site Coordinates

	Decimal Degrees	Deg./Min./Sec.	UTM's
Latitude	35.04416667	35 N 2 ' 39 "	3879184 N
Longitude	-106.64944444	-106 E 38 ' 58 "	349559 E
Coordinate Method		Latitude/Longitude Degrees	
Latitude/Longitude Method		GPS Unit - NAD 83 Reference System	

Specimen Information

Sex	N/A	**Age**	N/A
Egg Condition	N/A		

Specimen Abnormalities *(These observations were not necessarily determined by a pathologist)*
N/A

Sample Comments
Water sample collected with teflon nozzle intregated sampler using rod. Added to teflon churn, homogenized then split, unfiltered into container. Sample contains suspended sediment. Kept chilled and shipped chilled for chemical analysis.

Field Preparation Comments
Water sample collected with teflon nozzle intregated sampler using rod. Added to teflon churn, homogenized then split, unfiltered into 1 liter HDPE container. Sample contains suspended sediment. Kept chilled and shipped chilled for chemical analysis.
Analysis Requested Comments
Water sample contains suspended sediment, please homogenize before analysis.

Analysis Requested

Organic Analysis
209 PCB by HRGC/HRMS (EPA 1668a)
Organochlorine Scan (OC)
PCB Quantification of Individual Aroclors
Inorganic Analysis
Metals Scan

Appendix 3D. Moisture content and concentrations of elements and methyl mercury in Rio Grande silvery minnow carcasses from all sites expressed as µg/g dry weight. ["Sample ID", sample identification number, includes month and year sampled and site number; "µg/g", micrograms per gram; "<", less than laboratory method detection limit; see Table 2 for chemical names and abbreviations; see Table 1 for site number, descripton and location]

Sample ID	Collection Date	Site Number	Moisture Percent	Al µg/g	As µg/g
7-06-1	7/27/2006	1	73.2	1150	3.27
4-07-1	4/26/2007	1	74.1	382	3.02
1-08-1	1/30/2008	1	75.2	140	4.49
4-08-1	4/16/2008	1	73.0	383	3.41
7-08-1	7/16/2008	1	71.6	542	4.10
7-06-2	7/26/2006	2	72.5	970	3.08
1-07-2	1/24/2007	2	76.2	141	3.50
4-07-2	4/25/2007	2	75.3	429	3.29
1-08-2	1/29/2008	2	74.7	162	3.92
4-08-2	4/15/2008	2	72.0	317	3.70
7-08-2	7/15/2008	2	71.5	419	3.75
7-06-3	7/25/2006	3	71.5	767	2.68
1-07-3	1/23/2007	3	76.6	96	3.43
4-07-3	4/24/2007	3	74.0	256	2.74
1-08-3	1/28/2008	3	73.2	95	3.77
4-08-3	4/14/2008	3	71.0	111	2.67
7-08-3	7/14/2008	3	72.8	81	2.83
7-06-4	7/20/2006	4	66.5	626	2.68
2-07-4	2/1/2007	4	72.2	61	3.13
4-07-4	4/19/2007	4	74.3	450	3.09
1-08-4	1/24/2008	4	72.6	142	2.70
4-08-4	4/8/2008	4	73.0	214	2.78
7-08-4	7/10/2008	4	73.5	394	3.43
7-06-5	7/19/2006	5	68.8	328	2.80
1-07-5	1/31/2007	5	72.2	109	2.96
4-07-5	4/18/2007	5	73.5	201	2.79
1-08-5	1/23/2008	5	72.7	36	3.10
7-08-5	7/9/2008	5	71.5	205	3.20
7-06-6	7/18/2006	6	69.8	289	2.55
1-07-6	1/30/2007	6	72.9	127	3.26
1-08-6	1/22/2008	6	70.7	73	2.73
4-08-6	4/9/2008	6	73.3	95	2.67
7-08-6	7/8/2008	6	71.3	31	2.80

Appendix 3D. Moisture content and concentrations of elements and methyl mercury in Rio Grande silvery minnow carcasses from all sites expressed as µg/g dry weight. ["Sample ID", sample identification number, includes month and year sampled and site number; "µg/g", micrograms per gram; "<", less than laboratory method detection limit; see Table 2 for chemical names and abbreviations; see Table 1 for site number, descripton and location]

Sample ID	B µg/g	Ba µg/g	Be µg/g	Cd µg/g	Cr µg/g	Cu µg/g	Fe µg/g
7-06-1	1.050	88.0	0.0578	0.1150	3.730	31.70	1180
4-07-1	< 0.511	79.0	< 0.0511	< 0.0193	0.618	3.49	457
1-08-1	< 0.519	88.4	< 0.0519	0.0627	0.753	3.78	149
4-08-1	< 0.509	69.5	< 0.0509	0.0436	0.925	3.34	408
7-08-1	0.562	71.4	< 0.0468	0.0776	< 0.468	3.72	749
7-06-2	0.705	74.3	< 0.0517	0.0542	2.630	18.20	1060
1-07-2	< 0.462	79.3	< 0.0462	0.0695	< 0.462	3.30	203
4-07-2	< 0.513	70.0	< 0.0513	< 0.0194	0.783	3.25	496
1-08-2	< 0.508	69.3	< 0.0508	0.0359	1.230	46.70	181
4-08-2	< 0.511	67.7	< 0.0511	< 0.0193	0.662	3.17	344
7-08-2	< 0.463	67.6	< 0.0463	0.0230	< 0.463	3.19	574
7-06-3	0.651	59.8	< 0.0500	< 0.0189	2.580	3.28	838
1-07-3	< 0.431	68.0	< 0.0431	0.0371	< 0.431	3.20	151
4-07-3	< 0.512	65.2	< 0.0512	< 0.0193	0.513	2.79	290
1-08-3	< 0.520	62.1	< 0.0520	< 0.0196	< 0.520	3.47	125
4-08-3	< 0.508	49.8	< 0.0508	< 0.0192	< 0.508	20.00	141
7-08-3	< 0.484	47.5	< 0.0484	< 0.0097	< 0.484	2.56	130
7-06-4	0.576	65.3	< 0.0524	< 0.0198	2.060	2.81	634
2-07-4	< 0.434	47.8	< 0.0434	< 0.0087	< 0.434	2.67	106
4-07-4	< 0.507	59.0	< 0.0507	< 0.0191	0.752	3.57	511
1-08-4	< 0.508	56.1	< 0.0508	< 0.0192	< 0.508	3.76	178
4-08-4	< 0.512	55.3	< 0.0512	< 0.0193	0.602	14.90	233
7-08-4	< 0.449	64.2	< 0.0449	< 0.0090	< 0.449	3.07	590
7-06-5	< 0.513	54.4	< 0.0513	< 0.0194	0.993	2.31	344
1-07-5	0.477	22.1	< 0.0425	< 0.0086	< 0.425	2.69	162
4-07-5	0.758	29.7	< 0.0493	< 0.0099	< 0.493	3.00	294
1-08-5	< 0.459	34.7	< 0.0459	< 0.0092	< 0.459	2.89	66
7-08-5	< 0.470	42.0	< 0.0470	< 0.0094	< 0.470	3.22	314
7-06-6	< 0.521	57.6	< 0.0521	< 0.0197	0.728	2.32	309
1-07-6	< 0.439	37.3	< 0.0439	< 0.0088	< 0.439	2.58	178
1-08-6	< 0.502	22.1	< 0.0502	< 0.0190	< 0.502	6.91	101
4-08-6	< 0.506	30.6	< 0.0506	< 0.0191	< 0.506	3.01	163
7-08-6	< 0.456	26.3	< 0.0456	< 0.0091	< 0.456	2.78	69

Appendix 3D. Moisture content and concentrations of elements and methyl mercury in Rio Grande silvery minnow carcasses from all sites expressed as µg/g dry weight. ["Sample ID", sample identification number, includes month and year sampled and site number; "µg/g", micrograms per gram; "<", less than laboratory method detection limit; see Table 2 for chemical names and abbreviations; see Table 1 for site number, descripton and location]

Sample ID	Hg µg/g	MeHg µg/g	Mg µg/g	Mn µg/g	Mo µg/g	Ni µg/g	Pb µg/g
7-06-1	0.277		2000	58.2	< 1.04	13.30	1.120
4-07-1	0.282		1730	32.4	< 1.02	1.24	0.376
1-08-1	0.150		1560	17.8	< 1.04	0.74	0.152
4-08-1	0.129		1410	27.1	< 1.02	1.25	0.298
7-08-1	0.192	0.181	1570	56.6	< 0.94	0.90	0.959
7-06-2	0.273		1700	52.4	< 1.03	12.90	1.470
1-07-2	0.315	0.221	1550	24.0	< 0.92	0.47	0.416
4-07-2	0.251		1570	34.1	< 1.03	1.15	0.522
1-08-2	0.146		1440	17.0	< 1.02	7.50	0.291
4-08-2	0.123		1470	22.9	< 1.02	0.95	0.289
7-08-2	0.195	0.197	1440	62.8	< 0.93	0.87	0.722
7-06-3	0.205		1780	39.9	< 1.00	2.24	0.763
1-07-3	0.296	0.241	1590	16.4	< 0.86	0.54	0.395
4-07-3	0.202		1390	24.3	< 1.02	0.92	0.290
1-08-3	0.125		1400	10.7	< 1.04	0.86	0.166
4-08-3	0.106		1310	13.0	< 1.02	4.58	0.305
7-08-3	0.136	0.143	1300	14.8	< 0.97	< 0.48	0.343
7-06-4	0.181		1690	36.8	< 1.05	1.87	0.668
2-07-4	0.141	0.141	1330	12.5	< 0.87	< 0.43	0.326
4-07-4	0.161		1710	28.4	< 1.01	1.26	0.557
1-08-4	0.086		1390	12.9	< 1.02	0.69	0.191
4-08-4	0.111		1460	17.2	< 1.02	3.27	0.293
7-08-4	0.219	0.199	1520	35.6	< 0.90	0.87	0.908
7-06-5	0.185		1640	49.9	< 1.03	1.10	0.392
1-07-5	0.198	0.164	1150	16.3	< 0.85	0.47	0.272
4-07-5	0.216		1360	35.1	< 0.99	< 0.49	0.562
1-08-5	0.102	0.100	1280	13.1	< 0.92	< 0.46	0.214
7-08-5	0.204	0.221	1470	38.8	< 0.94	0.62	0.435
7-06-6	0.232		1530	31.7	< 1.04	1.19	0.338
1-07-6	0.258	0.256	1290	15.6	< 0.88	< 0.44	0.290
1-08-6	0.102		1240	11.8	< 1.00	3.45	0.115
4-08-6	0.120		1450	15.8	< 1.01	0.64	0.120
7-08-6	0.357	0.360	1260	14.0	< 0.91	< 0.46	0.136

Appendix 3D. Moisture content and concentrations of elements and methyl mercury in Rio Grande silvery minnow carcasses from all sites expressed as µg/g dry weight. ["Sample ID", sample identification number, includes month and year sampled and site number; "µg/g", micrograms per gram; "<", less than laboratory method detection limit; see Table 2 for chemical names and abbreviations; see Table 1 for site number, descripton and location]

Sample ID	Se µg/g	Sr µg/g	V µg/g	Zn µg/g
7-06-1	2.09	162	3.15	156
4-07-1	2.00	138	1.32	140
1-08-1	2.06	154	0.55	165
4-08-1	1.36	107	1.30	136
7-08-1	1.74	120	1.47	149
7-06-2	2.42	123	2.48	140
1-07-2	2.16	176	1.26	179
4-07-2	1.66	145	1.84	135
1-08-2	2.08	144	0.91	138
4-08-2	1.81	131	1.13	122
7-08-2	1.89	105	1.22	143
7-06-3	1.74	164	2.81	147
1-07-3	2.52	147	0.85	159
4-07-3	1.31	150	1.49	140
1-08-3	1.70	147	0.71	136
4-08-3	1.68	109	0.71	119
7-08-3	1.69	104	< 0.48	135
7-06-4	1.81	154	1.74	170
2-07-4	1.81	108	< 0.43	114
4-07-4	2.07	129	1.56	126
1-08-4	1.89	121	0.79	132
4-08-4	1.53	124	0.94	127
7-08-4	1.84	121	1.35	145
7-06-5	1.42	187	1.19	146
1-07-5	3.19	143	0.67	114
4-07-5	2.33	155	0.98	125
1-08-5	2.03	134	< 0.46	112
7-08-5	1.92	128	0.71	153
7-06-6	1.40	170	1.12	165
1-07-6	1.85	134	0.66	110
1-08-6	2.84	175	0.61	131
4-08-6	1.69	186	0.79	133
7-08-6	1.92	99	< 0.46	132

Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]

Sample ID	Collection Date	Site Number	Percent Lipid	Percent Moisture	Aldrin	alpha BHC	alpha chlordane
7-06-1	7/27/2006	1	5.5	73.9			
10-06-1	10/11/2006	1	7.7	73.3	< 0.122	< 0.209	1.710
10-07-1	10/31/2007	1	5.3	71.7	< 0.122	< 0.209	0.269
1-08-1	1/30/2008	1	4.1	76.4			
4-08-1	4/16/2008	1	7.5	76.2			
7-06-2	7/26/2006	2	6.8	74.1			
10-06-2	10/12/2006	2	5.4	74.6	< 0.122	< 0.209	2.540
10-07-2	10/30/2007	2	6.9	71.4	< 0.122	< 0.209	1.360
1-08-2	1/29/2008	2	4.3	73.8			
4-08-2	4/15/2008	2	6.1	74.3			
7-06-3	7/25/2006	3	5.9	74.2			
10-06-3	10/17/2006	3	6.6	70.0	< 0.122	< 0.209	5.660
10-07-3	10/29/2007	3	8.2	70.6	< 0.122	< 0.209	9.380
1-08-3	1/28/2008	3	5.0	72.6			
4-08-3	4/14/2008	3	7.7	72.4			
7-06-4	7/20/2006	4	10.8	69.0			
10-06-4	10/16/2006	4	7.8	71.3	< 0.122	< 0.209	6.490
10-07-4	10/25/2007	4	5.7	76.4	< 0.122	< 0.209	6.750
1-08-4	1/24/2008	4	6.6	74.1			
4-08-4	4/8/2008	4	5.5	73.3			
7-06-5	7/19/2006	5	7.7	72.1			
10-06-5	10/18/2006	5	7.9	69.9	< 0.122	< 0.209	3.170
10-07-5	10/24/2007	5	5.8	73.7	< 0.122	< 0.209	3.230
1-08-5	1/23/2008	5					
7-06-6	7/18/2006	6	8.0	73.5			
10-07-6	10/23/2007	6	6.0	74.5	< 0.122	< 0.209	0.419
1-08-6	1/22/2008	6	7.5	73.3			
4-08-6	4/9/2008	6	5.6	74.8	< 0.122	< 0.209	2.180

Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]

Sample ID	cis-nonachlor	gamma chlordane	oxy-chlordane	trans-nonachlor	beta BHC	gamma BHC (Lindane)	BHC-Total
7-06-1							
10-06-1	0.973	0.856	0.305	2.29	<0.224	<0.094	<0.359
10-07-1	0.157	<0.155	0.202	0.44	<0.224	<0.094	<0.359
1-08-1							
4-08-1							
7-06-2							
10-06-2	1.160	1.230	0.251	3.44	<0.224	<0.094	<0.359
10-07-2	0.656	0.946	0.265	2.20	<0.224	<0.094	<0.359
1-08-2							
4-08-2							
7-06-3							
10-06-3	2.210	3.220	0.332	7.21	<0.224	<0.094	<0.359
10-07-3	3.440	5.460	0.493	10.70	<0.224	2.640	2.640
1-08-3							
4-08-3							
7-06-4							
10-06-4	2.770	3.280	0.520	8.05	<0.224	<0.094	<0.359
10-07-4	3.370	3.240	0.398	8.76	<0.224	0.117	<0.359
1-08-4							
4-08-4							
7-06-5							
10-06-5	1.590	1.500	0.324	3.90	<0.224	<0.094	<0.359
10-07-5	1.740	1.580	0.314	4.10	<0.224	<0.094	<0.359
1-08-5							
7-06-6							
10-07-6	0.259	0.173	<0.121	0.57	<0.224	<0.094	<0.359
1-08-6							
4-08-6	0.977	1.130	0.274	2.48	<0.224	<0.094	<0.359

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**delta BHC**	**HCB**	**Heptachlor**	**heptachlor epoxide**	**DDMU**	**dieldrin**	**endrin**	**endosulfan I**
7-06-1								
10-06-1	<0.109	0.586	<0.133	0.188	0.703	0.727	<0.202	1.590
10-07-1	<0.109	<0.226	<0.133	<0.160	<0.128	<0.189	<0.202	0.347
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.109	0.251	<0.133	<0.160	0.427	0.855	<0.202	1.660
10-07-2	<0.109	0.290	<0.133	<0.160	<0.128	0.442	<0.202	0.642
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.109	0.714	<0.133	0.306	0.638	4.150	<0.202	2.490
10-07-3	<0.109	0.805	<0.133	0.312	<0.128	2.850	<0.202	4.180
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.109	0.442	<0.133	0.429	0.754	3.820	<0.202	3.450
10-07-4	<0.109	0.363	<0.133	<0.160	0.609	2.460	<0.202	4.050
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.109	0.398	<0.133	0.324	0.221	2.780	<0.202	2.350
10-07-5	<0.109	<0.226	<0.133	0.248	0.340	1.370	<0.202	2.140
1-08-5								
7-06-6								
10-07-6	<0.109	0.308	<0.133	<0.160	1.080	0.419	<0.202	0.577
1-08-6								
4-08-6	<0.109	0.391	<0.133	<0.160	0.639	1.360	<0.202	1.270

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Sample ID	**endosulfan II**	**endosulfan sulfate**	**o,p'-DDD**	**p,p'-DDD**	**o,p'-DDE**	**p,p'-DDE**	**o,p'-DDT**
7-06-1							
10-06-1	0.234	1.410	0.586	1.780	0.410	10.900	<0.155
10-07-1	<0.148	0.427	<0.206	0.696	<0.119	7.780	<0.155
1-08-1							
4-08-1							
7-06-2							
10-06-2	<0.148	2.400	0.490	1.600	0.465	13.900	<0.155
10-07-2	<0.148	1.100	<0.206	0.833	0.164	8.020	<0.155
1-08-2							
4-08-2							
7-06-3							
10-06-3	<0.148	4.340	0.574	1.750	0.421	18.600	<0.155
10-07-3	0.312	3.220	0.865	1.950	0.396	17.400	<0.155
1-08-3							
4-08-3							
7-06-4							
10-06-4	<0.148	4.600	0.832	2.950	0.741	24.600	<0.155
10-07-4	<0.148	4.320	1.060	2.870	0.527	22.600	<0.155
1-08-4							
4-08-4							
7-06-5							
10-06-5	0.648	2.840	0.663	1.560	0.884	13.700	<0.155
10-07-5	<0.148	1.980	0.399	1.460	0.375	8.650	<0.155
1-08-5							
7-06-6							
10-07-6	<0.148	0.481	<0.206	0.752	0.308	13.400	<0.155
1-08-6							
4-08-6	<0.148	1.520	0.300	1.060	0.209	10.600	<0.155

Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]

Sample ID	p,p'-DDT	DDT-Total	mirex	PCB-1242	PCB-1248	PCB-1254	PCB-1260
7-06-1							
10-06-1	0.234	14.60	<0.123	<0.183	<0.183	4.200	6.300
10-07-1	0.168	8.80	<0.123	0.571	1.140	2.570	1.430
1-08-1							
4-08-1							
7-06-2							
10-06-2	0.201	17.10	<0.123	<0.189	<0.189	3.330	7.770
10-07-2	0.303	9.32	<0.123	<0.197	2.520	5.050	0.841
1-08-2							
4-08-2							
7-06-3							
10-06-3	0.663	22.70	<0.123	<0.196	<0.196	4.860	11.300
10-07-3	0.673	21.20	<0.123	<0.195	7.640	26.700	3.820
1-08-3							
4-08-3							
7-06-4							
10-06-4	0.819	30.70	0.130	<0.198	<0.198	8.280	19.300
10-07-4	0.761	28.40	<0.123	<0.188	<0.188	15.400	10.200
1-08-4							
4-08-4							
7-06-5							
10-06-5	0.339	17.40	<0.123	<0.163	<0.163	4.500	10.500
10-07-5	0.290	11.50	<0.123	<0.160	<0.160	5.800	5.800
1-08-5							
7-06-6							
10-07-6	<0.166	15.50	<0.123	0.475	0.950	3.320	<0.155
1-08-6							
4-08-6	<0.166	13.00	<0.123	<0.151	<0.151	3.160	5.880

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB-TOTAL**	**PCB 1**	**PCB 10**	**PCB 103**	**PCB 104**	**PCB 105**	**PCB 106**	**PCB 107**
7-06-1								
10-06-1	10.500	<0.0367	<0.00917	<0.0917	<0.0917	0.124	<0.0917	<0.183
10-07-1	5.710	<0.0398	<0.00994	<0.0994	<0.0994	0.062	<0.0994	<0.199
1-08-1								
4-08-1								
7-06-2								
10-06-2	11.100	<0.0377	<0.00943	<0.0943	<0.0943	0.064	<0.0943	<0.189
10-07-2	8.410	<0.0393	<0.00983	<0.0983	<0.0983	0.123	<0.0983	<0.197
1-08-2								
4-08-2								
7-06-3								
10-06-3	16.200	<0.0392	<0.00980	<0.0980	<0.0980	0.112	<0.0980	<0.196
10-07-3	38.200	<0.039	<0.00976	<0.0976	<0.0976	0.628	<0.0976	<0.195
1-08-3								
4-08-3								
7-06-4								
10-06-4	27.600	<0.0395	<0.00988	<0.0988	<0.0988	0.194	<0.0988	<0.198
10-07-4	25.600	<0.0377	<0.00942	<0.0942	<0.0942	0.355	<0.0942	<0.188
1-08-4								
4-08-4								
7-06-5								
10-06-5	15.000	<0.0326	<0.00815	<0.0815	<0.0815	0.125	<0.0815	<0.163
10-07-5	11.600	<0.0320	<0.00801	<0.0801	<0.0801	0.127	<0.0801	<0.160
1-08-5								
7-06-6								
10-07-6	4.750	<0.0311	<0.00777	<0.0777	<0.0777	0.052	<0.0777	<0.155
1-08-6								
4-08-6	9.040	<0.0301	<0.00753	<0.0753	<0.0753	0.077	<0.0753	<0.151

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Sample ID	**PCB 108/124**	**PCB 11**	**PCB 110/115**	**PCB 111**	**PCB 112**	**PCB 114**	**PCB 117**	**PCB 118**
7-06-1								
10-06-1	<0.183	0.613	0.344	<0.183	<0.183	<0.0917	<0.0367	0.367
10-07-1	<0.199	0.676	<0.199	<0.199	<0.199	<0.0994	<0.0398	0.145
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.189	0.635	0.255	<0.189	<0.189	<0.0943	<0.0377	0.177
10-07-2	<0.197	0.810	0.367	<0.197	<0.197	<0.0983	<0.0393	0.333
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.196	0.659	0.434	<0.196	<0.196	<0.0980	<0.0392	0.319
10-07-3	<0.195	0.725	1.950	<0.195	<0.195	<0.0976	<0.0390	1.660
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.198	0.687	0.723	<0.198	<0.198	<0.0988	<0.0395	0.540
10-07-4	<0.188	0.653	1.120	<0.188	<0.188	<0.0942	<0.0377	0.987
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.163	0.571	0.423	<0.163	<0.163	<0.0815	<0.0326	0.337
10-07-5	<0.160	0.652	0.441	<0.160	<0.160	<0.0801	<0.0320	0.347
1-08-5								
7-06-6								
10-07-6	<0.155	0.594	0.188	<0.155	<0.155	<0.0777	<0.0311	0.146
1-08-6								
4-08-6	<0.151	0.527	0.303	<0.151	<0.151	<0.0753	<0.0301	0.235

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Sample ID	**PCB 12/13**	**PCB 120**	**PCB 121**	**PCB 122**	**PCB 123**	**PCB 126**	**PCB 127**	**PCB 128/166**
7-06-1								
10-06-1	<0.0183	<0.0917	<0.0917	<0.0917	<0.0917	<0.0917	<0.183	0.106
10-07-1	<0.0199	<0.0994	<0.0994	<0.0994	<0.0994	<0.0994	<0.199	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0189	<0.0943	<0.0943	<0.0943	<0.0943	<0.0943	<0.189	<0.0943
10-07-2	<0.0197	<0.0983	<0.0983	<0.0983	<0.0983	<0.0983	<0.197	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0196	<0.0980	<0.0980	<0.0980	<0.0980	<0.0980	<0.196	0.115
10-07-3	<0.0195	<0.0976	<0.0976	<0.0976	<0.0976	<0.0976	<0.195	0.446
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0198	<0.0988	<0.0988	<0.0988	<0.0988	<0.0988	<0.198	0.210
10-07-4	<0.0188	<0.0942	<0.0942	<0.0942	<0.0942	<0.0942	<0.188	0.293
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0163	<0.0815	<0.0815	<0.0815	<0.0815	<0.0815	<0.163	0.135
10-07-5	<0.016	<0.0801	<0.0801	<0.0801	<0.0801	<0.0801	<0.160	0.116
1-08-5								
7-06-6								
10-07-6	<0.0155	<0.0777	<0.0777	<0.0777	<0.0777	<0.0777	<0.155	<0.0777
1-08-6								
4-08-6	<0.0151	<0.0753	<0.0753	<0.0753	<0.0753	<0.0753	<0.151	0.077

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Sample ID	PCB 129/138/163	PCB 130	PCB 131	PCB 132	PCB 133	PCB 134	PCB 135/151
7-06-1							
10-06-1	0.879	<0.0917	<0.0917	0.1460	<0.0917	<0.0917	0.1640
10-07-1	0.265	<0.0990	<0.0994	<0.099	<0.0994	<0.0994	<0.0990
1-08-1							
4-08-1							
7-06-2							
10-06-2	0.887	<0.0943	<0.0943	0.2170	<0.0943	<0.0943	0.2580
10-07-2	0.650	<0.0983	<0.0983	0.1650	<0.0983	<0.0983	0.1220
1-08-2							
4-08-2							
7-06-3							
10-06-3	1.300	<0.0980	<0.0980	0.3070	<0.0980	<0.0980	0.3430
10-07-3	3.500	0.1850	<0.0976	0.8810	<0.0976	0.1220	0.6710
1-08-3							
4-08-3							
7-06-4							
10-06-4	2.370	0.1030	<0.0988	0.5290	<0.0988	<0.0988	0.5790
10-07-4	2.420	0.1240	<0.0942	0.5620	<0.0942	<0.0942	0.4320
1-08-4							
4-08-4							
7-06-5							
10-06-5	1.360	<0.0815	<0.0815	0.2900	<0.0815	<0.0815	0.2870
10-07-5	1.040	<0.0801	<0.0801	0.2460	<0.0801	<0.0801	0.2120
1-08-5							
7-06-6							
10-07-6	0.345	<0.0777	<0.0777	0.0884	<0.0777	<0.0777	0.0778
1-08-6							
4-08-6	0.769	<0.0753	<0.0753	0.1740	<0.0753	<0.0753	0.1810

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Sample ID	PCB 136	PCB 137	PCB 139/140	PCB 14	PCB 141	PCB 142	PCB 143	PCB 144
7-06-1								
10-06-1	0.0391	<0.183	<0.0917	<0.183	0.0937	<0.183	<0.0917	<0.0917
10-07-1	<0.0398	<0.199	<0.0994	<0.199	0.0448	<0.199	<0.0994	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	0.0724	<0.189	<0.0943	<0.189	0.2190	<0.189	<0.0943	<0.0943
10-07-2	0.0438	<0.197	<0.0983	<0.197	0.1130	<0.197	<0.0983	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	0.0948	<0.196	<0.0980	<0.196	0.3090	<0.196	<0.0980	<0.0980
10-07-3	0.2130	<0.195	<0.0976	<0.195	0.6490	<0.195	<0.0976	0.1090
1-08-3								
4-08-3								
7-06-4								
10-06-4	0.1520	<0.198	<0.0988	<0.198	0.5160	<0.198	<0.0988	<0.0988
10-07-4	0.1250	<0.188	<0.0942	<0.188	0.4500	<0.188	<0.0942	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	0.0744	<0.163	<0.0815	<0.163	0.2520	<0.163	<0.0815	<0.0815
10-07-5	0.0612	<0.160	<0.0801	<0.160	0.1970	<0.160	<0.0801	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0311	<0.155	<0.0777	<0.155	0.0582	<0.155	<0.0777	<0.0777
1-08-6								
4-08-6	0.0508	<0.151	<0.0753	<0.151	0.1530	<0.151	<0.0753	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 145**	**PCB 146**	**PCB 147/149**	**PCB 148**	**PCB 15**	**PCB 150**	**PCB 152**	**PCB 153/168**
7-06-1								
10-06-1	<0.183	0.1890	0.4560	<0.183	<0.0917	<0.183	<0.183	0.970
10-07-1	<0.199	<0.0994	0.1590	<0.199	<0.0994	<0.199	<0.199	0.248
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.189	0.1320	0.7050	<0.189	<0.0943	<0.189	<0.189	1.020
10-07-2	<0.197	<0.0983	0.3900	<0.197	<0.0983	<0.197	<0.197	0.571
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.196	0.1820	0.9660	<0.196	<0.0980	<0.196	<0.196	1.440
10-07-3	<0.195	0.4240	2.0900	<0.195	<0.0976	<0.195	<0.195	3.050
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.198	0.3430	1.7000	<0.198	<0.0988	<0.198	<0.198	2.590
10-07-4	<0.188	0.3100	1.4200	<0.188	<0.0942	<0.188	<0.188	2.250
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.163	0.1960	0.8430	<0.163	<0.0815	<0.163	<0.163	1.430
10-07-5	<0.160	0.1380	0.6450	<0.160	<0.0801	<0.160	<0.160	1.000
1-08-5								
7-06-6								
10-07-6	<0.155	<0.0777	0.2210	<0.155	<0.0777	<0.155	<0.155	0.351
1-08-6								
4-08-6	<0.151	0.1100	0.5220	<0.151	<0.0753	<0.151	<0.151	0.810

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 154**	**PCB 155**	**PCB 156/157**	**PCB 158**	**PCB 159**	**PCB 16**	**PCB 160**	**PCB 161**
7-06-1								
10-06-1	<0.0917	<0.183	<0.0917	0.0486	<0.183	0.0420	<0.0917	<0.183
10-07-1	<0.0994	<0.199	<0.0994	<0.0398	<0.199	0.0490	<0.0994	<0.199
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0943	<0.189	<0.0943	0.0734	<0.189	0.0338	<0.0943	<0.189
10-07-2	<0.0983	<0.197	<0.0983	0.0648	<0.197	0.0385	<0.0983	<0.197
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0980	<0.196	<0.098	0.1100	<0.196	0.0458	<0.0980	<0.196
10-07-3	<0.0976	<0.195	0.3780	0.3370	<0.195	0.0459	<0.0976	<0.195
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0988	<0.198	0.1790	0.2070	<0.198	0.0505	<0.0988	<0.198
10-07-4	<0.0942	<0.188	0.2490	0.2270	<0.188	0.0345	<0.0942	<0.188
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0815	<0.163	0.1050	0.1140	<0.163	0.0302	<0.0815	<0.163
10-07-5	<0.0801	<0.160	0.0923	0.0930	<0.160	0.0314	<0.0801	<0.160
1-08-5								
7-06-6								
10-07-6	<0.0777	<0.155	<0.0777	<0.0311	<0.155	0.0285	<0.0777	<0.155
1-08-6								
4-08-6	<0.0753	<0.151	<0.0753	0.0654	<0.151	0.0231	<0.0753	<0.151

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 162**	**PCB 164**	**PCB 165**	**PCB 167**	**PCB 169**	**PCB 17**	**PCB 170**	**PCB 171/173**
7-06-1								
10-06-1	<0.183	<0.0917	<0.183	<0.0917	<0.0917	<0.0367	0.1640	<0.183
10-07-1	<0.199	<0.0994	<0.199	<0.0994	<0.0994	<0.0398	<0.0994	<0.199
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.189	<0.0943	<0.189	<0.0943	<0.0943	<0.0377	0.3460	<0.189
10-07-2	<0.197	<0.0983	<0.197	<0.0983	<0.0983	<0.0393	0.1270	<0.197
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.196	<0.098	<0.196	<0.0980	<0.0980	<0.0392	0.5110	<0.196
10-07-3	<0.195	0.2140	<0.195	0.1380	<0.0976	0.0403	0.9670	<0.195
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.198	0.1500	<0.198	<0.0988	<0.0988	0.0438	0.9400	<0.198
10-07-4	<0.188	0.1460	<0.188	<0.0942	<0.0942	<0.0377	0.7660	<0.188
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.163	0.0862	<0.163	<0.0815	<0.0815	<0.0326	0.4710	<0.163
10-07-5	<0.160	<0.0801	<0.160	<0.0801	<0.0801	<0.0320	0.3030	<0.160
1-08-5								
7-06-6								
10-07-6	<0.155	<0.0777	<0.155	<0.0777	<0.0777	<0.0311	<0.0777	<0.155
1-08-6								
4-08-6	<0.151	<0.0753	<0.151	<0.0753	<0.0753	<0.0301	0.2430	<0.151

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 172**	**PCB 174**	**PCB 175**	**PCB 176**	**PCB 177**	**PCB 178**	**PCB 179**	**PCB 18/30**
7-06-1								
10-06-1	<0.183	0.1340	<0.183	<0.183	0.0979	<0.0917	<0.0917	<0.0917
10-07-1	<0.199	<0.0994	<0.199	<0.199	<0.0994	<0.0994	<0.0994	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.189	0.3680	<0.189	<0.189	0.1710	<0.0943	0.1080	<0.0943
10-07-2	<0.197	0.1190	<0.197	<0.197	<0.0983	<0.0983	<0.0983	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.196	0.5390	<0.196	<0.196	0.2440	<0.098	0.1460	<0.0980
10-07-3	<0.195	0.8950	<0.195	<0.195	0.4350	0.1500	0.2270	<0.0976
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.198	0.9380	<0.198	<0.198	0.5040	0.1800	0.2520	<0.0988
10-07-4	<0.188	0.6710	<0.188	<0.188	0.3830	0.1200	0.1580	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.163	0.4760	<0.163	<0.163	0.2390	0.0967	0.1270	<0.0815
10-07-5	<0.160	0.3110	<0.160	<0.160	0.1590	<0.0801	0.0815	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.155	<0.0777	<0.155	<0.155	<0.0777	<0.0777	<0.0777	<0.0777
1-08-6								
4-08-6	<0.151	0.2580	<0.151	<0.151	0.1230	<0.0753	<0.0753	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 180/193**	**PCB 181**	**PCB 182**	**PCB 183**	**PCB 184**	**PCB 185**	**PCB 186**	**PCB 187**
7-06-1								
10-06-1	0.4440	<0.183	<0.183	<0.183	<0.183	<0.183	<0.183	0.382
10-07-1	0.2220	<0.199	<0.199	<0.199	<0.199	<0.199	<0.199	0.111
1-08-1								
4-08-1								
7-06-2								
10-06-2	0.9810	<0.189	<0.189	0.251	<0.189	<0.189	<0.189	0.470
10-07-2	0.3290	<0.197	<0.197	<0.197	<0.197	<0.197	<0.197	0.157
1-08-2								
4-08-2								
7-06-3								
10-06-3	1.4000	<0.196	<0.196	0.348	<0.196	<0.196	<0.196	0.655
10-07-3	2.4200	<0.195	<0.195	0.582	<0.195	<0.195	<0.195	1.020
1-08-3								
4-08-3								
7-06-4								
10-06-4	2.6600	<0.198	<0.198	0.647	<0.198	<0.198	<0.198	1.240
10-07-4	1.9700	<0.188	<0.188	0.476	<0.188	<0.188	<0.188	0.858
1-08-4								
4-08-4								
7-06-5								
10-06-5	1.3200	<0.163	<0.163	0.324	<0.163	<0.163	<0.163	0.670
10-07-5	0.8210	<0.160	<0.160	0.208	<0.160	<0.160	<0.160	0.384
1-08-5								
7-06-6								
10-07-6	0.1800	<0.155	<0.155	<0.155	<0.155	<0.155	<0.155	0.110
1-08-6								
4-08-6	0.6820	<0.151	<0.151	0.173	<0.151	<0.151	<0.151	0.338

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 188**	**PCB 189**	**PCB 19**	**PCB 190**	**PCB 191**	**PCB 192**	**PCB 194**	**PCB 195**
7-06-1								
10-06-1	<0.0917	<0.0917	0.0284	<0.0917	<0.183	<0.183	<0.0917	<0.183
10-07-1	<0.0994	<0.0994	0.0282	<0.0994	<0.199	<0.199	<0.0994	<0.199
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0943	<0.0943	<0.0189	<0.0943	<0.189	<0.189	0.0962	<0.189
10-07-2	<0.0983	<0.0983	<0.0197	<0.0983	<0.197	<0.197	<0.0983	<0.197
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0980	<0.0980	<0.0196	0.1150	<0.196	<0.196	0.1630	<0.196
10-07-3	<0.0976	<0.0976	<0.0195	0.1950	<0.195	<0.195	0.2480	<0.195
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0988	<0.0988	<0.0198	0.2080	<0.198	<0.198	0.2860	<0.198
10-07-4	<0.0942	<0.0942	<0.0188	0.1650	<0.188	<0.188	0.2600	<0.188
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0815	<0.0815	<0.0163	0.1130	<0.163	<0.163	0.1260	<0.163
10-07-5	<0.0801	<0.0801	<0.0160	<0.0801	<0.160	<0.160	0.0883	<0.160
1-08-5								
7-06-6								
10-07-6	<0.0777	<0.0777	<0.0155	<0.0777	<0.155	<0.155	<0.0777	<0.155
1-08-6								
4-08-6	<0.0753	<0.0753	<0.0151	<0.0753	<0.151	<0.151	<0.0753	<0.151

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 196**	**PCB 197**	**PCB 198/199**	**PCB 2**	**PCB 20/28**	**PCB 200**	**PCB 201**	**PCB 202**
7-06-1								
10-06-1	<0.183	<0.183	0.1460	0.0031	0.1240	<0.183	<0.183	<0.183
10-07-1	<0.199	<0.199	<0.0994	0.0034	0.1650	<0.199	<0.199	<0.199
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.189	<0.189	0.1730	0.0029	<0.0943	<0.189	<0.189	<0.189
10-07-2	<0.197	<0.197	<0.0983	0.0047	0.1450	<0.197	<0.197	<0.197
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.196	<0.196	0.2520	0.0036	0.1620	<0.196	<0.196	<0.196
10-07-3	<0.195	<0.195	0.4030	0.0039	0.2630	<0.195	<0.195	<0.195
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.198	<0.198	0.4800	0.0040	0.2080	<0.198	<0.198	<0.198
10-07-4	<0.188	<0.188	0.3580	0.0040	0.1060	<0.188	<0.188	<0.188
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.163	<0.163	0.2740	0.0027	0.0903	<0.163	<0.163	<0.163
10-07-5	<0.160	<0.160	0.1530	0.0031	<0.0801	<0.160	<0.160	<0.160
1-08-5								
7-06-6								
10-07-6	<0.155	<0.155	<0.0777	0.0029	<0.0777	<0.155	<0.155	<0.155
1-08-6								
4-08-6	<0.151	<0.151	0.1250	0.0024	0.0835	<0.151	<0.151	<0.151

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 203**	**PCB 204**	**PCB 205**	**PCB 206**	**PCB 207**	**PCB 208**	**PCB 209**	**PCB 21/33**
7-06-1								
10-06-1	<0.183	<0.183	<0.183	<0.183	<0.183	<0.183	<0.0917	0.0661
10-07-1	<0.199	<0.199	<0.199	<0.199	<0.199	<0.199	<0.0994	0.0727
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.189	<0.189	<0.189	<0.189	<0.189	<0.189	<0.0943	0.0641
10-07-2	<0.197	<0.197	<0.197	<0.197	<0.197	<0.197	<0.0983	0.0591
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.196	<0.196	<0.196	<0.196	<0.196	<0.196	<0.0980	0.0771
10-07-3	0.265	<0.195	<0.195	<0.195	<0.195	<0.195	<0.0976	0.0770
1-08-3								
4-08-3								
7-06-4								
10-06-4	0.323	<0.198	<0.198	<0.198	<0.198	<0.198	<0.0988	0.0873
10-07-4	0.242	<0.188	<0.188	<0.188	<0.188	<0.188	<0.0942	0.0535
1-08-4								
4-08-4								
7-06-5								
10-06-5	0.181	<0.163	<0.163	<0.163	<0.163	<0.163	<0.0815	0.0494
10-07-5	<0.160	<0.160	<0.160	<0.160	<0.160	<0.160	<0.0801	0.0473
1-08-5								
7-06-6								
10-07-6	<0.155	<0.155	<0.155	<0.155	<0.155	<0.155	<0.0777	0.0392
1-08-6								
4-08-6	<0.151	<0.151	<0.151	<0.151	<0.151	<0.151	<0.0753	0.0342

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 22**	**PCB 23**	**PCB 24**	**PCB 25**	**PCB 26/29**	**PCB 27**	**PCB 3**	**PCB 31**
7-06-1								
10-06-1	0.0673	<0.0367	<0.0367	<0.0367	<0.0367	<0.0367	<0.0367	0.121
10-07-1	0.0790	<0.0398	<0.0398	<0.0398	<0.0398	<0.0398	<0.0398	0.136
1-08-1								
4-08-1								
7-06-2								
10-06-2	0.0516	<0.0377	<0.0377	<0.0377	<0.0377	<0.0377	<0.0377	0.099
10-07-2	0.0454	<0.0393	<0.0393	<0.0393	<0.0393	<0.0393	<0.0393	0.103
1-08-2								
4-08-2								
7-06-3								
10-06-3	0.0814	<0.0392	<0.0392	<0.0392	<0.0392	<0.0392	<0.0392	0.155
10-07-3	0.0952	<0.0390	<0.0390	<0.0390	<0.0390	<0.0390	<0.0390	0.176
1-08-3								
4-08-3								
7-06-4								
10-06-4	0.1120	<0.0395	<0.0395	<0.0395	<0.0395	<0.0395	<0.0395	0.191
10-07-4	0.0487	<0.0377	<0.0377	<0.0377	<0.0377	<0.0377	<0.0377	0.095
1-08-4								
4-08-4								
7-06-5								
10-06-5	0.0493	<0.0326	<0.0326	<0.0326	<0.0326	<0.0326	<0.0326	0.093
10-07-5	0.0380	<0.0320	<0.0320	<0.0320	<0.0320	<0.0320	<0.0320	<0.080
1-08-5								
7-06-6								
10-07-6	<0.0311	<0.0311	<0.0311	<0.0311	<0.0311	<0.0311	<0.0311	<0.078
1-08-6								
4-08-6	0.0316	<0.0301	<0.0301	<0.0301	<0.0301	<0.0301	<0.0301	<0.075

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 32**	**PCB 34**	**PCB 35**	**PCB 36**	**PCB 37**	**PCB 38**	**PCB 39**	**PCB 4**
7-06-1								
10-06-1	<0.0367	<0.0367	<0.0367	<0.0367	<0.0917	<0.0367	<0.0367	<0.0917
10-07-1	<0.0398	<0.0398	<0.0398	<0.0398	<0.0994	<0.0398	<0.0398	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0377	<0.0377	<0.0377	<0.0377	<0.0943	<0.0377	<0.0377	<0.0943
10-07-2	<0.0393	<0.0393	<0.0393	<0.0393	<0.0983	<0.0393	<0.0393	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0392	<0.0392	<0.0392	<0.0392	<0.0980	<0.0392	<0.0392	<0.0980
10-07-3	<0.0390	<0.0390	<0.0390	<0.0390	<0.0976	<0.0390	<0.0390	<0.0976
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0395	<0.0395	<0.0395	<0.0395	<0.0988	<0.0395	<0.0395	<0.0988
10-07-4	<0.0377	<0.0377	<0.0377	<0.0377	<0.0942	<0.0377	<0.0377	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0326	<0.0326	<0.0326	<0.0326	<0.0815	<0.0326	<0.0326	<0.0815
10-07-5	<0.0320	<0.0320	<0.0320	<0.0320	<0.0801	<0.0320	<0.0320	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0311	<0.0311	<0.0311	<0.0311	<0.0777	<0.0311	<0.0311	<0.0777
1-08-6								
4-08-6	<0.0301	<0.0301	<0.0301	<0.0301	<0.0753	<0.0301	<0.0301	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 40/41/70**	**PCB 42**	**PCB 43/73**	**PCB 44/47/65**	**PCB 45/51**	**PCB 46**	**PCB 48**	**PCB 49/69**
7-06-1								
10-06-1	<0.0917	<0.0367	<0.0917	0.1380	<0.0367	<0.0367	<0.0367	0.0922
10-07-1	<0.0994	<0.0398	<0.0994	0.1230	<0.0398	<0.0398	<0.0398	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0943	<0.0377	<0.0943	<0.0943	<0.0377	<0.0377	<0.0377	<0.0943
10-07-2	<0.0983	<0.0393	<0.0983	0.1120	<0.0393	<0.0393	<0.0393	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0980	<0.0392	<0.0980	0.1560	<0.0392	<0.0392	<0.0392	<0.0980
10-07-3	<0.0976	0.0657	<0.0976	0.3480	<0.0390	<0.0390	<0.0390	0.2200
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0988	0.0553	<0.0988	0.2180	<0.0395	<0.0395	<0.0395	0.1460
10-07-4	<0.0942	<0.0377	<0.0942	0.1610	<0.0377	<0.0377	<0.0377	0.1010
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0815	<0.0326	<0.0815	0.1130	<0.0326	<0.0326	<0.0326	<0.0815
10-07-5	<0.0801	<0.0320	<0.0801	0.0920	<0.0320	<0.0320	<0.0320	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0777	<0.0311	<0.0777	<0.0777	<0.0311	<0.0311	<0.0311	<0.0777
1-08-6								
4-08-6	<0.0753	<0.0301	<0.0753	0.0761	<0.0301	<0.0301	<0.0301	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 5**	**PCB 50/53**	**PCB 52**	**PCB 54**	**PCB 55**	**PCB 56**	**PCB 57**	**PCB 58**
7-06-1								
10-06-1	<0.0917	<0.0367	0.1830	<0.0917	<0.0917	0.0411	<0.0917	<0.0917
10-07-1	<0.0994	<0.0398	0.1520	<0.0994	<0.0994	0.0454	<0.0994	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0943	<0.0377	0.1230	<0.0943	<0.0943	<0.0377	<0.0943	<0.0943
10-07-2	<0.0983	<0.0393	0.1780	<0.0983	<0.0983	<0.0393	<0.0983	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0980	<0.0392	0.2280	<0.0980	<0.0980	0.0580	<0.0980	<0.0980
10-07-3	<0.0976	<0.0390	0.6910	<0.0976	<0.0976	0.1240	<0.0976	<0.0976
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0988	<0.0395	0.2960	<0.0988	<0.0988	0.0955	<0.0988	<0.0988
10-07-4	<0.0942	<0.0377	0.2920	<0.0942	<0.0942	0.0593	<0.0942	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0815	<0.0326	0.1550	<0.0815	<0.0815	0.0413	<0.0815	<0.0815
10-07-5	<0.0801	<0.0320	0.1500	<0.0801	<0.0801	0.0321	<0.0801	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0777	<0.0311	0.0908	<0.0777	<0.0777	<0.0311	<0.0777	<0.0777
1-08-6								
4-08-6	<0.0753	<0.0301	0.1160	<0.0753	<0.0753	<0.0301	<0.0753	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 59/62/75**	**PCB 6**	**PCB 60**	**PCB 63**	**PCB 64**	**PCB 66**	**PCB 67**	**PCB 68**
7-06-1								
10-06-1	<0.0367	0.0162	<0.0917	<0.0917	0.0552	0.1070	<0.0917	<0.0917
10-07-1	<0.0398	<0.0099	<0.0994	<0.0994	0.0617	0.1290	<0.0994	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0377	0.0116	<0.0943	<0.0943	0.0391	<0.0943	<0.0943	<0.0943
10-07-2	<0.0393	<0.0098	<0.0983	<0.0983	0.0462	<0.0983	<0.0983	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0392	<0.0098	<0.0980	<0.0980	0.0766	0.1180	<0.0980	<0.0980
10-07-3	<0.0390	0.0142	<0.0976	<0.0976	0.1620	0.2960	<0.0976	<0.0976
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0395	<0.0099	<0.0988	<0.0988	0.1200	0.1860	<0.0988	<0.0988
10-07-4	<0.0377	0.0119	<0.0942	<0.0942	0.0760	0.1380	<0.0942	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0326	<0.0082	<0.0815	<0.0815	0.0532	0.0877	<0.0815	<0.0815
10-07-5	<0.0320	0.0120	<0.0801	<0.0801	0.0409	<0.0801	<0.0801	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0311	0.0118	<0.0777	<0.0777	<0.0311	<0.0777	<0.0777	<0.0777
1-08-6								
4-08-6	<0.0301	<0.0075	<0.0753	<0.0753	0.0337	<0.0753	<0.0753	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 7**	**PCB 70/61/74/76**	**PCB 72**	**PCB 77**	**PCB 78**	**PCB 79**	**PCB 8**	**PCB 80**
7-06-1								
10-06-1	<0.0917	0.2130	<0.0917	<0.0917	<0.0917	<0.0917	<0.0917	<0.0917
10-07-1	<0.0994	0.2280	<0.0994	<0.0994	<0.0994	<0.0994	<0.0994	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0943	0.1350	<0.0943	<0.0943	<0.0943	<0.0943	<0.0943	<0.0943
10-07-2	<0.0983	0.1910	<0.0983	<0.0983	<0.0983	<0.0983	<0.0983	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0980	0.2460	<0.0980	<0.0980	<0.0980	<0.0980	<0.0980	<0.0980
10-07-3	<0.0976	0.7760	<0.0976	<0.0976	<0.0976	<0.0976	<0.0976	<0.0976
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0988	0.3830	<0.0988	<0.0988	<0.0988	<0.0988	<0.0988	<0.0988
10-07-4	<0.0942	0.3470	<0.0942	<0.0942	<0.0942	<0.0942	<0.0942	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0815	0.1810	<0.0815	<0.0815	<0.0815	<0.0815	<0.0815	<0.0815
10-07-5	<0.0801	0.1620	<0.0801	<0.0801	<0.0801	<0.0801	<0.0801	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0777	0.0834	<0.0777	<0.0777	<0.0777	<0.0777	<0.0777	<0.0777
1-08-6								
4-08-6	<0.0753	0.1180	<0.0753	<0.0753	<0.0753	<0.0753	<0.0753	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 81**	**PCB 82**	**PCB 83/99**	**PCB 84**	**PCB 85/116**	**PCB 86/87/ 97/109/119/125**	**PCB 88/91**	**PCB 89**
7-06-1								
10-06-1	<0.0917	<0.0917	0.2810	<0.0917	0.0509	0.2070	<0.0917	<0.0917
10-07-1	<0.0994	<0.0994	<0.0994	<0.0994	<0.0398	0.1090	<0.0994	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0943	<0.0943	<0.0943	<0.0943	<0.0377	0.1300	<0.0943	<0.0943
10-07-2	<0.0983	<0.0983	0.1710	<0.0983	0.0462	0.2230	<0.0983	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0980	<0.098	0.1710	<0.098	0.0471	0.2230	<0.0980	<0.0980
10-07-3	<0.0976	0.1360	0.8260	0.2750	0.2360	1.0900	0.1610	<0.0976
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0988	<0.0988	0.2800	<0.0988	0.0801	0.3390	<0.0988	<0.0988
10-07-4	<0.0942	<0.0942	0.4600	0.1400	0.1290	0.5900	<0.0942	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0815	<0.0815	0.1800	<0.0815	0.0492	0.2110	<0.0815	<0.0815
10-07-5	<0.0801	<0.0801	0.1740	<0.0801	0.0517	0.2320	<0.0801	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0777	<0.0777	0.0944	<0.0777	<0.0311	0.1080	<0.0777	<0.0777
1-08-6								
4-08-6	<0.0753	<0.0753	0.1290	<0.0753	0.0311	0.1550	<0.0753	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**PCB 9**	**PCB 90/101/113**	**PCB 92**	**PCB 94**	**PCB 95**	**PCB 96**	**PCB 98/100**	**PCB 98/102**
7-06-1								
10-06-1	<0.0917	0.4280	<0.0917	<0.0917	0.180	<0.0917	<0.0917	<0.0917
10-07-1	<0.0994	<0.1990	<0.0994	<0.0994	0.103	<0.0994	<0.0994	<0.0994
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0943	0.3730	<0.0943	<0.0943	0.185	<0.0943	<0.0943	<0.0943
10-07-2	<0.0983	0.3930	<0.0983	<0.0983	0.200	<0.0983	<0.0983	<0.0983
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0980	0.5510	<0.0980	<0.0980	0.270	<0.0980	<0.0980	<0.0980
10-07-3	<0.0976	1.9800	<0.0976	<0.0976	0.900	<0.0976	<0.0976	<0.0976
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0988	0.8490	<0.0988	<0.0988	0.375	<0.0988	<0.0988	<0.0988
10-07-4	<0.0942	1.1400	<0.0942	<0.0942	0.461	<0.0942	<0.0942	<0.0942
1-08-4								
4-08-4								
7-06-5								
10-06-5	<0.0815	0.4630	<0.0815	<0.0815	0.187	<0.0815	<0.0815	<0.0815
10-07-5	<0.0801	0.4520	<0.0801	<0.0801	0.211	<0.0801	<0.0801	<0.0801
1-08-5								
7-06-6								
10-07-6	<0.0777	0.2160	<0.0777	<0.0777	0.108	<0.0777	<0.0777	<0.0777
1-08-6								
4-08-6	<0.0753	0.3460	<0.0753	<0.0753	0.172	<0.0753	<0.0753	<0.0753

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**pentachloro-anisole**	**chlorpyrifos**	**toxaphene**	**1,2,3,4-Tetrachloro-benzene**	**1,2,4,5-Tetrachloro-benzene**
7-06-1					
10-06-1	0.481	<0.277	<10.5	<0.151	<0.269
10-07-1	0.168	<0.277	<10.5	<0.151	<0.269
1-08-1					
4-08-1					
7-06-2					
10-06-2	0.226	<0.277	<10.5	<0.151	<0.269
10-07-2	<0.152	<0.277	<10.5	<0.151	<0.269
1-08-2					
4-08-2					
7-06-3					
10-06-3	0.982	<0.277	<10.5	<0.151	<0.269
10-07-3	0.865	<0.277	<10.5	<0.151	<0.269
1-08-3					
4-08-3					
7-06-4					
10-06-4	1.050	<0.277	<10.5	<0.151	<0.269
10-07-4	0.386	<0.277	<10.5	<0.151	<0.269
1-08-4					
4-08-4					
7-06-5					
10-06-5	1.370	<0.277	<10.5	<0.151	<0.269
10-07-5	0.314	<0.277	<10.5	<0.151	<0.269
1-08-5					
7-06-6					
10-07-6	0.296	<0.277	<10.5	<0.151	<0.269
1-08-6					
4-08-6	0.261	<0.277	<10.5	<0.151	<0.269

Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]

Sample ID	Pentachloro-benzene	BDE 1	BDE 10	BDE 100	BDE 11	BDE 116	BDE 118	BDE 119
7-06-1		<0.176	<0.176	6.1	<0.176	<0.186	<0.186	<0.186
10-06-1	2.60							
10-07-1	2.49							
1-08-1								
4-08-1		<0.179	<0.179	1.0	<0.179	<0.190	<0.190	<0.190
7-06-2		<0.177	<0.177	6.8	<0.177	<0.188	<0.188	0.200
10-06-2	2.59							
10-07-2	2.18							
1-08-2								
4-08-2		<0.176	<0.176	3.1	<0.176	<0.186	<0.186	<0.186
7-06-3		<0.179	<0.179	26.9	<0.179	<0.190	<0.190	1.400
10-06-3	2.73							
10-07-3	3.15							
1-08-3								
4-08-3		<0.176	<0.176	8.9	<0.176	<0.186	<0.186	0.300
7-06-4		<0.172	<0.172	44.7	<0.172	<0.182	<0.182	2.000
10-06-4	3.13							
10-07-4	2.86							
1-08-4								
4-08-4		<0.182	<0.182	16.1	<0.182	<0.192	<0.192	0.800
7-06-5		<0.173	<0.173	15.5	<0.173	<0.183	<0.183	0.800
10-06-5	3.79							
10-07-5	2.49							
1-08-5								
7-06-6		<0.167	<0.167	9.1	<0.167	<0.177	<0.177	0.500
10-07-6	3.01							
1-08-6								
4-08-6	3.00	<0.174	<0.174	3.5	<0.174	<0.184	<0.184	<0.184

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**BDE 12**	**BDE 126**	**BDE 13**	**BDE 138**	**BDE 15**	**BDE 153**	**BDE 154**	**BDE 155**
7-06-1	<0.176	<0.186	<0.176	<0.139	<0.176	0.50	1.6	<0.139
10-06-1								
10-07-1								
1-08-1								
4-08-1	<0.179	<0.190	<0.179	0.200	<0.179	0.20	0.4	<0.141
7-06-2	<0.177	<0.188	<0.177	<0.140	<0.177	0.60	1.9	<0.140
10-06-2								
10-07-2								
1-08-2								
4-08-2	<0.176	<0.186	<0.176	<0.138	<0.176	0.40	1.0	<0.138
7-06-3	<0.179	0.400	<0.179	<0.141	0.700	3.30	7.0	<0.141
10-06-3								
10-07-3								
1-08-3								
4-08-3	<0.176	0.200	<0.176	<0.139	<0.176	1.30	2.7	<0.139
7-06-4	<0.172	0.800	<0.172	<0.135	0.800	4.20	12.8	<0.135
10-06-4								
10-07-4								
1-08-4								
4-08-4	<0.182	0.300	<0.182	<0.143	0.200	2.20	4.9	<0.143
7-06-5	<0.173	<0.183	<0.173	<0.136	0.300	1.00	4.2	<0.136
10-06-5								
10-07-5								
1-08-5								
7-06-6	<0.167	<0.177	<0.167	<0.132	0.200	0.70	2.6	<0.132
10-07-6								
1-08-6								
4-08-6	<0.174	<0.184	<0.174	<0.137	<0.174	0.50	1.2	<0.137

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**BDE 166**	**BDE 17**	**BDE 181**	**BDE 183**	**BDE 190**	**BDE 194**	**BDE 195**	**BDE 196**
7-06-1	<0.139	<0.176	<0.129	<0.129	<0.097	<0.097	<0.097	<0.097
10-06-1								
10-07-1								
1-08-1								
4-08-1	<0.141	<0.179	<0.131	0.200	0.500	<0.099	<0.099	<0.099
7-06-2	<0.140	<0.177	<0.130	<0.130	<0.098	<0.098	<0.098	<0.098
10-06-2								
10-07-2								
1-08-2								
4-08-2	<0.138	<0.176	<0.128	0.200	0.400	<0.097	<0.097	<0.097
7-06-3	<0.141	0.600	<0.131	<0.131	<0.099	<0.099	<0.099	<0.099
10-06-3								
10-07-3								
1-08-3								
4-08-3	<0.139	<0.176	<0.129	<0.129	<0.097	<0.097	<0.097	<0.097
7-06-4	<0.135	1.000	<0.126	<0.126	<0.095	<0.095	<0.095	<0.095
10-06-4								
10-07-4								
1-08-4								
4-08-4	<0.143	<0.182	<0.133	0.300	0.200	<0.101	<0.101	<0.101
7-06-5	<0.136	0.300	<0.127	<0.127	<0.096	<0.096	<0.096	<0.096
10-06-5								
10-07-5								
1-08-5								
7-06-6	<0.132	0.200	<0.122	<0.122	<0.092	<0.092	<0.092	<0.092
10-07-6								
1-08-6								
4-08-6	<0.137	<0.174	<0.127	<0.127	<0.096	<0.096	<0.096	<0.096

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**BDE 197**	**BDE 198/ 199/203/200**	**BDE 2**	**BDE 201**	**BDE 202**	**BDE 204**	**BDE 205**	**BDE 206**
7-06-1	<0.097	<0.097	<0.176	<0.097	<0.097	<0.097	0.4	0.600
10-06-1								
10-07-1								
1-08-1								
4-08-1	<0.099	<0.099	<0.179	<0.099	<0.099	<0.099	0.5	0.700
7-06-2	<0.098	<0.098	<0.177	<0.098	<0.098	<0.098	0.5	<0.098
10-06-2								
10-07-2								
1-08-2								
4-08-2	<0.097	<0.097	<0.176	<0.097	<0.097	<0.097	0.5	0.600
7-06-3	<0.099	<0.099	<0.179	<0.099	<0.099	<0.099	0.6	<0.099
10-06-3								
10-07-3								
1-08-3								
4-08-3	<0.097	<0.097	<0.176	<0.097	<0.097	<0.097	0.4	<0.097
7-06-4	<0.095	<0.095	<0.172	<0.095	<0.095	<0.095	0.4	0.400
10-06-4								
10-07-4								
1-08-4								
4-08-4	<0.101	<0.101	<0.182	<0.101	<0.101	<0.101	0.6	0.600
7-06-5	<0.096	<0.096	<0.173	<0.096	<0.096	<0.096	0.5	0.500
10-06-5								
10-07-5								
1-08-5								
7-06-6	<0.092	<0.092	<0.167	<0.092	<0.092	<0.092	0.5	<0.092
10-07-6								
1-08-6								
4-08-6	<0.096	<0.096	<0.174	<0.096	<0.096	<0.096	0.7	<0.096

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]								
Sample ID	**BDE 207**	**BDE 208**	**BDE 209**	**BDE 25**	**BDE 28**	**BDE 3**	**BDE 30**	**BDE 32**	**BDE 33**
7-06-1	0.7	0.400	<1.45	<0.176	2.7	<0.176	<0.176	<0.176	<0.176
10-06-1									
10-07-1									
1-08-1									
4-08-1	0.8	<0.099	<1.47	<0.179	0.3	<0.179	<0.179	<0.179	<0.179
7-06-2	0.7	0.300	<1.46	<0.177	2.2	<0.177	<0.177	<0.177	<0.177
10-06-2									
10-07-2									
1-08-2									
4-08-2	0.8	0.400	<1.44	<0.176	0.9	<0.176	<0.176	<0.176	<0.176
7-06-3	0.6	0.500	<1.47	<0.179	5.1	3.300	<0.179	<0.179	<0.179
10-06-3									
10-07-3									
1-08-3									
4-08-3	0.7	0.500	<1.45	<0.176	1.7	0.600	<0.176	<0.176	<0.176
7-06-4	0.5	0.300	<1.41	<0.172	8.5	1.600	<0.172	<0.172	<0.172
10-06-4									
10-07-4									
1-08-4									
4-08-4	0.7	0.400	<1.49	<0.182	2.5	0.700	<0.182	<0.182	<0.182
7-06-5	0.8	0.400	<1.42	<0.173	3.6	<0.173	<0.173	<0.173	<0.173
10-06-5									
10-07-5									
1-08-5									
7-06-6	0.6	<0.092	<1.37	<0.167	2.1	<0.167	<0.167	<0.167	<0.167
10-07-6									
1-08-6									
4-08-6	0.9	0.600	<1.43	<0.174	0.6	<0.174	<0.174	<0.174	<0.174

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]								
Sample ID	**BDE 35**	**BDE 37**	**BDE 47**	**BDE 49/71**	**BDE 66**	**BDE 7**	**BDE 75**	**BDE 77**	**BDE 8**
7-06-1	<0.176	<0.176	48.70	2.00	<0.112	<0.176	<0.205	0.400	<0.176
10-06-1									
10-07-1									
1-08-1									
4-08-1	<0.179	<0.179	6.00	<0.153	<0.114	<0.179	<0.209	<0.209	<0.179
7-06-2	<0.177	<0.177	50.60	2.10	<0.113	<0.177	<0.207	0.700	<0.177
10-06-2									
10-07-2									
1-08-2									
4-08-2	<0.176	<0.176	22.00	0.90	<0.112	<0.176	<0.205	0.300	<0.176
7-06-3	<0.179	0.200	266.00	5.70	<0.114	<0.179	<0.209	<0.209	<0.179
10-06-3									
10-07-3									
1-08-3									
4-08-3	<0.176	<0.176	75.10	2.00	<0.112	<0.176	<0.205	<0.205	<0.176
7-06-4	<0.172	0.500	385.00	12.50	<0.110	<0.172	<0.200	<0.200	<0.172
10-06-4									
10-07-4									
1-08-4									
4-08-4	<0.182	<0.182	129.00	3.20	<0.116	<0.182	<0.212	0.500	<0.182
7-06-5	<0.173	0.300	127.00	6.50	<0.111	<0.173	0.400	<0.201	<0.173
10-06-5									
10-07-5									
1-08-5									
7-06-6	<0.167	0.500	70.40	4.00	<0.107	<0.167	<0.194	1.000	<0.167
10-07-6									
1-08-6									
4-08-6	<0.174	<0.174	26.60	1.00	<0.111	<0.174	<0.203	<0.203	<0.174

Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]

Sample ID	BDE 85	BDE 99	BDE-TOTAL	n-decane	n-undecane	n-dodecane	n-tridecane
7-06-1	<0.384	<0.186	64.3				
10-06-1							
10-07-1							
1-08-1				199.0	18500.0	22.8	168.000
4-08-1	<0.391	<0.190	11.0				
7-06-2	<0.387	<0.188	66.7				
10-06-2							
10-07-2							
1-08-2				47.9	41.0	32.7	333.000
4-08-2	<0.383	<0.186	31.5				
7-06-3	0.500	<0.190	323.0				
10-06-3							
10-07-3							
1-08-3				66.0	72.5	56.9	912.000
4-08-3	<0.384	<0.186	94.6				
7-06-4	1.100	<0.182	477.0				
10-06-4							
10-07-4							
1-08-4				69.5	93.3	52.6	577.000
4-08-4	0.400	<0.192	164.0				
7-06-5	<0.377	<0.183	162.0				
10-06-5							
10-07-5							
1-08-5							
7-06-6	<0.364	<0.177	92.4				
10-07-6							
1-08-6				90.1	90.8	<9.2	1090.000
4-08-6	<0.380	<0.184	35.7				

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**n-tetradecane**	**n-pentadecane**	**n-hexadecane**	**n-heptadecane**	**n-octadecane**
7-06-1					
10-06-1					
10-07-1					
1-08-1	102.0	3150.0	271.0	2730	2750.000
4-08-1					
7-06-2					
10-06-2					
10-07-2					
1-08-2	132.0	5610.0	337.0	6490	3360.000
4-08-2					
7-06-3					
10-06-3					
10-07-3					
1-08-3	343.0	11200.0	780.0	7660	6920.000
4-08-3					
7-06-4					
10-06-4					
10-07-4					
1-08-4	86.7	6690.0	849.0	4120	4790.000
4-08-4					
7-06-5					
10-06-5					
10-07-5					
1-08-5					
7-06-6					
10-07-6					
1-08-6	137.0	2870.0	517.0	17600	4660.000
4-08-6					

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**n-nonadecane**	**n-eicosane**	**n-heneicosane**	**n-docosane**	**n-tricosane**
7-06-1					
10-06-1					
10-07-1					
1-08-1	219.0	1210.0	70.2	<16.4	10.9
4-08-1					
7-06-2					
10-06-2					
10-07-2					
1-08-2	419.0	<16.5	<17.4	<16.2	6.4
4-08-2					
7-06-3					
10-06-3					
10-07-3					
1-08-3	5320.0	185.0	1450.0	889.0	3.1
4-08-3					
7-06-4					
10-06-4					
10-07-4					
1-08-4	173.0	327.0	103.0	<16.4	< 0.02
4-08-4					
7-06-5					
10-06-5					
10-07-5					
1-08-5					
7-06-6					
10-07-6					
1-08-6	825.0	<16.2	<17.1	<15.9	3.8
4-08-6					

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**n-tetracosane**	**n-pentacosane**	**n-hexacosane**	**n-heptacosane**	**n-octacosane**
7-06-1					
10-06-1					
10-07-1					
1-08-1	562	1230	216	294.0	52.8
4-08-1					
7-06-2					
10-06-2					
10-07-2					
1-08-2	211	327	91	102.0	<14.3
4-08-2					
7-06-3					
10-06-3					
10-07-3					
1-08-3	67	846	312	463.0	188.0
4-08-3					
7-06-4					
10-06-4					
10-07-4					
1-08-4	90	434	545	1210.0	<14.4
4-08-4					
7-06-5					
10-06-5					
10-07-5					
1-08-5					
7-06-6					
10-07-6					
1-08-6	134	353	567	132.0	<14.0
4-08-6					

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**n-nonacosane**	**n-triacontane**	**n-hentriacontane**	**n-dotriacontane**	**n-tritriacontane**
7-06-1					
10-06-1					
10-07-1					
1-08-1	58.700	59.6	111.00	<14.9	<12.7
4-08-1					
7-06-2					
10-06-2					
10-07-2					
1-08-2	<13.2	<14.0	<15.1	<14.8	<12.6
4-08-2					
7-06-3					
10-06-3					
10-07-3					
1-08-3	6880.000	270.0	5850.00	<14.8	<12.6
4-08-3					
7-06-4					
10-06-4					
10-07-4					
1-08-4	8260.000	256.0	<15.2	<14.9	<12.7
4-08-4					
7-06-5					
10-06-5					
10-07-5					
1-08-5					
7-06-6					
10-07-6					
1-08-6	<13.0	<13.8	<14.8	<14.5	<12.3
4-08-6					

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]					
Sample ID	**n-tetratriacontane**	**phytane**	**pristane**	**1,6,7-trimethyl-naphthalene**	**1-methyl-naphthalene**	**1-methyl-phenanthrene**
7-06-1						
10-06-1						
10-07-1						
1-08-1	<14.6	204	235	<0.451	2.6	<0.51
4-08-1						
7-06-2						
10-06-2						
10-07-2						
1-08-2	<14.5	175	132	<0.446	3.7	7.80
4-08-2						
7-06-3						
10-06-3						
10-07-3						
1-08-3	<14.5	362	251	<0.447	3.4	1.80
4-08-3						
7-06-4						
10-06-4						
10-07-4						
1-08-4	<14.6	428	308	<0.450	3.6	<0.51
4-08-4						
7-06-5						
10-06-5						
10-07-5						
1-08-5						
7-06-6						
10-07-6						
1-08-6	<14.2	278	210	<0.438	2.1	4.70
4-08-6						

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**2,6-dimethyl-naphthalene**	**2-methyl-naphthalene**	**acenaphthalene**	**acenaphthene**	**anthracene**
7-06-1					
10-06-1					
10-07-1					
1-08-1	0.70	5.1	0.5	<0.41	0.3
4-08-1					
7-06-2					
10-06-2					
10-07-2					
1-08-2	1.00	6.8	0.4	<0.41	0.3
4-08-2					
7-06-3					
10-06-3					
10-07-3					
1-08-3	1.30	5.9	1.0	0.70	0.4
4-08-3					
7-06-4					
10-06-4					
10-07-4					
1-08-4	1.00	5.7	1.3	0.70	0.5
4-08-4					
7-06-5					
10-06-5					
10-07-5					
1-08-5					
7-06-6					
10-07-6					
1-08-6	<0.64	3.8	0.7	0.50	0.3
4-08-6					

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]					
Sample ID	**benzo(a) anthracene**	**benzo(a) pyrene**	**benzo(b) fluoranthene**	**benzo(e) pyrene**	**benzo(g,h,i) perylene**	**benzo(k) fluoranthene**
7-06-1						
10-06-1						
10-07-1						
1-08-1	<0.378	<0.451	0.80	<0.475	<0.536	<0.414
4-08-1						
7-06-2						
10-06-2						
10-07-2						
1-08-2	<0.374	<0.446	0.40	<0.470	<0.530	<0.410
4-08-2						
7-06-3						
10-06-3						
10-07-3						
1-08-3	<0.374	<0.447	0.70	<0.471	<0.532	<0.411
4-08-3						
7-06-4						
10-06-4						
10-07-4						
1-08-4	<0.377	<0.450	1.30	0.600	<0.535	<0.413
4-08-4						
7-06-5						
10-06-5						
10-07-5						
1-08-5						
7-06-6						
10-07-6						
1-08-6	<0.367	<0.438	<0.32	<0.462	<0.521	<0.403
4-08-6						

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**biphenyl**	**C1-chrysenes**	**C1-dibenzo-thiophenes**	**C1-Fluoranthenes & Pyrenes**	**C1-fluorenes**
7-06-1					
10-06-1					
10-07-1					
1-08-1	1.1	<0.938	<0.682	<1.28	1.700
4-08-1					
7-06-2					
10-06-2					
10-07-2					
1-08-2	1.8	<0.928	<0.675	<1.27	2.600
4-08-2					
7-06-3					
10-06-3					
10-07-3					
1-08-3	5.6	<0.930	3.100	<1.27	<0.821
4-08-3					
7-06-4					
10-06-4					
10-07-4					
1-08-4	2.8	<0.936	4.100	<1.28	<0.826
4-08-4					
7-06-5					
10-06-5					
10-07-5					
1-08-5					
7-06-6					
10-07-6					
1-08-6	1.7	<0.912	1.600	<1.24	<0.806
4-08-6					

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]			
Sample ID	**C1-naphthalenes**	**C1-Phenanthrenes & Anthracenes**	**C2-chrysenes**	**C2-dibenzo-thiophenes**
7-06-1				
10-06-1				
10-07-1				
1-08-1	5.0	<0.597	<0.938	<0.682
4-08-1				
7-06-2				
10-06-2				
10-07-2				
1-08-2	6.8	7.900	<0.928	<0.675
4-08-2				
7-06-3				
10-06-3				
10-07-3				
1-08-3	6.0	2.600	<0.930	<0.677
4-08-3				
7-06-4				
10-06-4				
10-07-4				
1-08-4	6.0	<0.596	<0.936	<0.681
4-08-4				
7-06-5				
10-06-5				
10-07-5				
1-08-5				
7-06-6				
10-07-6				
1-08-6	3.8	5.300	<0.912	<0.664
4-08-6				

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]			
Sample ID	**C2-fluorenes**	**C2-naphthalenes**	**C2-Phenanthrenes & Anthracenes**	**C3-chrysenes**
7-06-1				
10-06-1				
10-07-1				
1-08-1	<0.828	2.3	<0.597	<0.938
4-08-1				
7-06-2				
10-06-2				
10-07-2				
1-08-2	<0.820	3.7	<0.591	<0.928
4-08-2				
7-06-3				
10-06-3				
10-07-3				
1-08-3	<0.821	3.8	<0.592	<0.930
4-08-3				
7-06-4				
10-06-4				
10-07-4				
1-08-4	<0.826	3.6	<0.596	<0.936
4-08-4				
7-06-5				
10-06-5				
10-07-5				
1-08-5				
7-06-6				
10-07-6				
1-08-6	<0.806	2.7	<0.581	<0.912
4-08-6				

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]			
Sample ID	**C3-dibenzo-thiophenes**	**C3-fluorenes**	**C3-naphthalenes**	**C3-Phenanthrenes & Anthracenes**
7-06-1				
10-06-1				
10-07-1				
1-08-1	<0.682	<0.828	<2.00	<0.597
4-08-1				
7-06-2				
10-06-2				
10-07-2				
1-08-2	<0.675	<0.820	<1.98	<0.591
4-08-2				
7-06-3				
10-06-3				
10-07-3				
1-08-3	<0.677	<0.821	2.60	<0.592
4-08-3				
7-06-4				
10-06-4				
10-07-4				
1-08-4	<0.681	<0.826	2.90	<0.596
4-08-4				
7-06-5				
10-06-5				
10-07-5				
1-08-5				
7-06-6				
10-07-6				
1-08-6	<0.664	<0.806	<1.94	<0.581
4-08-6				

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]				
Sample ID	**C4-chrysenes**	**C4-naphthalenes**	**C4-Phenanthrenes & Anthracenes**	**chrysene**	**Dibenz(a,h) anthracene**
7-06-1					
10-06-1					
10-07-1					
1-08-1	<0.938	<2.00	<0.597	0.50	<0.426
4-08-1					
7-06-2					
10-06-2					
10-07-2					
1-08-2	<0.928	<1.98	<0.591	0.50	<0.422
4-08-2					
7-06-3					
10-06-3					
10-07-3					
1-08-3	<0.930	<1.98	<0.592	0.50	<0.423
4-08-3					
7-06-4					
10-06-4					
10-07-4					
1-08-4	<0.936	<1.99	<0.596	0.90	<0.425
4-08-4					
7-06-5					
10-06-5					
10-07-5					
1-08-5					
7-06-6					
10-07-6					
1-08-6	<0.912	<1.94	<0.581	<0.46	<0.415
4-08-6					

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]					
Sample ID	**dibenzo-thiophene**	**fluoranthene**	**fluorene**	**indeno(1,2,3-cd) pyrene**	**naphthalene**	**perylene**
7-06-1						
10-06-1						
10-07-1						
1-08-1	<0.341	1.0	1.2	<0.694	13.0	<0.645
4-08-1						
7-06-2						
10-06-2						
10-07-2						
1-08-2	0.400	1.8	1.5	<0.687	13.9	<0.639
4-08-2						
7-06-3						
10-06-3						
10-07-3						
1-08-3	0.400	1.2	2.2	<0.689	14.8	<0.640
4-08-3						
7-06-4						
10-06-4						
10-07-4						
1-08-4	0.500	2.8	1.9	<0.693	15.4	<0.644
4-08-4						
7-06-5						
10-06-5						
10-07-5						
1-08-5						
7-06-6						
10-07-6						
1-08-6	<0.332	0.5	1.4	<0.675	17.3	<0.628
4-08-6						

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]					
Sample ID	**phenanthrene**	**pyrene**	**1,2,3,4,6,7,8-HpCDD**	**1,2,3,4,6,7,8-HpCDF**	**1,2,3,4,7,8,9-HpCDF**	**1,2,3,4,7,8-HxCDD**
7-06-1						
10-06-1			<0.0092	<0.0092	<0.0092	<0.0092
10-07-1			<0.0099	<0.0099	<0.0099	<0.0099
1-08-1	2.9	0.6				
4-08-1						
7-06-2						
10-06-2			<0.0094	<0.0094	<0.0094	<0.0094
10-07-2			<0.0098	<0.0098	<0.0098	<0.0098
1-08-2	4.7	0.8				
4-08-2						
7-06-3						
10-06-3			<0.0098	<0.0098	<0.0098	<0.0098
10-07-3			<0.0098	<0.0098	<0.0098	<0.0098
1-08-3	4.4	0.7				
4-08-3						
7-06-4						
10-06-4			<0.0099	<0.0099	<0.0099	<0.0099
10-07-4			<0.0094	<0.0094	<0.0094	<0.0094
1-08-4	5.0	1.8				
4-08-4						
7-06-5						
10-06-5						
10-07-5						
1-08-5						
7-06-6						
10-07-6						
1-08-6	1.9	<0.4				
4-08-6						

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]					
Sample ID	**1,2,3,4,7,8-HxCDF**	**1,2,3,6,7,8-HxCDD**	**1,2,3,6,7,8-HxCDF**	**1,2,3,7,8,9-HxCDD**	**1,2,3,7,8,9-HxCDF**	**1,2,3,7,8-PeCDD**
7-06-1						
10-06-1	<0.0092	<0.0092	<0.0092	<0.0092	<0.0092	<0.0092
10-07-1	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099
1-08-1						
4-08-1						
7-06-2						
10-06-2	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094
10-07-2	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098
1-08-2						
4-08-2						
7-06-3						
10-06-3	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098
10-07-3	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098
1-08-3						
4-08-3						
7-06-4						
10-06-4	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099
10-07-4	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094
1-08-4						
4-08-4						
7-06-5						
10-06-5						
10-07-5						
1-08-5						
7-06-6						
10-07-6						
1-08-6						
4-08-6						

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]						
Sample ID	**1,2,3,7,8-PeCDF**	**2,3,4,6,7,8-HxCDF**	**2,3,4,7,8-PeCDF**	**2,3,7,8-TCDD**	**2,3,7,8-TCDF**	**Cl4-PCDD**	**Cl4-PCDF**
7-06-1							
10-06-1	<0.0092	<0.0092	<0.0092	<0.002	<0.002	<0.002	<0.002
10-07-1	<0.0099	<0.0099	<0.0099	<0.002	<0.002	<0.002	<0.002
1-08-1							
4-08-1							
7-06-2							
10-06-2	<0.0094	<0.0094	<0.0094	<0.002	<0.002	<0.002	<0.002
10-07-2	<0.0098	<0.0098	<0.0098	<0.002	<0.002	<0.002	<0.002
1-08-2							
4-08-2							
7-06-3							
10-06-3	<0.0098	<0.0098	<0.0098	<0.002	<0.002	<0.002	<0.002
10-07-3	<0.0098	<0.0098	<0.0098	<0.002	<0.002	<0.002	<0.002
1-08-3							
4-08-3							
7-06-4							
10-06-4	<0.0099	<0.0099	<0.0099	<0.002	<0.002	<0.002	<0.002
10-07-4	<0.0094	<0.0094	<0.0094	<0.002	<0.002	<0.002	<0.002
1-08-4							
4-08-4							
7-06-5							
10-06-5							
10-07-5							
1-08-5							
7-06-6							
10-07-6							
1-08-6							
4-08-6							

	Appendix 3E. Concentrations of organic contaminants in Rio Grande silvery minnow carcasses from all sites expressed as micrograms per kilogram (µg/kg) wet weight. ["Sample ID", Sample identification number, includes month and year sampled and site number; "<", less than laboratory method detection limit (as indicated); blank values indicate no results available; see Table 2 for names and abbreviations; see Table 1 for site number, description, and location]							
Sample ID	**Cl5-PCDD**	**Cl5-PCDF**	**Cl6-PCDD**	**Cl6-PCDF**	**Cl7-PCDD**	**Cl7-PCDF**	**OCDD**	**OCDF**
7-06-1								
10-06-1	<0.0092	<0.0092	<0.0092	<0.0092	<0.0092	<0.0092	<0.002	<0.002
10-07-1	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099	<0.002	<0.002
1-08-1								
4-08-1								
7-06-2								
10-06-2	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094	<0.002	<0.002
10-07-2	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.002	<0.002
1-08-2								
4-08-2								
7-06-3								
10-06-3	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.002	<0.002
10-07-3	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.0098	<0.002	<0.002
1-08-3								
4-08-3								
7-06-4								
10-06-4	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099	<0.0099	<0.002	<0.002
10-07-4	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094	<0.0094	<0.002	<0.002
1-08-4								
4-08-4								
7-06-5								
10-06-5								
10-07-5								
1-08-5								
7-06-6								
10-07-6								
1-08-6								
4-08-6								